



# **In vivo bioorthogonal chemistry for imaging and therapy**

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October 1, 2020

# Outline

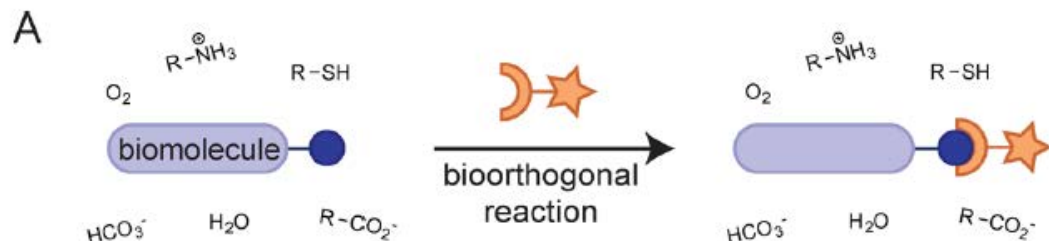
- **Part 1 - Click**

- Bioorthogonal reactions recap
- Quick introduction to nuclear imaging and “internal radiotherapy
- Tumor pretargeting for radioimmuno-imaging and –therapy
- Chemical approach to tumor pretargeting
- Summary

- **Part 2 – Click-to-release**

- “Click-to-release” ADC approach to cancer therapy
- Designing a suitable activator for in vivo applications
- Other applications of the IEDDA pyridazine elimination reaction

# Bioorthogonal reactions recap



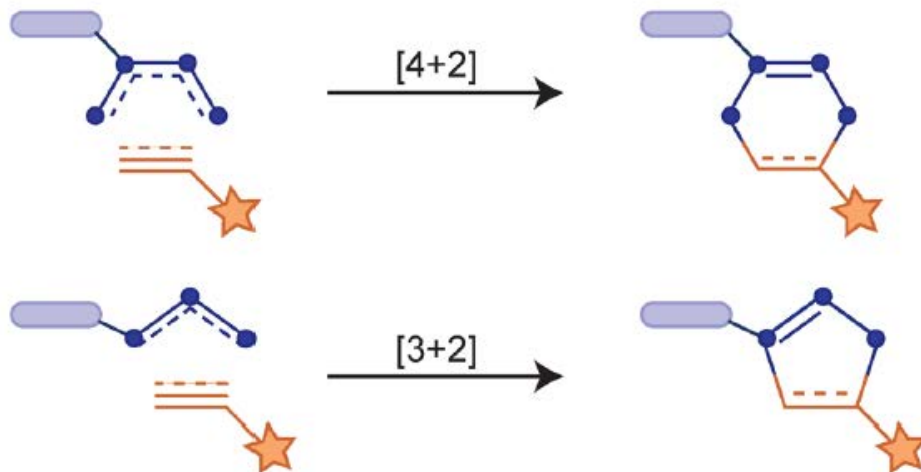
Examples:

**B** polar reactions



Staudinger Ligation

cycloadditions

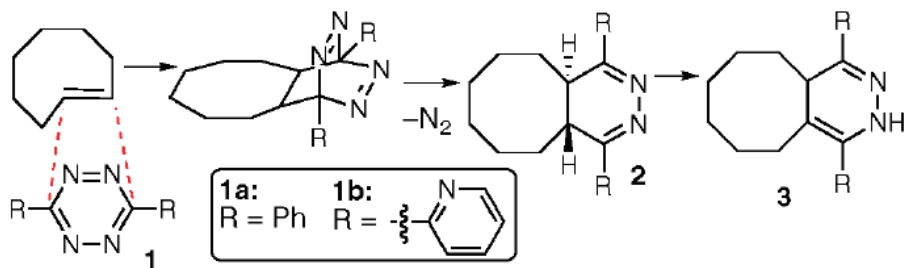


Inverse-electron demand Diels-Alder cycloaddition (IEDDA)

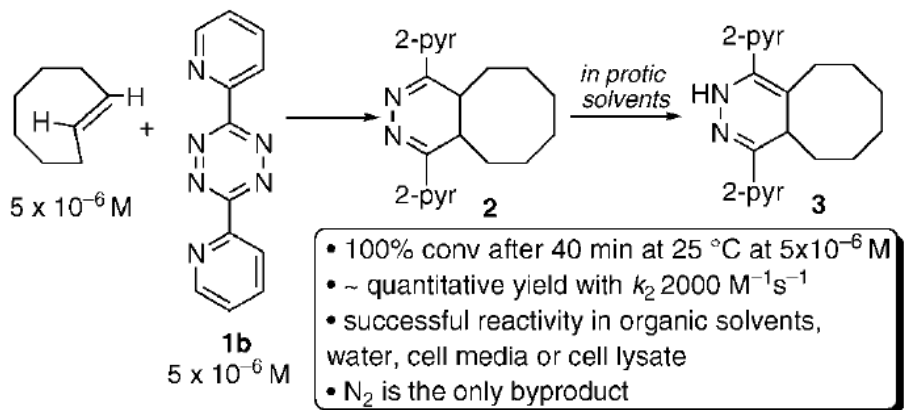
Strain-promoted azide-alkyn cycloaddition (SPAAC)

# Inverse-electron demand Diels-Alder cycloaddition (IEDDA)

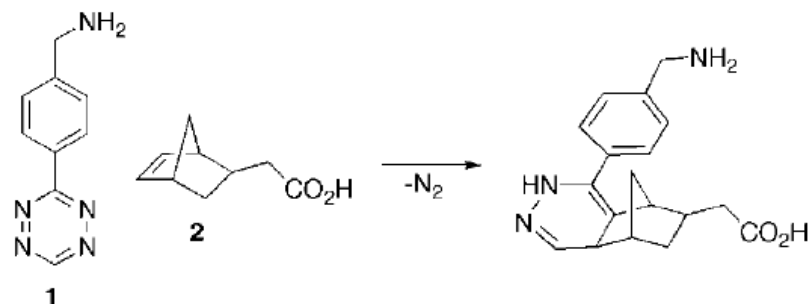
**Scheme 1.** Diels–Alder Reactions of Tetrazines with *trans*-Cyclooctene



**Scheme 2.** Fast Reactivity at Low Micromolar Concentrations



tetrazine + *trans*-cyclooctene



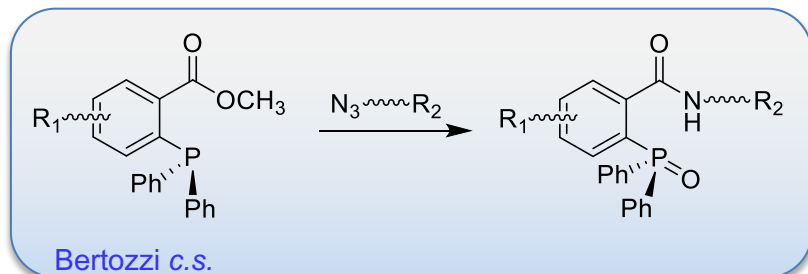
tetrazine + norbornadiene

Devaraj et al., *Bioconjug. Chem.* **2008**, 19, 2297-2299

Reactivity increases with increased strain, EWG-functionalized TZs and in protic solvents

# Bioorthogonal chemistry for in vivo applications

## Staudinger Ligation

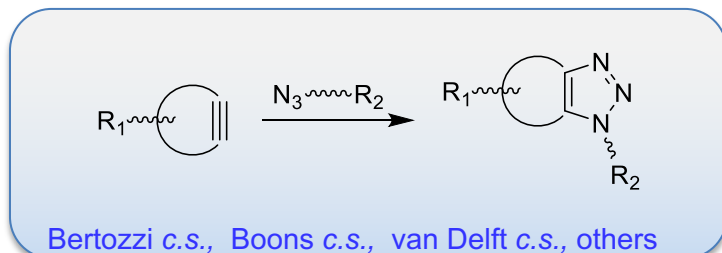


$$K_2 = 2 \times 10^{-3} \text{ M}^{-1} \text{ s}^{-1} (\text{CD}_3\text{CN} + 5\% \text{ H}_2\text{O})$$

$$[\text{A}] = [\text{B}] = 1 \mu\text{M}$$

**15.9 yr**

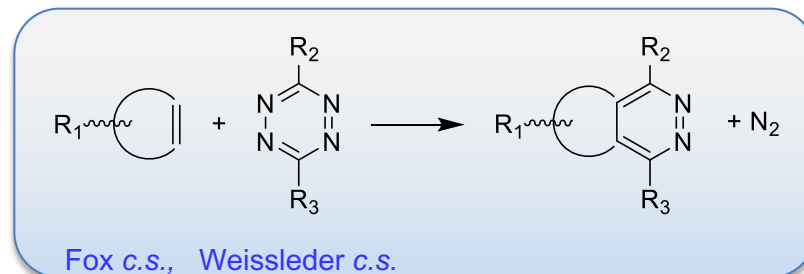
## Strain-Promoted Azide-Alkyn Cycloaddition



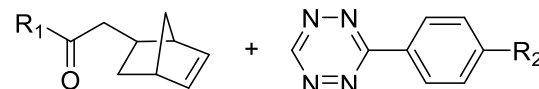
$$K_2 = 4.2 \times 10^{-2} - 1.7 \text{ M}^{-1} \text{ s}^{-1} (\text{CD}_3\text{CN}, \text{MeOH} \text{ or } \text{PBS})$$

**6.8 d – 9.2 mo**

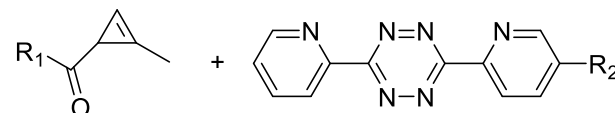
## Inverse-Electron Demand Diels-Alder



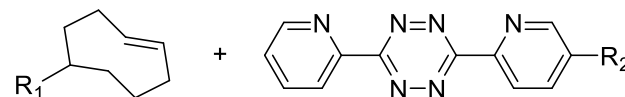
$$K_2 = 1.9 - 2.3 \times 10^6 \text{ M}^{-1} \text{ s}^{-1} (\text{PBS})$$



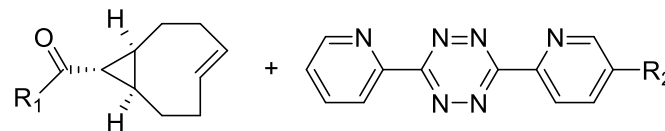
**6.1 d**



**21.4 h**



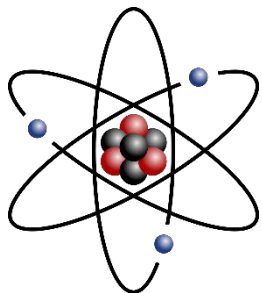
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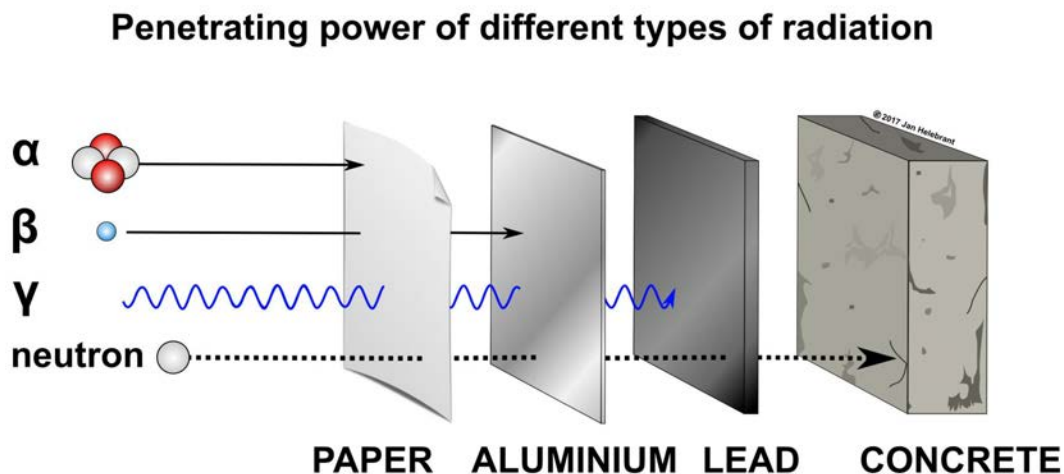
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# Quick introduction to nuclear imaging and “internal” radiotherapy

# Types of radiations for nuclear imaging and “internal” radiotherapy



**Radionuclides** are unstable nuclei that decay to more stable states by emitting some sort of radiation

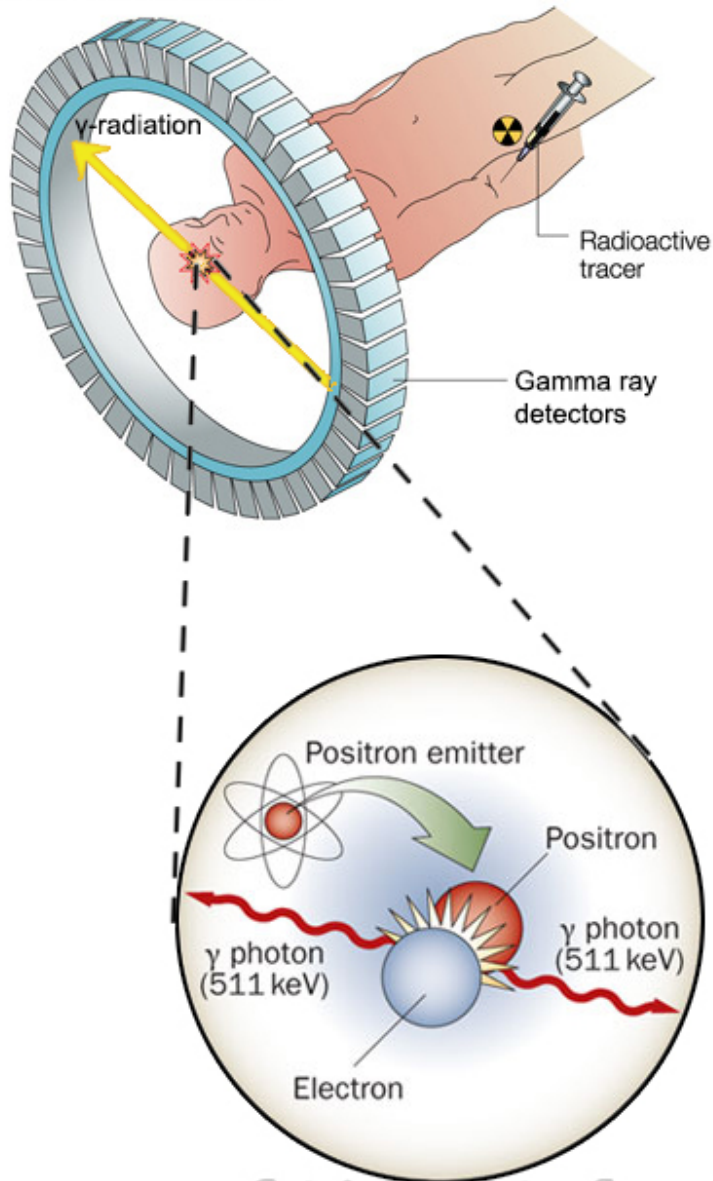


Radionuclides emitting  $\gamma$  ( $^{111}\text{In}$ ,  $^{99\text{m}}\text{Tc}$ , etc) and  $\beta^+$  ( $^{18}\text{F}$ ,  $^{89}\text{Zr}$ , etc) can be used for **Single Photon Emission Computed Tomography (SPECT)** and **Positron Emission Tomography (PET)**, respectively

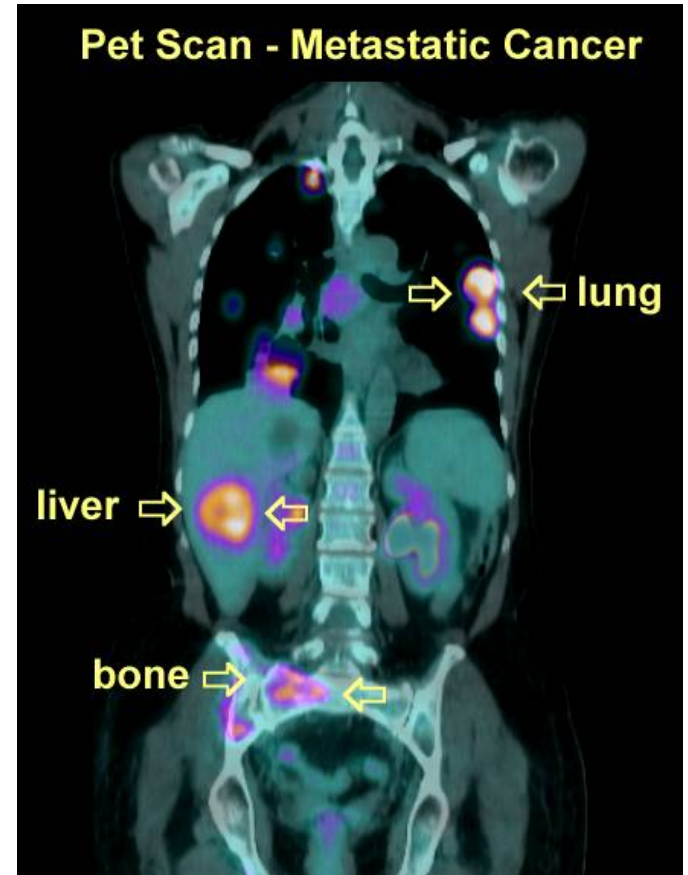
Radionuclides emitting  $\beta^-$  ( $^{177}\text{Lu}$ ,  $^{186}\text{Re}$ , etc) and  $\alpha$  particles ( $^{225}\text{Ac}$ ,  $^{212}\text{Pb}$ ,  $^{223}\text{Ra}$ , etc) can be used for **radiotherapy** of cancer



# Positron Emission Tomography

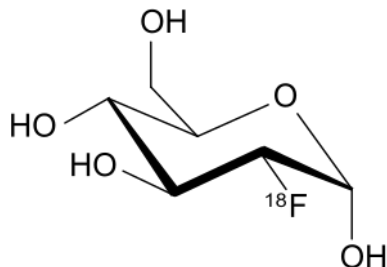


*Gabriel Gonzalez-Escamilla*

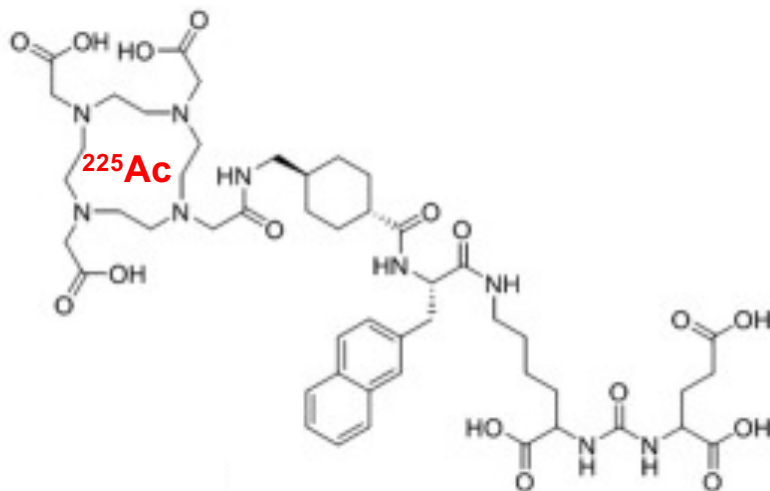




# Cancer radiotherapy



[ $^{18}\text{F}$ ]fluorodeoxyglucose (FDG) for PET imaging of tumor metabolism



$^{225}\text{Ac}$ -PSMA-617 for targeted radiotherapy of PSMA expressing tumors



FDG-PET of a patient with metastatic prostate cancer before and after  $^{225}\text{Ac}$ -PSMA-617 treatment

# Tumor pretargeting for radioimmuno- imaging and -therapy

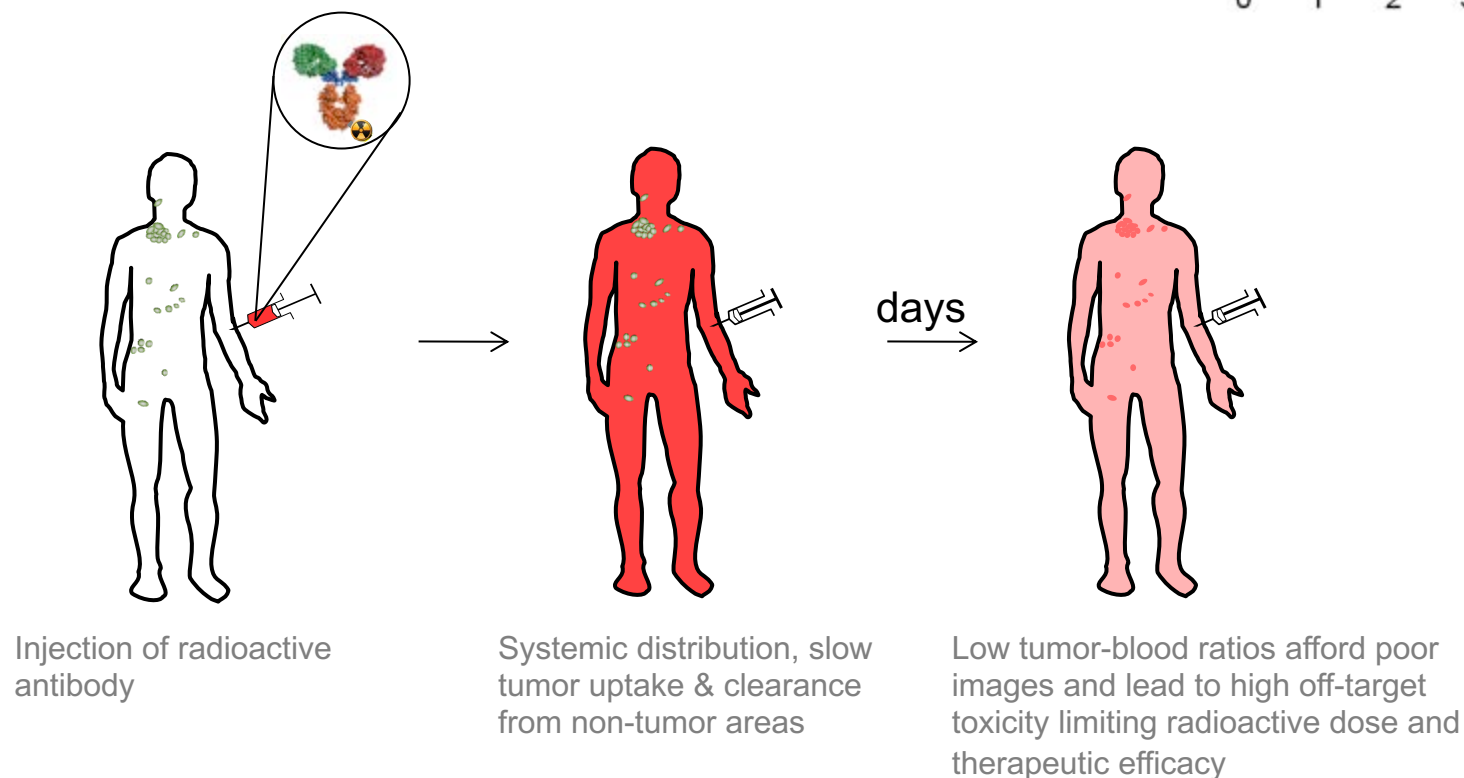
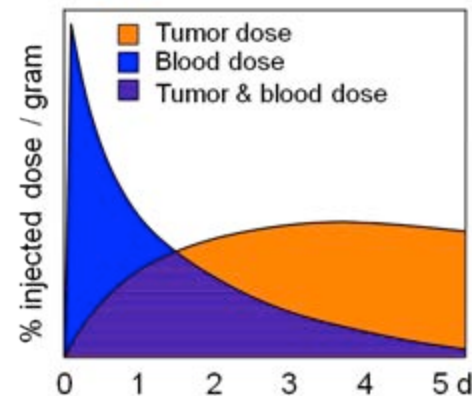


# Directly labeled antibodies for imaging and therapy

Antibodies (mAbs) give superior target uptake compared to fragments

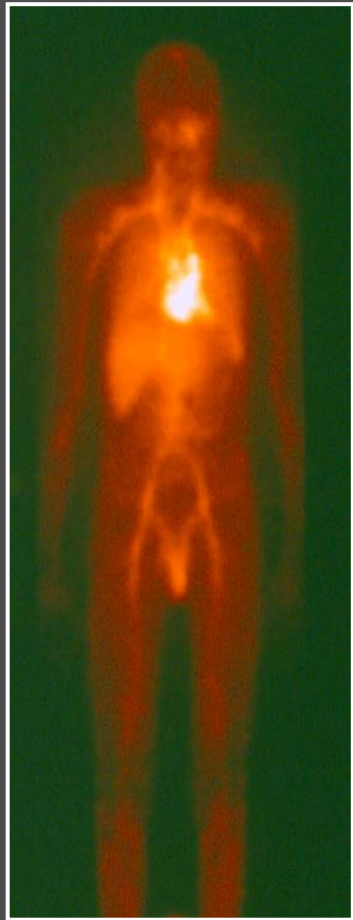
**BUT:**

- They have slow target (e.g. tumor) uptake and slow blood clearance,
- hampering imaging applications
- and solid tumors remain out of reach of radioimmunotherapy (RIT) due to radiation dose-limiting side effects

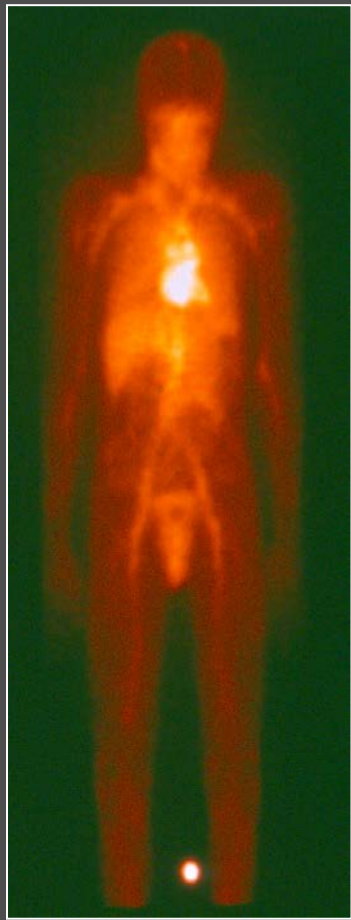


# Conventional radioimmunoimaging & –therapy: low target-blood ratios & high radiation dose to bone marrow

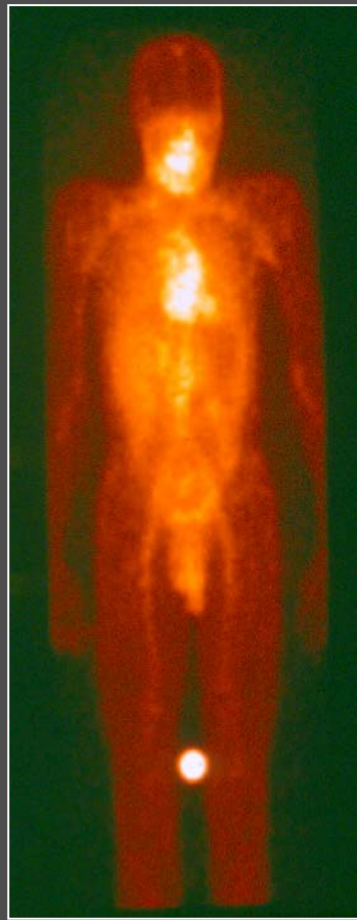
[ $^{186}\text{Re}$ ]Re-U36 in head and neck cancer patient



**< 1 hour**



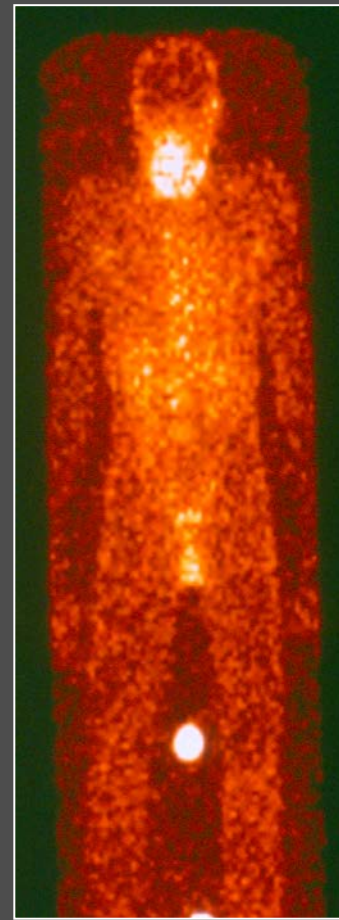
**24 hours**



**48 hours**



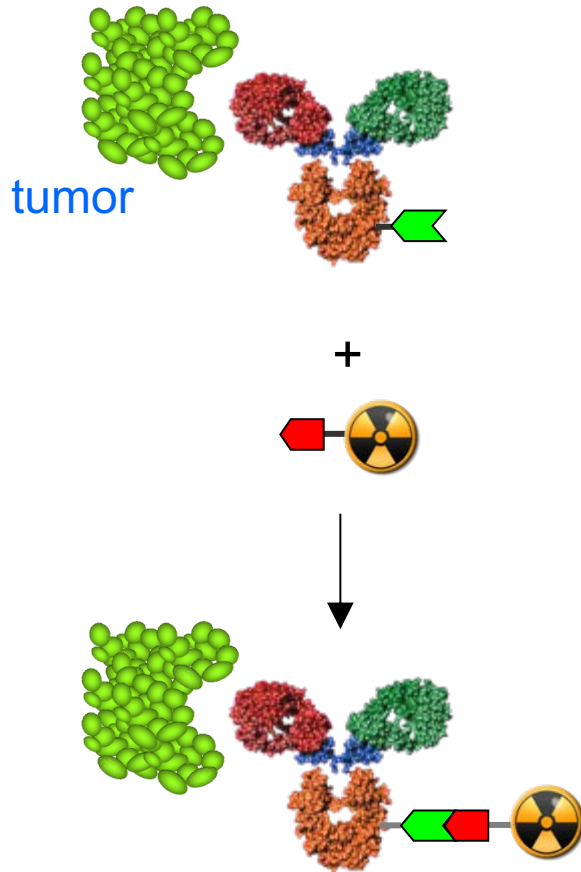
**72 hours**



**312 hours**

# Pretargeting..

..improves radioimmunotherapy and -imaging of tumors via a 2-step tumor targeting scheme



Step 1:  
Slow tumor binding with antibody



Step 2:  
Fast binding with small probe with



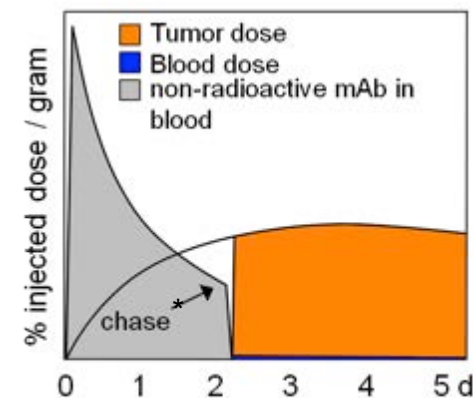
High tumor-background ratio &  
increased efficacy



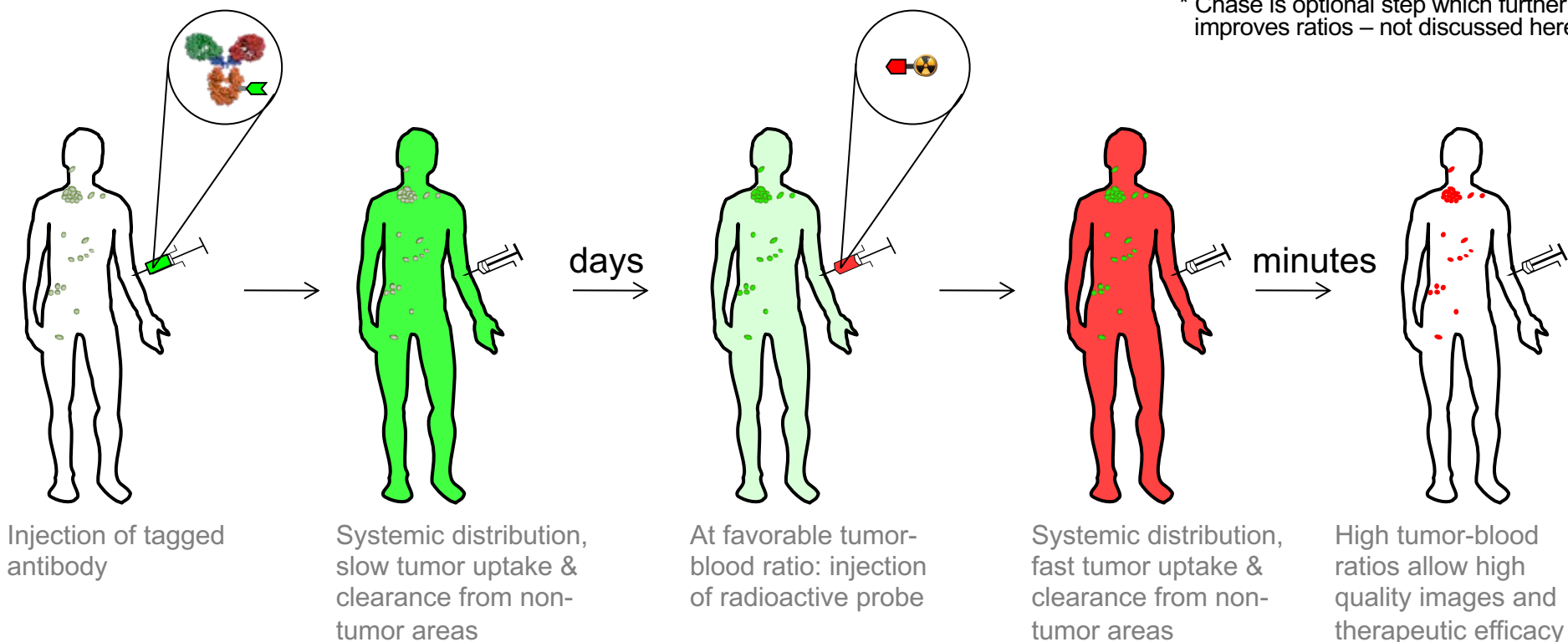
# Pretargeting - general



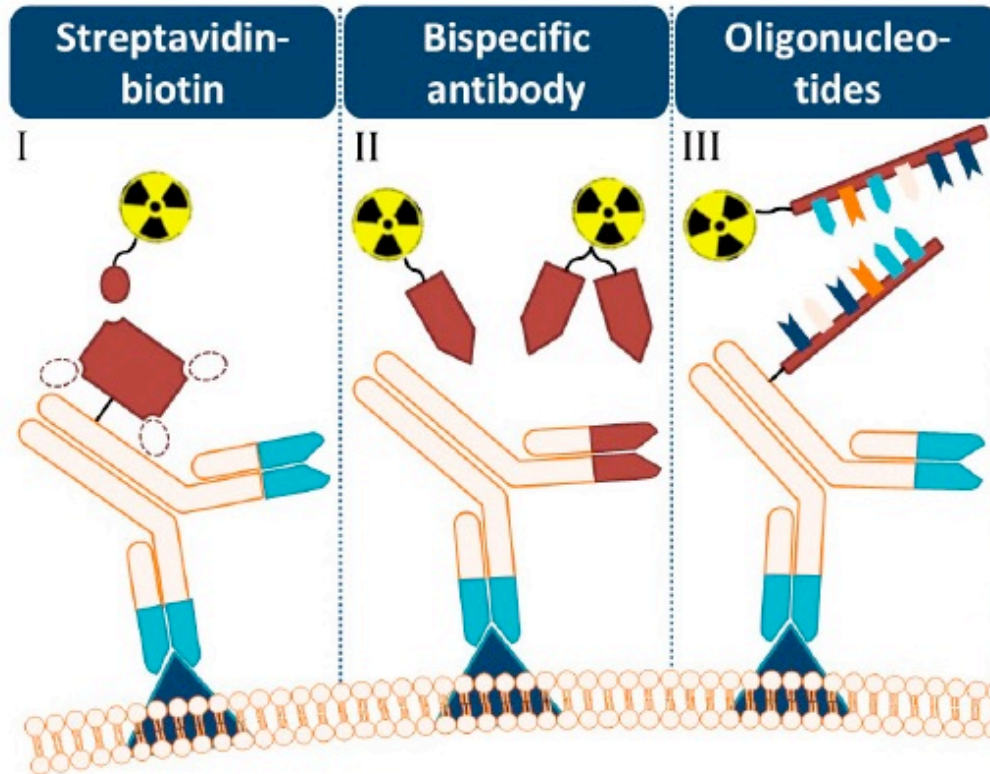
- Overcomes limitations by separating tumor targeting from delivery of radioactive component
- 2 steps: tumor targeting of a “tagged” mAb followed by rapid binding of a small radiolabeled probe to the tumor-bound mAb in 2<sup>nd</sup> step
- Non-bound probe is excreted in a matter of minutes



\* Chase is optional step which further improves ratios – not discussed here



# Pretargeting pairs

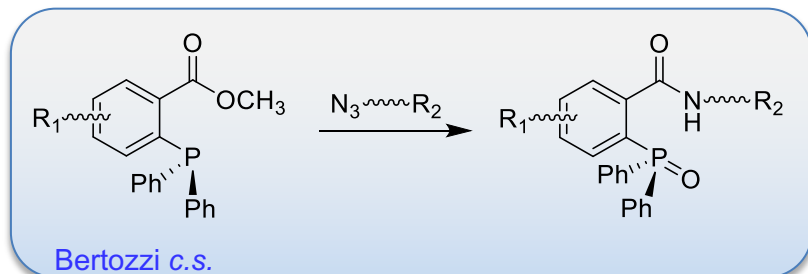




# Chemical approach to tumor pretargeting

# Bioorthogonal chemistry for in vivo applications: **Speed is key**

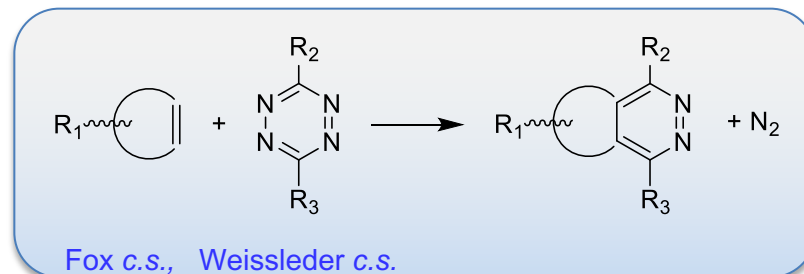
## Staudinger Ligation



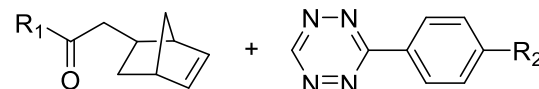
$$K_2 = 2 \times 10^{-3} \text{ M}^{-1}\text{s}^{-1} \text{ (CD}_3\text{CN + 5\% H}_2\text{O)}$$

**[A]=[B]= 1  $\mu$ M**  
**15.9 yr**

## Inverse-Electron Demand Diels-Alder

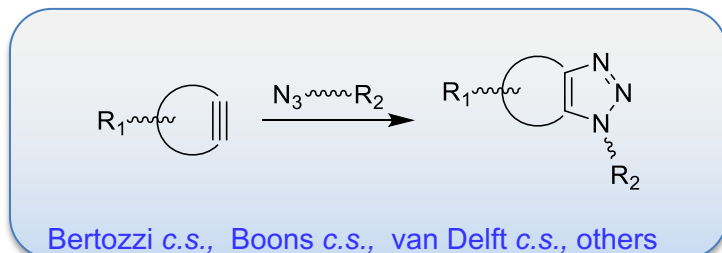


$$K_2 = 1.9 - 2.3 \times 10^6 \text{ M}^{-1}\text{s}^{-1} \text{ (PBS)}$$



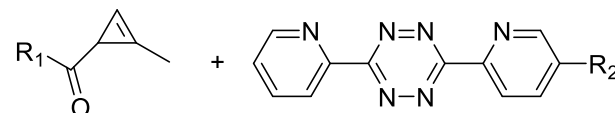
**6.1 d**

## Strain-Promoted Azide-Alkyn Cycloaddition

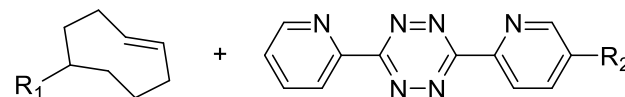


$$K_2 = 4.2 \times 10^{-2} - 1.7 \text{ M}^{-1}\text{s}^{-1} \text{ (CD}_3\text{CN, MeOH or PBS)}$$

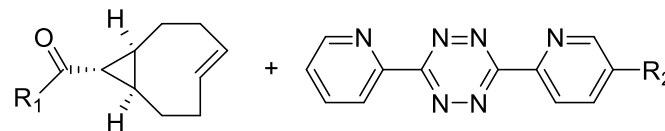
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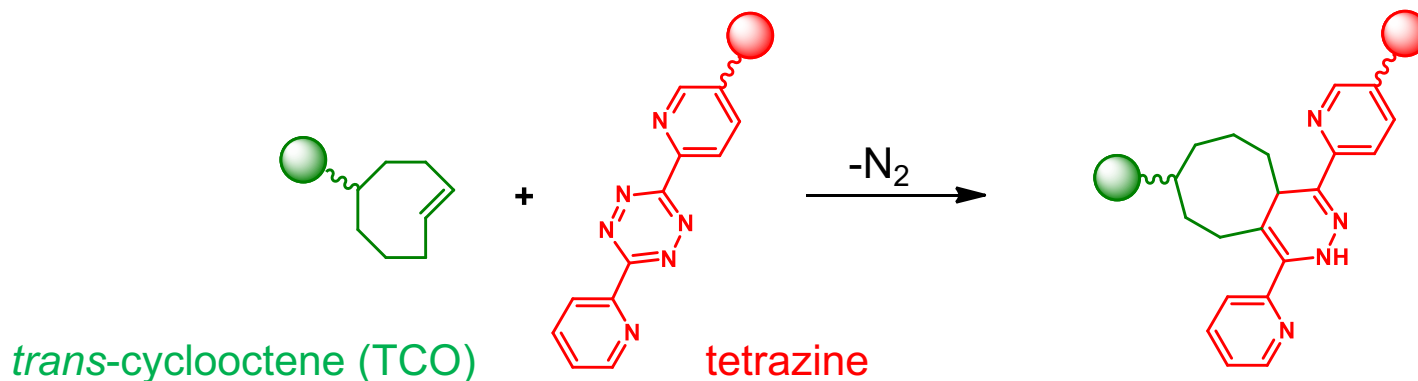
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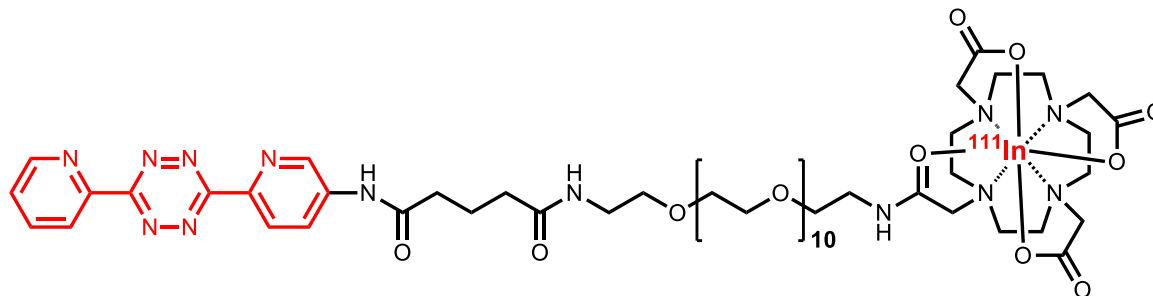
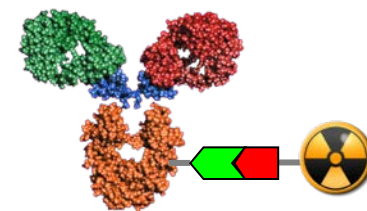
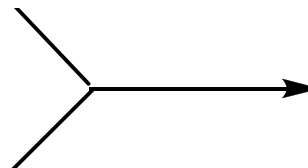
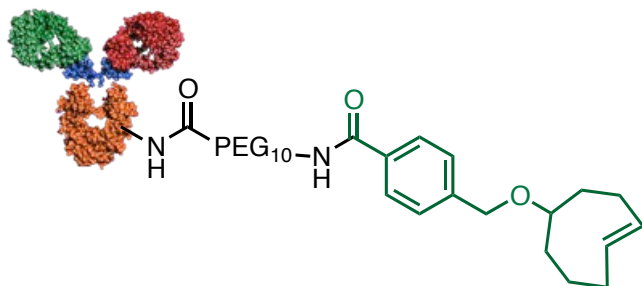
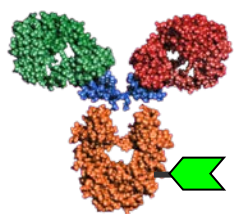
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# Bioorthogonal chemistry for pretargeting

with CC49 antibody against TAG72, a pan-carcinoma marker (colon, breast, ovarian, lung, ..)

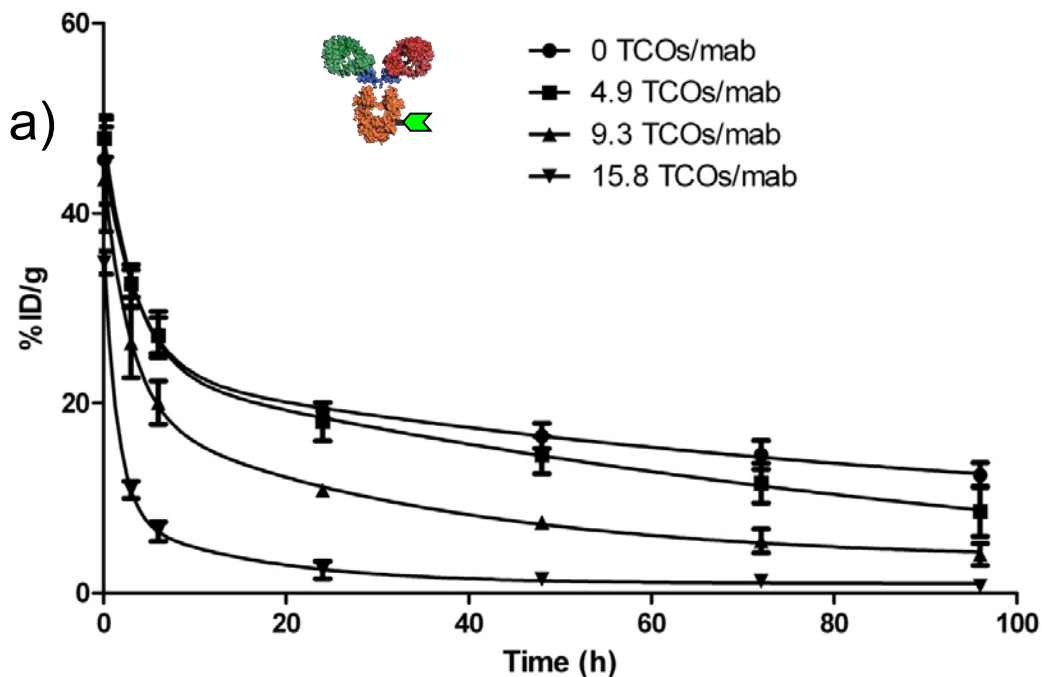


Fox c.s. *J. Am. Chem. Soc.* **2008**, 130, 13518



# Blood clearance of both components

Nude mice (n=4) injected i.v. with a)  $^{125}\text{I}$ -CC49 or  $^{125}\text{I}$ -CC49-TCO tag (100  $\mu\text{g}$ ); or b) with  $^{111}\text{In}$ -DOTA-tetrazine (21  $\mu\text{g}$ )

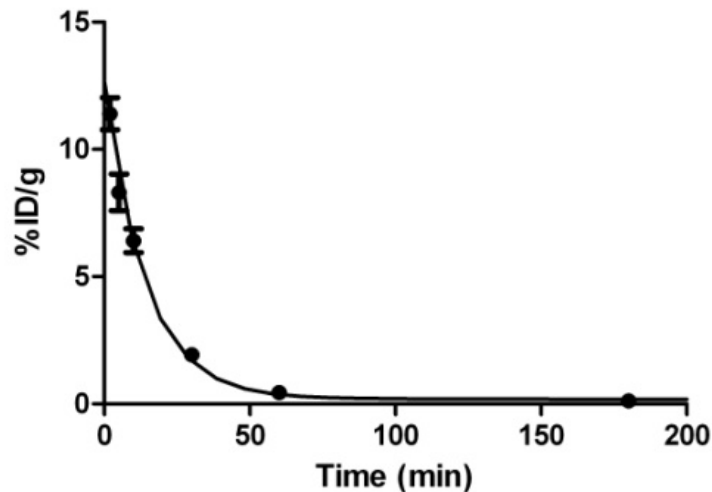


0 TCO  
4.9 TCO  
9.3 TCO  
15.8 TCO

$t_{\alpha} = 2.58 \text{ h}; t_{\beta} = 64.36 \text{ h}$   
 $t_{\alpha} = 2.39 \text{ h}; t_{\beta} = 74.55 \text{ h}$   
 $t_{\alpha} = 1.97 \text{ h}; t_{\beta} = 23.49 \text{ h}$   
 $t_{\alpha} = 1.05 \text{ h}; t_{\beta} = 10.55 \text{ h}$

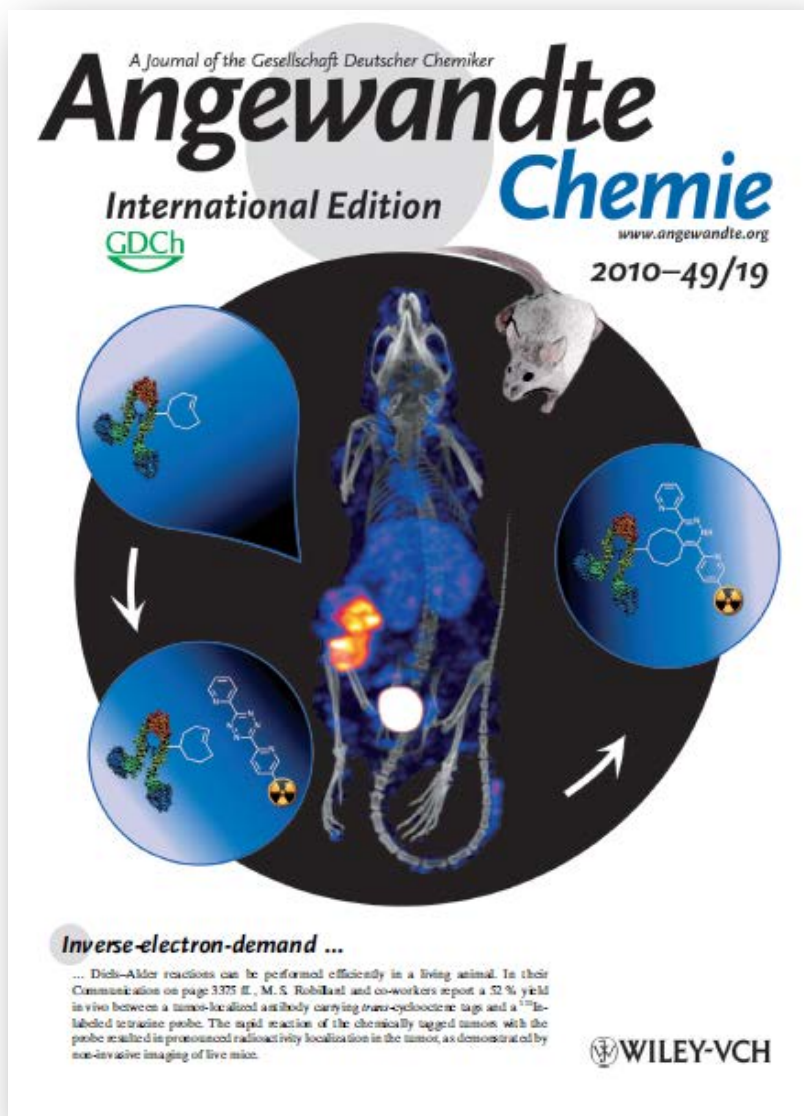
b)

$^{111}\text{In}$ -tetrazine



$t_{1/2} = 9.8 \text{ min}$   
 $V_D = \text{Dose}/C_0 = 0.32 \text{ L/kg}$

# Proof of concept: in vivo reaction in equimolar conditions



Colorectal cancer mouse model (LS174T).  
Injection of CC49-TCO<sub>7</sub> followed 24h later by  
[<sup>111</sup>In]In-tetrazine (25 eq)

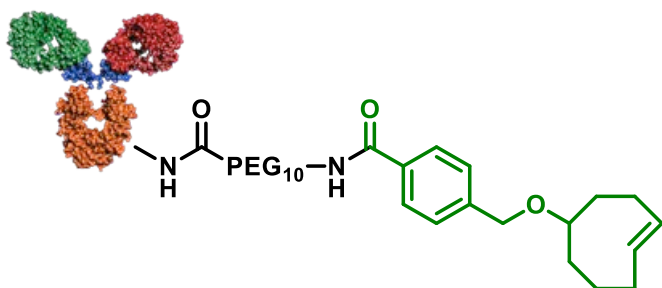
- 52 % reaction yield on the tumor
- TCO in vivo stability:  $t_{1/2} = 2.62 \text{ d}$
- TCO  $k_2 = 2.7 \times 10^4 \text{ M}^{-1} \text{ s}^{-1}$

## Next aims:

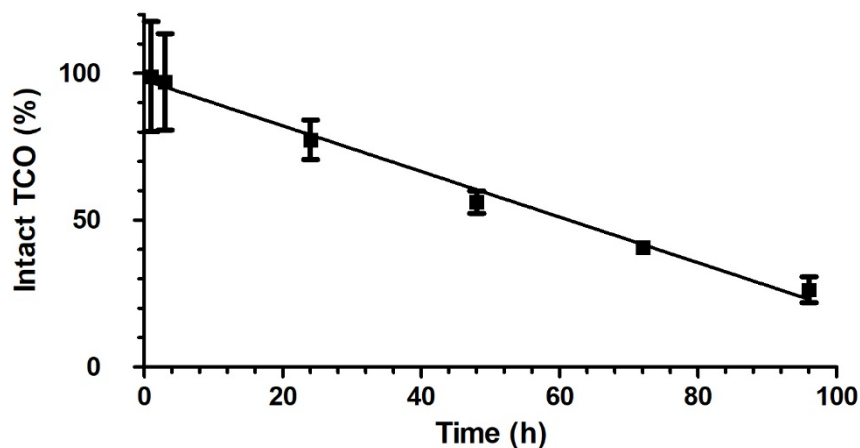
- Improve tag: stability, reactivity, PK
- Improve tumor-blood ratio
- Other targets and targeting agents

## Slow TCO deactivation in circulation

In vivo in mice *trans*-cyclooctene slowly becomes unreactive towards tetrazines

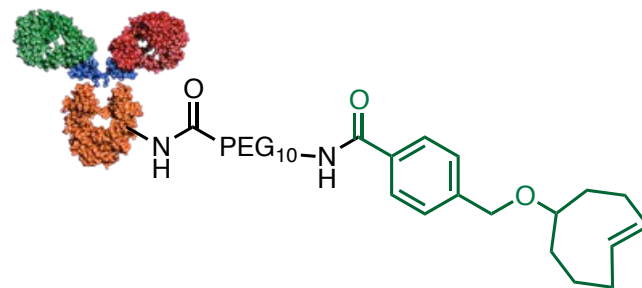
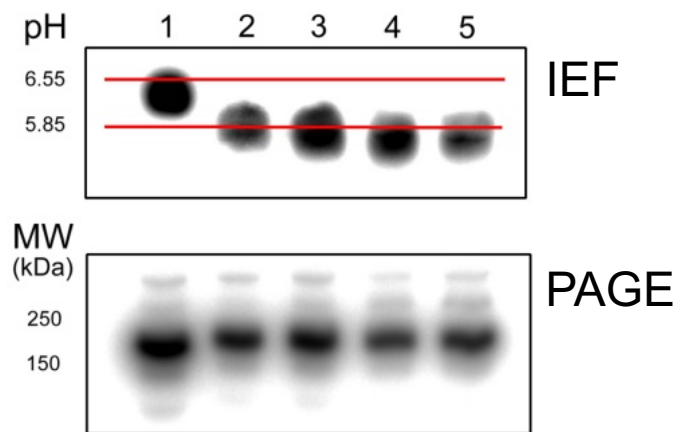


# Half-life 2.62 days



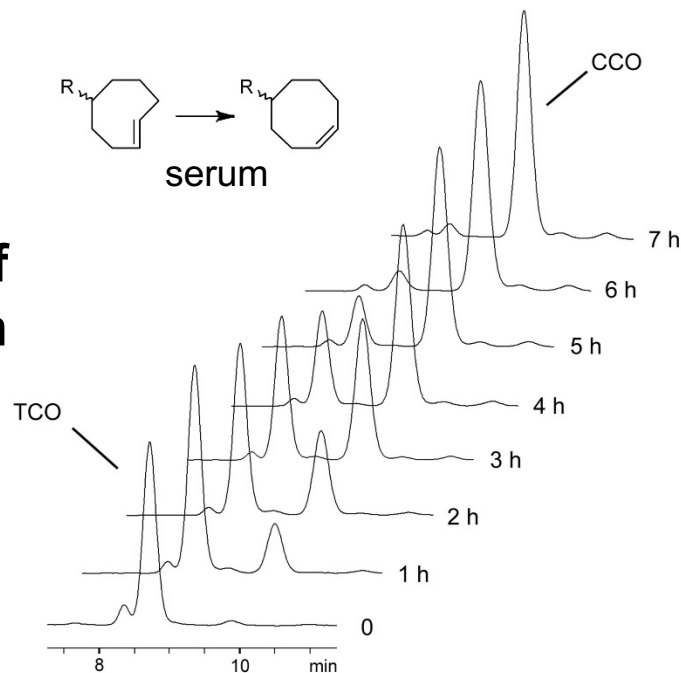
In vivo TCO stability in blood in mice (n=3) assessed ex vivo with  $^{177}\text{Lu}$ -DOTA-tetrazine, corrected for  $^{125}\text{I}$ -ADC blood clearance

# Mechanism of TCO deactivation



- <sup>125</sup>I-CC49 (lane 1, t = 0) and <sup>125</sup>I-CC49-TCO<sub>7</sub> after (lanes 2 to 5) 0, 1 h, 24 h and 48 h incubation at 37 °C in 50% mouse serum.
- Amide bonds in linker are stable

Instead, we found in vitro **isomerization of TCO** to the unreactive *cis*-cyclooctene (CCO) in serum, 37°C





# Stability of the TCO tag

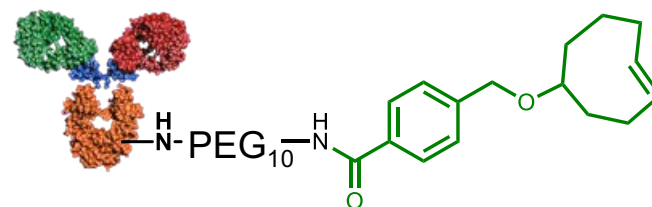
In vitro isomerization TCO to *cis*-cyclooctene (CCO) in serum (components), 37 °C

Medium	$t_{1/2}$
Fresh serum	3.26 h
PBS	>> 7 d
Vitamin B12	>> 7 d
Transferrin	>> 7 d
Ceruloplasmin	6.25 d
Transcuprein	1.39 h

Albumin (MSA)	0.65 h
Metal-depleted albumin (MD-MSA)	2.84 h

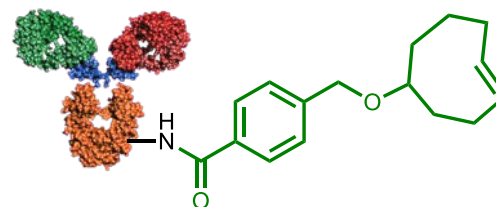
Albumin-bound Cu plays a role in TCO isomerization

By removing the linker TCO is less accessible and stability increases



$$t_{1/2} = 2.62 \text{ d}$$

$$k_2 = 2.7 \times 10^4 \text{ M}^{-1}\text{s}^{-1}$$

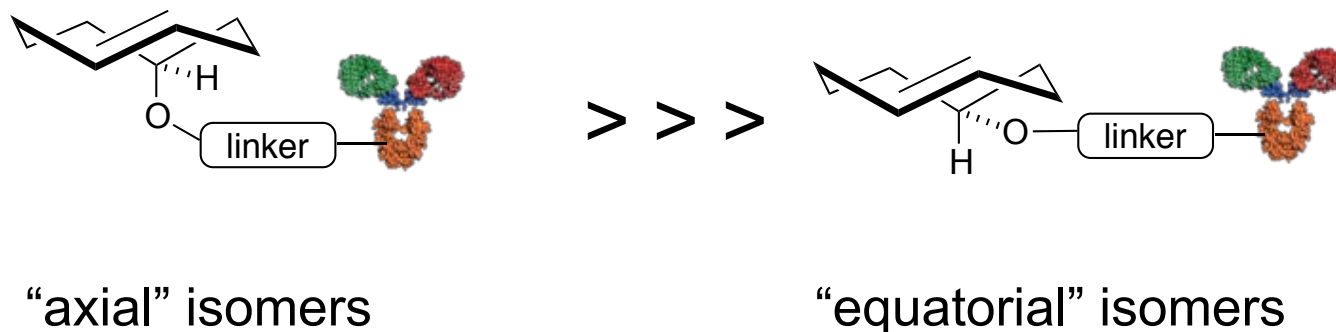


$$t_{1/2} = 6.19 \text{ d}$$

$$k_2 = 3.2 \times 10^4 \text{ M}^{-1}\text{s}^{-1}$$

# Reactivity of the TCO tag

Axial TCOs are 10 times more reactive than the equatorial isomers due to higher energy (strain)

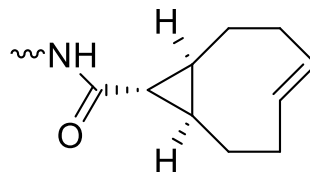


2<sup>nd</sup> order constants  $k_2 \times 10^4 \text{ M}^{-1}\text{s}^{-1}$  between CC49-TCO's & <sup>177</sup>Lu-tetrazine, 37 °C, PBS

TCO structures / isomers					
Axial	27.3±0.5	10.0±1.3	16.3±0.3	14.8±1.2	13.5±0.1
Equatorial	3.2±0.2	1.3±0.1	2.9±0.2	2.5±0.4	2.1±0.0

## Reactivity of the TCO tag

Higher reactivity comes  
at the expenses of in  
vivo instability:  
**need for compromise**



cyclopropyl-TCO  
 $k_2 = 280 \times 10^4 \text{ M}^{-1}\text{s}^{-1}$   
 $t_{1/2} = 0.67 \text{ days}$

$t_{1/2} = 3.94 \text{ days}$

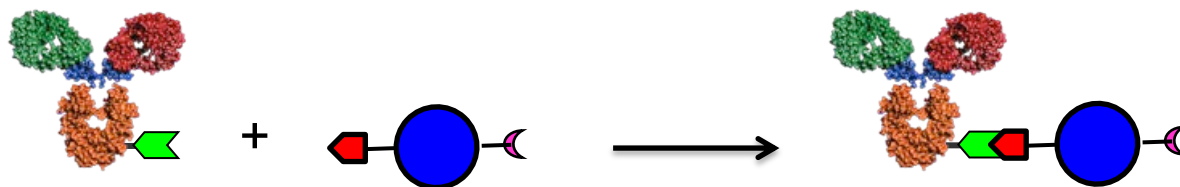
$t_{1/2} = 10.3 \text{ days}$

2<sup>nd</sup> order constants  $k_2 \times 10^4 \text{ M}^{-1}\text{s}^{-1}$  between CC49-TCO's &  $^{177}\text{Lu}$ -tetrazine, 37 °C, PBS

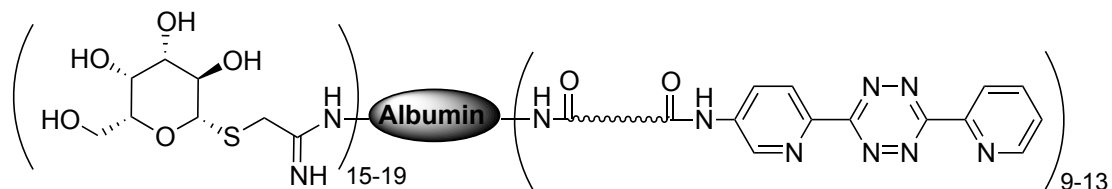
TCO structures / isomers					
Axial	27.3±0.5	10.0±1.3	16.3±0.3	14.8±1.2	13.5±0.1
Equatorial	3.2±0.2	1.3±0.1	2.9±0.2	2.5±0.4	2.1±0.0

# Clearing agents

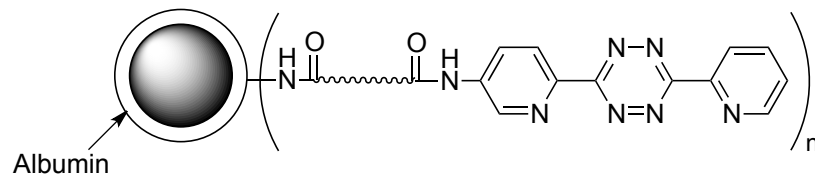
- Tetrazine-functionalized & liver-targeted albumin or microparticles enable instantaneous clearance of tagged constructs from blood
- Removal of residual mAb-tag from circulation before administration of the radioactive probe boosts the target-blood ratio

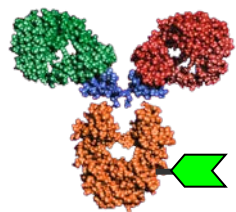
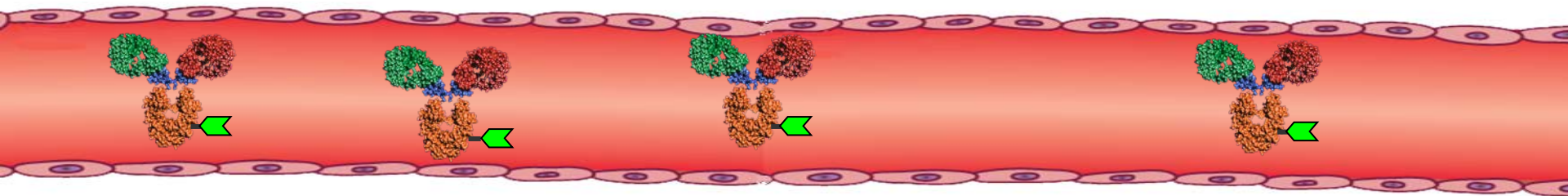


**CA-1:** Galactose/tetrazine-functionalized albumin; hepatocyte Ashwell receptor uptake

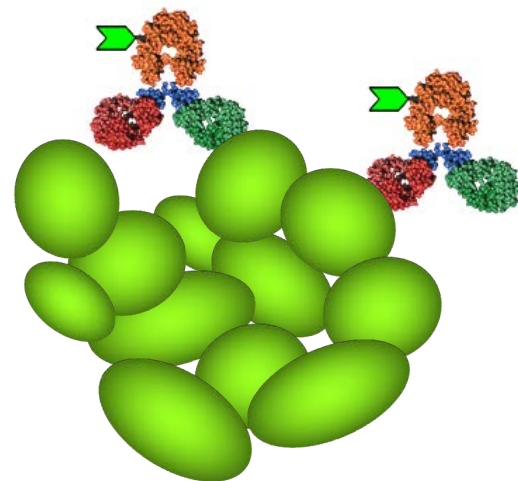


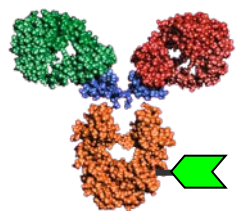
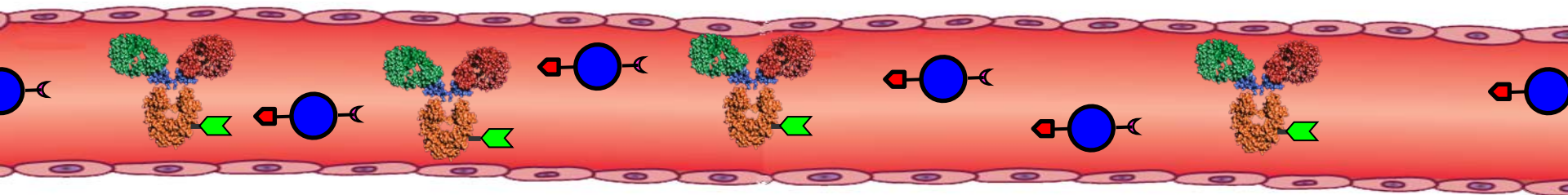
**CA-2:** Microparticles coated with tetrazine-functionalized albumin; size-related RES uptake



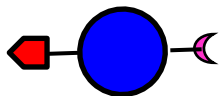


Step 1:  
Slow tumor binding and clearance of antibody



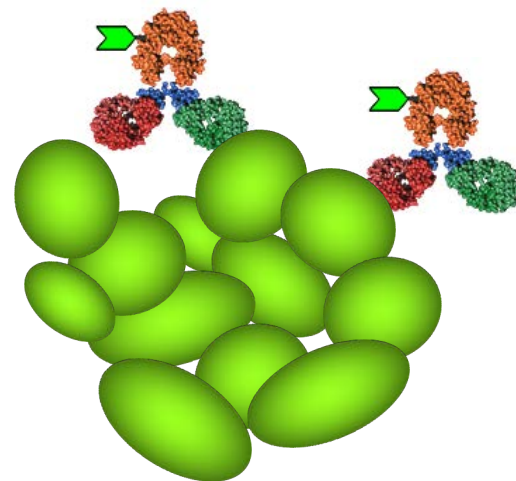


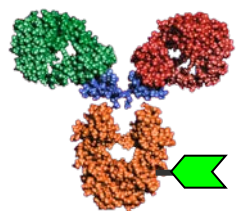
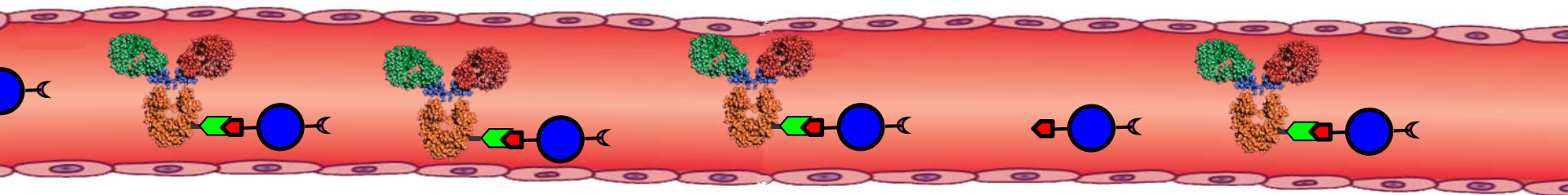
+



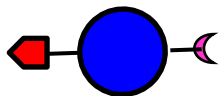
Step 1:  
Slow tumor binding and clearance of antibody

Step 2:  
Administration of clearing agent; fast  
binding with circulating antibody



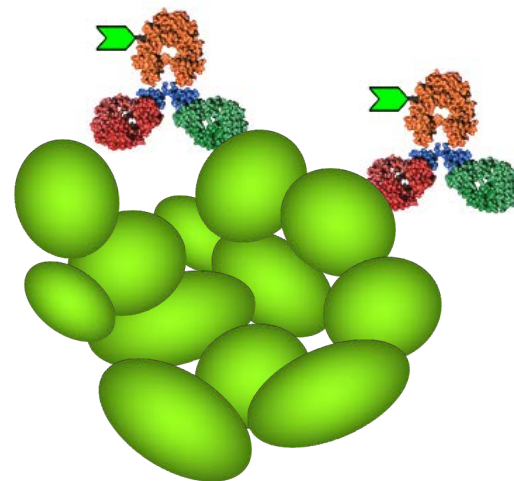


+

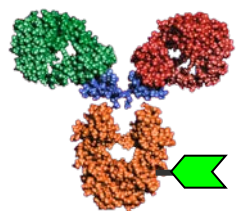
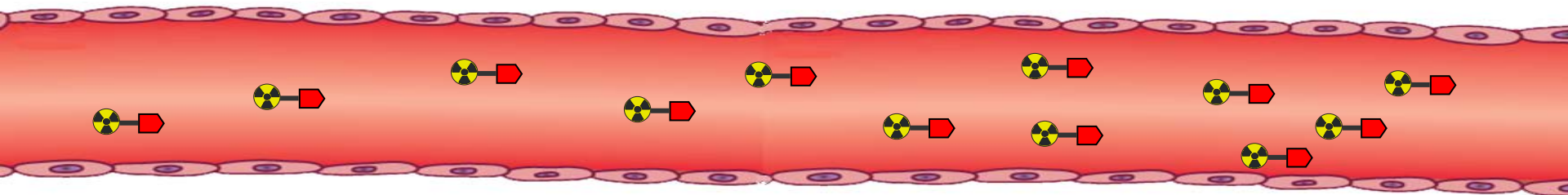


Step 1:  
Slow tumor binding and clearance of antibody

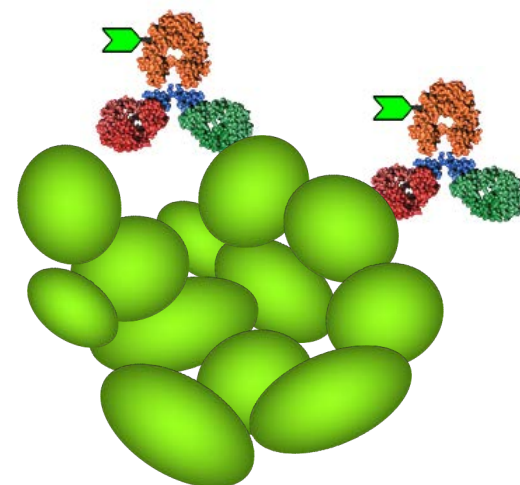
Step 2:  
Administration of clearing agent; fast  
binding with circulating antibody



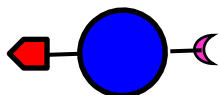




Step 1:  
Slow tumor binding and clearance of antibody



+

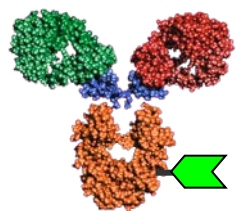
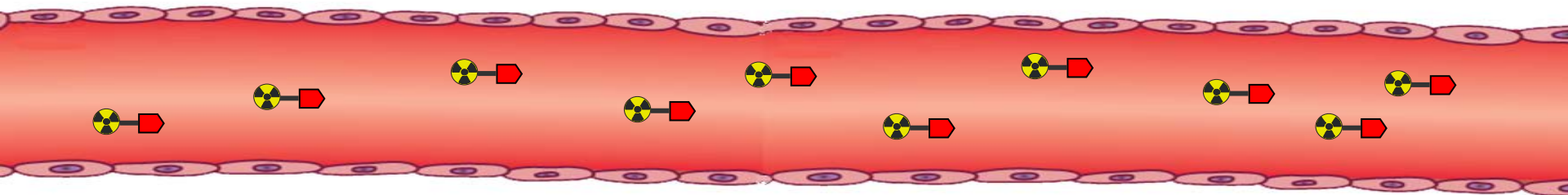


Step 2:  
Administration of clearing agent; fast  
binding with circulating antibody

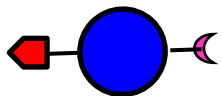
+



Step 3:  
Fast binding with small radiolabeled probe



+



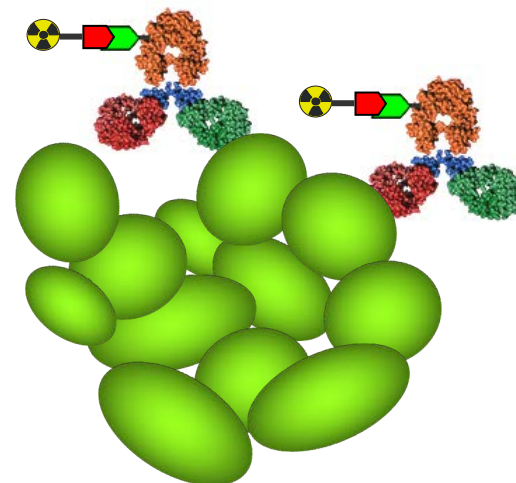
+



Step 1:  
Slow tumor binding and clearance of antibody

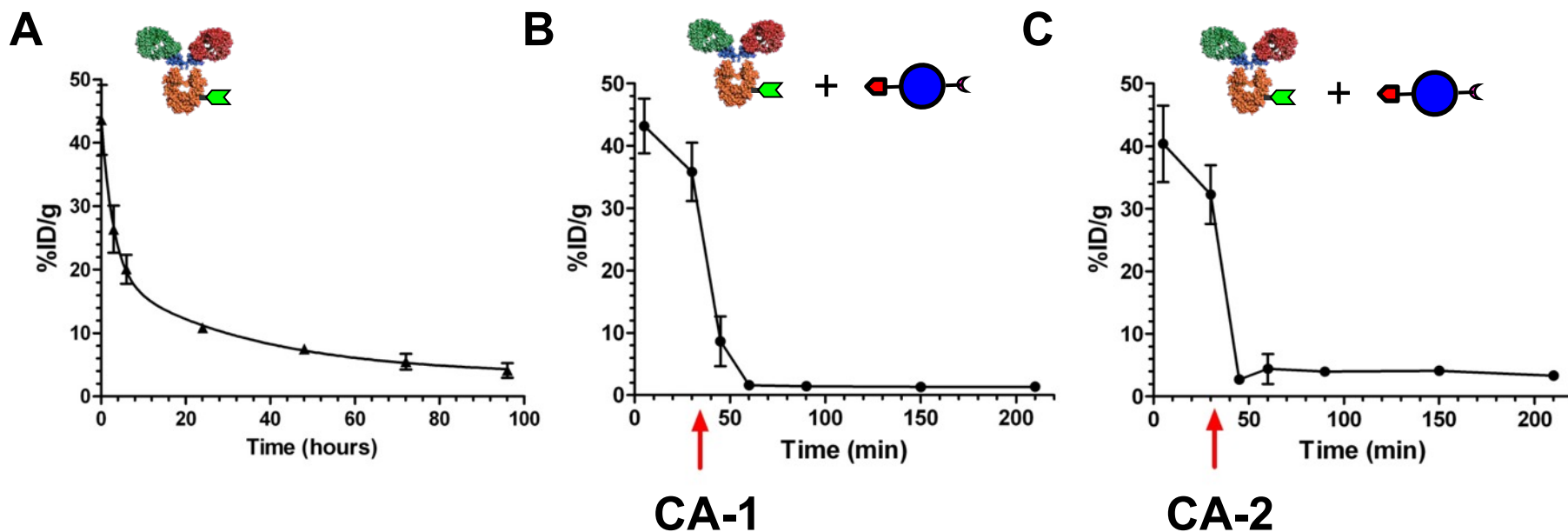
Step 2:  
Administration of clearing agent; fast  
binding with circulating antibody

Step 3:  
Fast binding with small radiolabeled probe



# Blood clearance of $^{125}\text{I}$ -CC49-TCO

Tumor-free mice (n=3) injected with (A) 100  $\mu\text{g}$  [ $^{125}\text{I}$ ]I-CC49-TCO<sub>9</sub>, (B) 20  $\mu\text{g}$  [ $^{125}\text{I}$ ]O-CC49-TCO<sub>9</sub> followed by 120  $\mu\text{g}$  albumin CA-1, (C) 20  $\mu\text{g}$  [ $^{125}\text{I}$ ]I-CC49-TCO<sub>9</sub> followed by  $4 \times 10^7$  particles CA-2

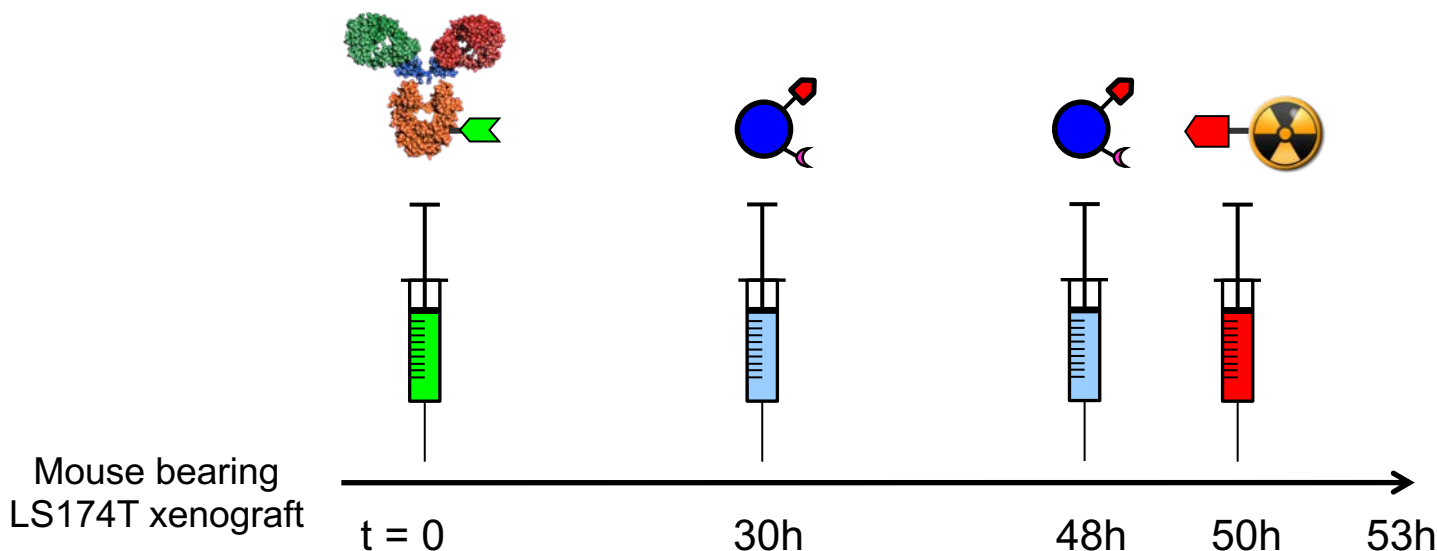


- Highly reactive towards mAb-tag in blood
- Efficient clearance of mAb-tag from blood through liver
- No extravasation into tumor
- No formation of small tag-reactive metabolites

# Optimized pretargeting protocol

Injection of 100  $\mu\text{g}$  CC49-TCO<sub>7</sub>, clearing agent (160  $\mu\text{g}$ ; 30 & 48 h), [<sup>111</sup>In]In-tetrazine (10 eq to CC49; 50 h), image @ 53 h

SPECT/CT



➤ 46 % yield; product stable in vivo > 1 week

➤ **7 day <sup>177</sup>Lu dosimetry: 8-fold higher tumor dose (171Gy) vs. [<sup>177</sup>Lu]Lu-CC49**

T/B: 254 (ex vivo)

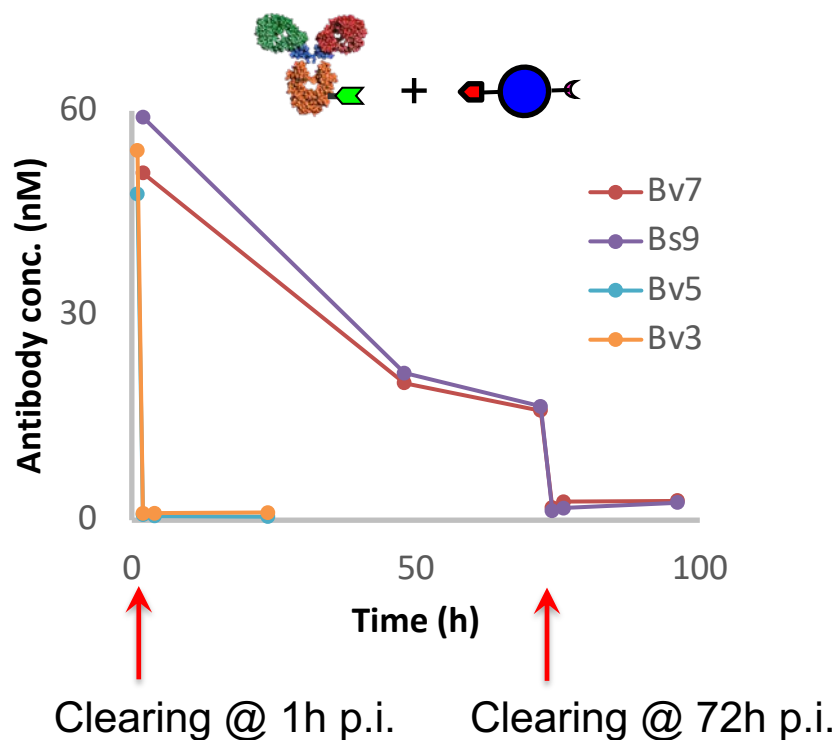
T/M: 190

T/L: 55

T/K: 8

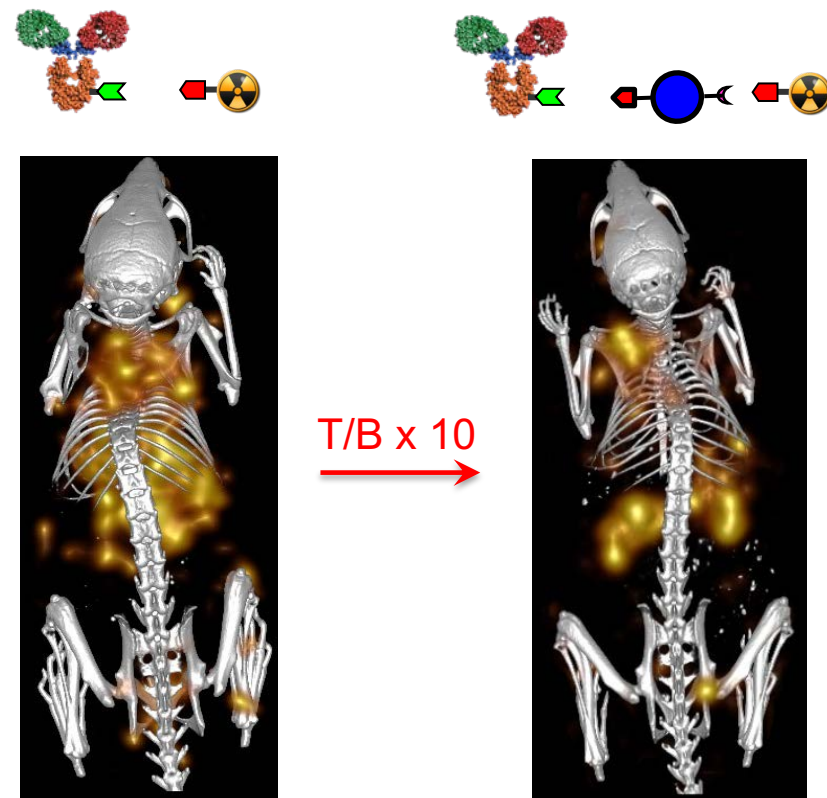
# Triggered clearance of other mAbs

Amyloid-beta mAb for Alzheimer imaging



Collaboration with Syvänen, Uppsala Univ

Prostate cancer stem cells (PC3MA3)  
with anti-CD44v6-TCO &  $^{68}\text{Ga}$ -NOTA-Tz



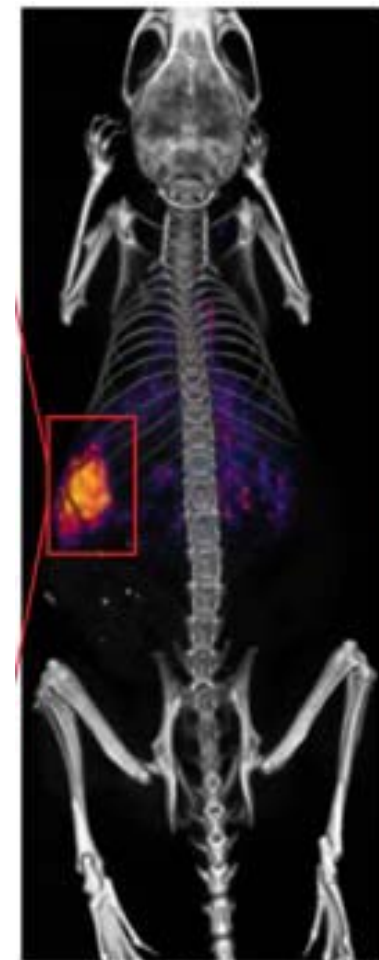
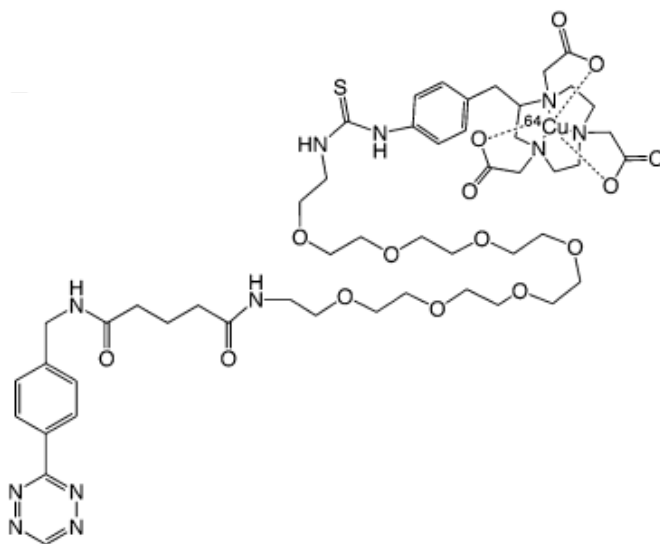
Clear 48h, probe 50h, image 53 h

Collaboration with Quinn, Missouri Univ

# Pretargeted mAb imaging without clearing agent

Orthotopic CA19.9+ Capan-2 pancreas xenograft; 200  $\mu$ g 5B1-TCO + 1.1 eq  $^{64}\text{Cu}$ -NOTA-PEG7-Tz 72h later.

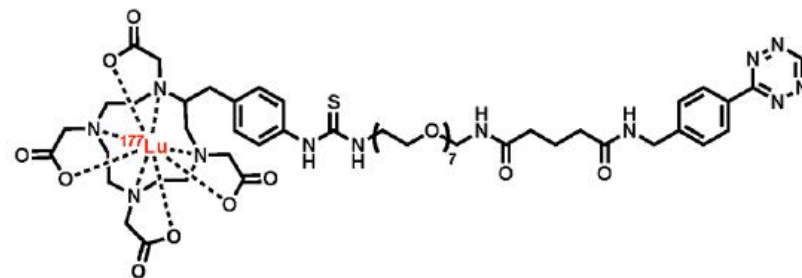
- Rapid shedding and moderate internalization of target
- Tetrazine tumor uptake:  $8.2 \pm 1.7$  %ID/g; T/B = 4
- Dosimetry: >25-fold reduction in total body radiation exposure relative to  $^{89}\text{Zr}$ -DFO-5B1; >70-fold when using  $^{68}\text{Ga}$  instead of  $^{64}\text{Cu}$





# Pretargeted $\beta^-$ radioimmunotherapy

S.c. A33+ SW1222 xenograft; 100  $\mu$ g huA33-TCO followed by 1.4 eq [ $^{177}\text{Lu}$ ]Lu-DOTA-PEG7-Tz 24 h later.

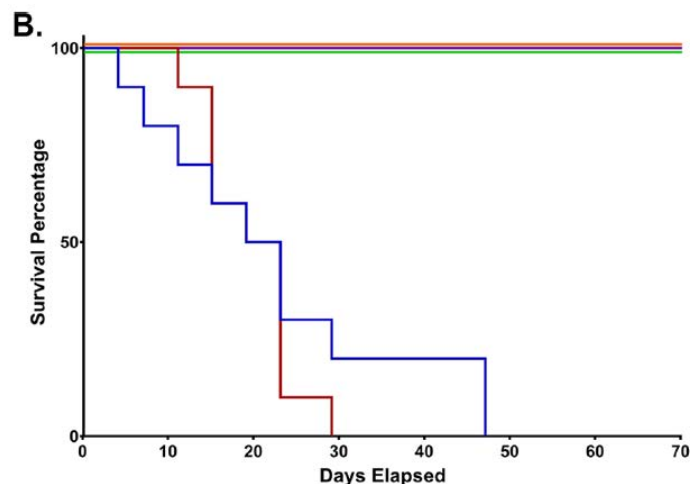
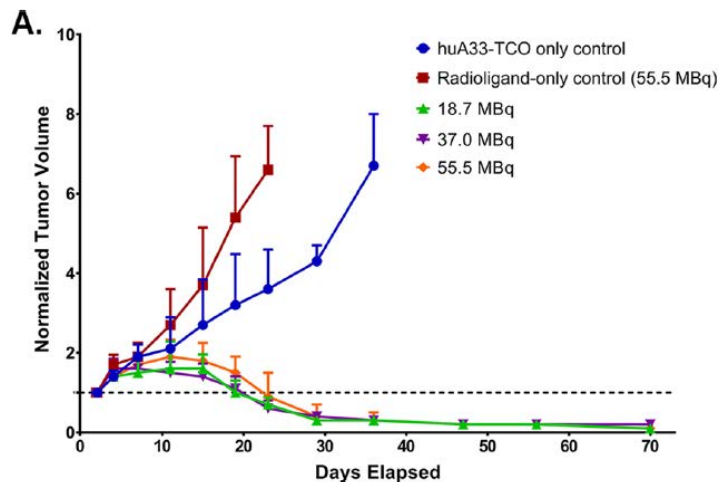


Complete remission already at the lowest dose (18.5 MBq)

Controls:  
Saline and [ $^{177}\text{Lu}$ ]Lu-DOTA-Tz alone (55.5 MBq)

**Directly labeled mAb is missing**

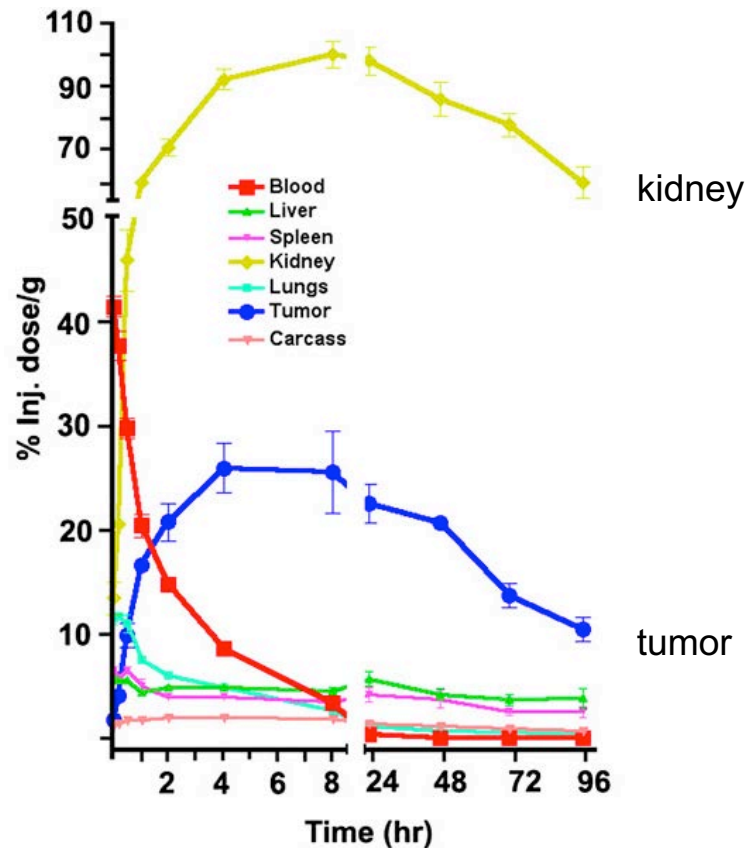
Dosimetry: 2-to-3 lower dose to bone marrow and osteogenic cells





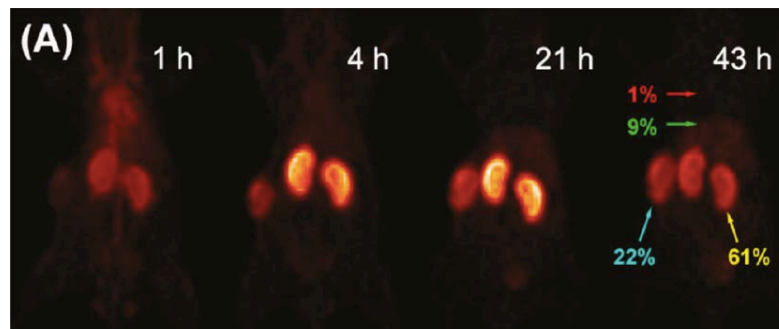
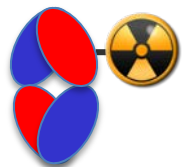
# Tumor PT with a CC49 diabody (50 kDa)

- Radiolabeled diabody-DOTA gives very poor tumor-to-kidney ratios
- Pretargeting may address this, enabling radioimaging and –therapy



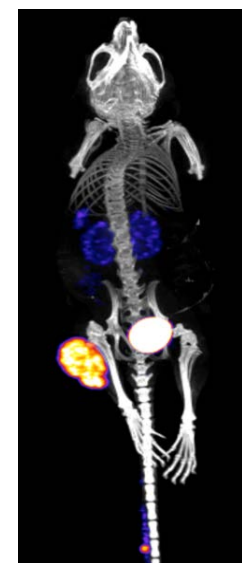
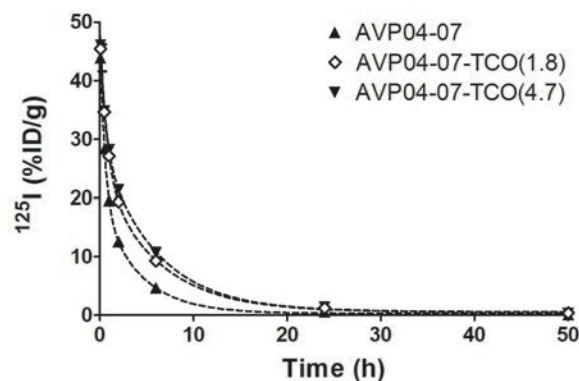
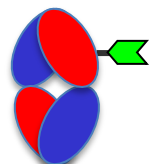
# Pretargeting improves tumor-kidney ratios for radiolabeled proteins

$^{64}\text{Cu}$ -DOTA-CC49 diabody (55 kDa)



TCO-CC49 diabody +  $^{111}\text{In}$ -tetrazine

LS174T-tumored mice

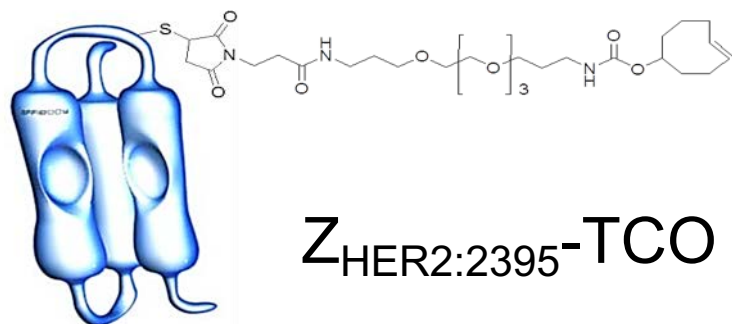


t = 0

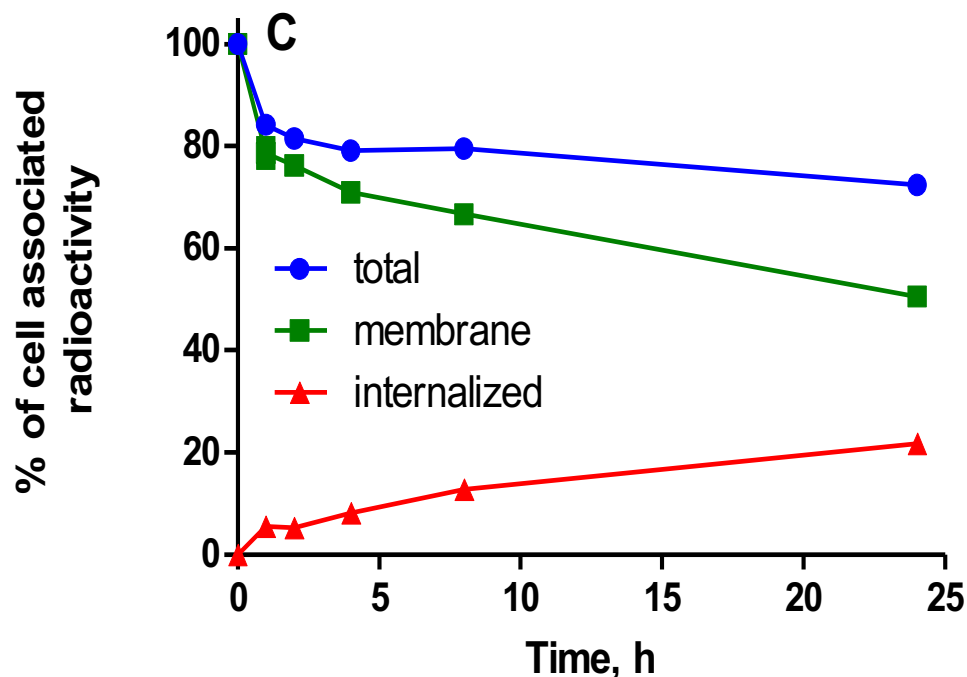
48h

50h

- Radiolabeled Affibody-DOTA gives very poor tumor-to-kidney ratios
- Pretargeting may address this, enabling radioimaging and –therapy
- Affibodies internalize slower than full mAbs – pretargeting with dynamic targets ?

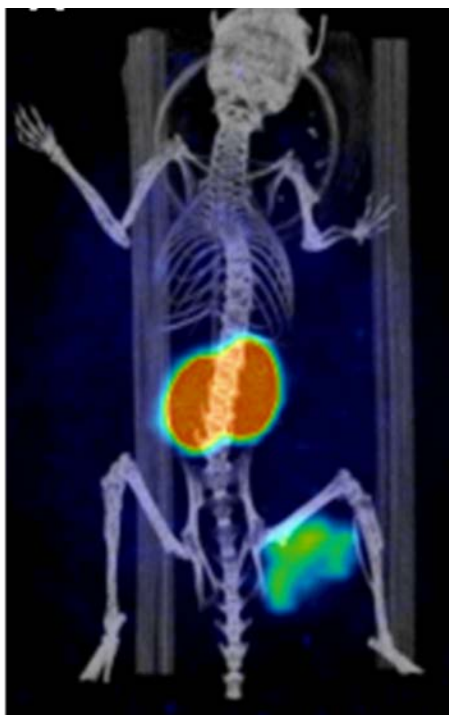


Cellular processing of  $Z_{2395}\text{-TCO}$  in SKOV-3 cells



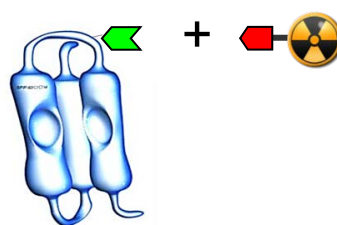
# Pretargeting improves tumor-kidney ratios for radiolabeled peptide

anti-HER2 Affibody-DOTA- $^{111}\text{In}$  (7kD)



Tumor/kidney: 0.06

Affibody-TCO +  $^{111}\text{In}$ -Tz

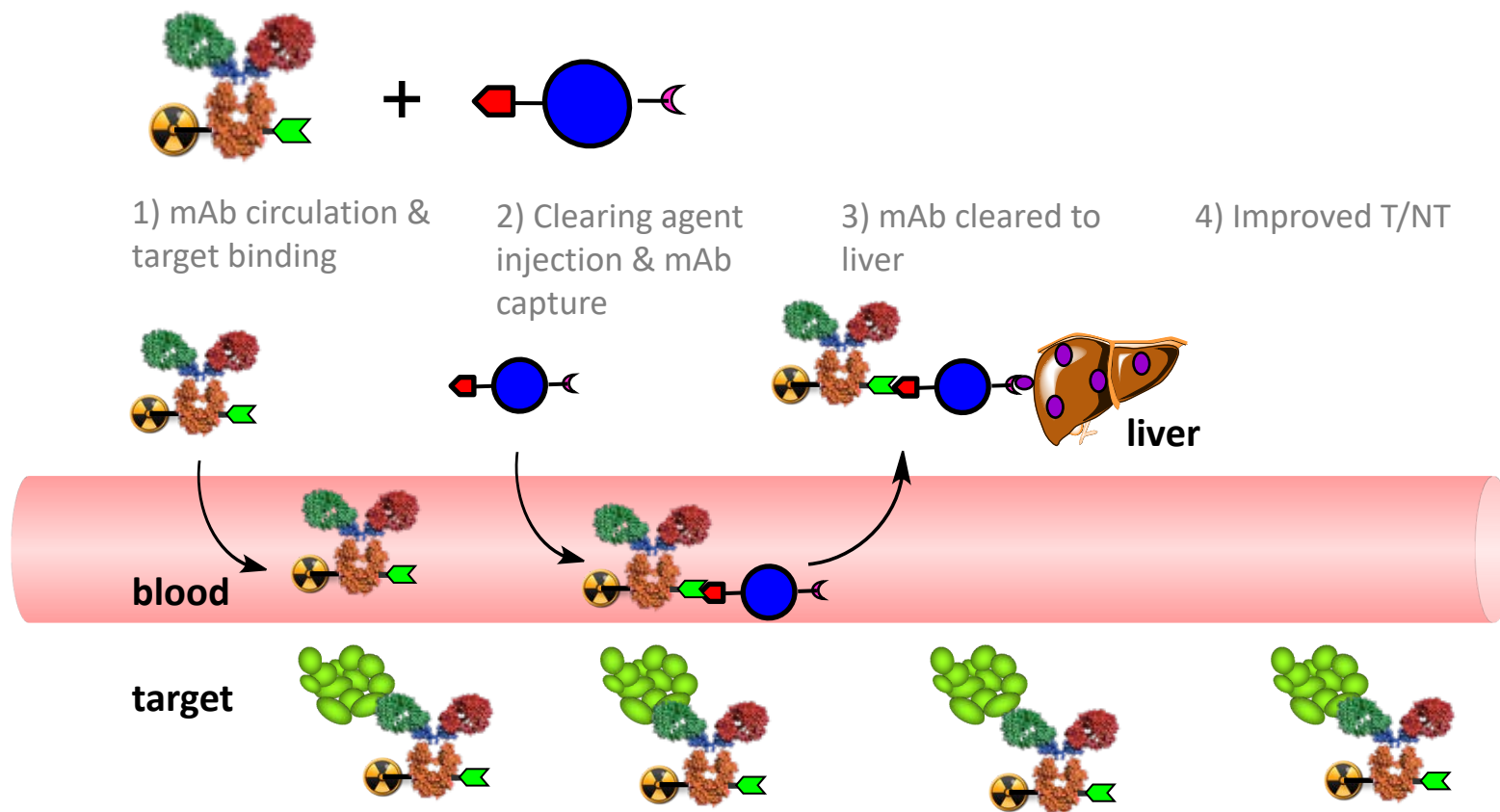


Tumor/kidney: 2

SKOV3 tumored mice: 30  $\mu\text{g}$   $\text{Z}_{2395}\text{-TCO}$ , 1 eq  $^{111}\text{In}$ -DOTA-tetrazine (t = 4h); imaging @ 5 h

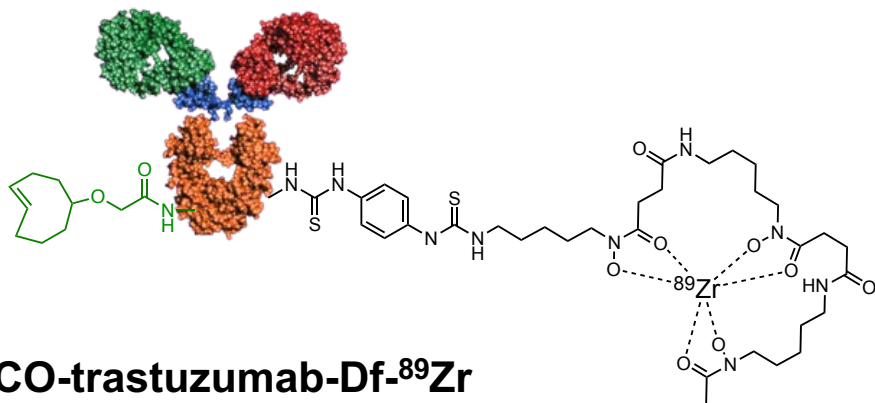
# Clearing radiolabeled mAbs

- Removal of tagged & labeled (e.g.  $^{89}\text{Zr}$ ,  $^{124}\text{I}$ ) mAbs to boosts target-blood ratios, also for internalizing mAbs
- Well suited to improve brain-blood ratios of BBB-crossing mAbs, e.g. in (pre)clinical imaging & development of Alzheimer mAbs

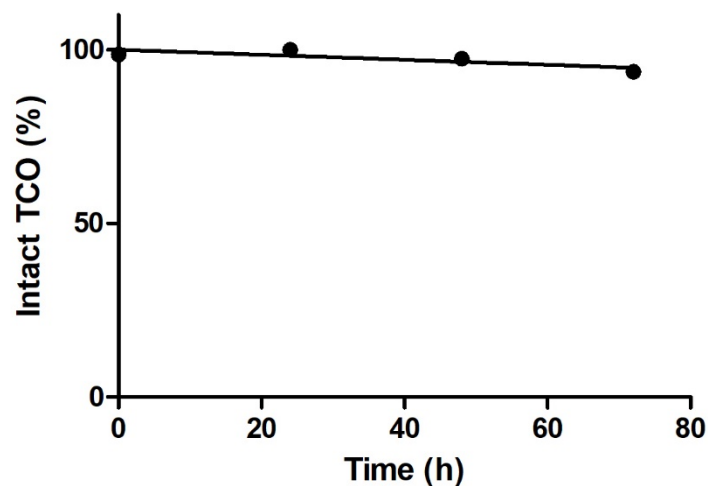


# Clearing radiolabeled mAbs

- Removal of tagged & labeled (e.g.  $^{89}\text{Zr}$ ,  $^{124}\text{I}$ ) mAbs to boost target-blood ratios
- Partnership with biopharma on BBB-crossing mAbs



## Stability in stock solution



TCO is stable in  $^{89}\text{Zr}$ -labeling conditions.

$^{89}\text{Zr}$ -labeled TCO-mAb-Df solutions can be stored at  $+4^\circ\text{C}$  for days (or can be shipped) without losses of TCO reactivity.

## Summary - Click

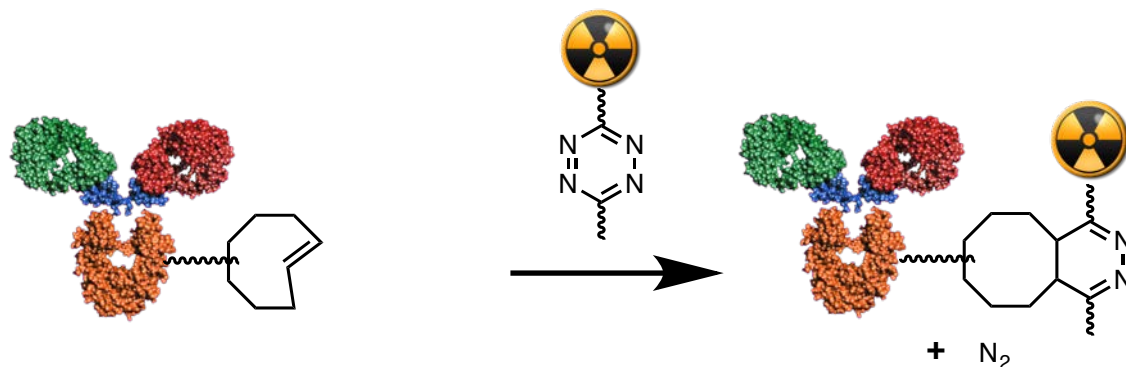
- Selective binding of radioactive probe to chemical tag of antibody via bioorthogonal reaction
- Very fast coupling system (up to  $k_2 \sim 3 \times 10^5 \text{ M}^{-1}\text{s}^{-1}$ ) approaching streptavidin-biotin. High in vivo stability of tag ( $t_{1/2}$  10 days) and reaction product ( $\gg 1$  week)
- Boosts target-to-blood ratios: improved imaging for e.g. companion diagnostics ( $^{18}\text{F}$ -tetrazines), and increased tumor dose in radioimmunotherapy
- Low likelihood of immunogenicity compared to biological pretargeting components: repetition
- Universal & straightforward tag conjugation with minimal perturbation: antibodies, fragments, peptides, particles, ..



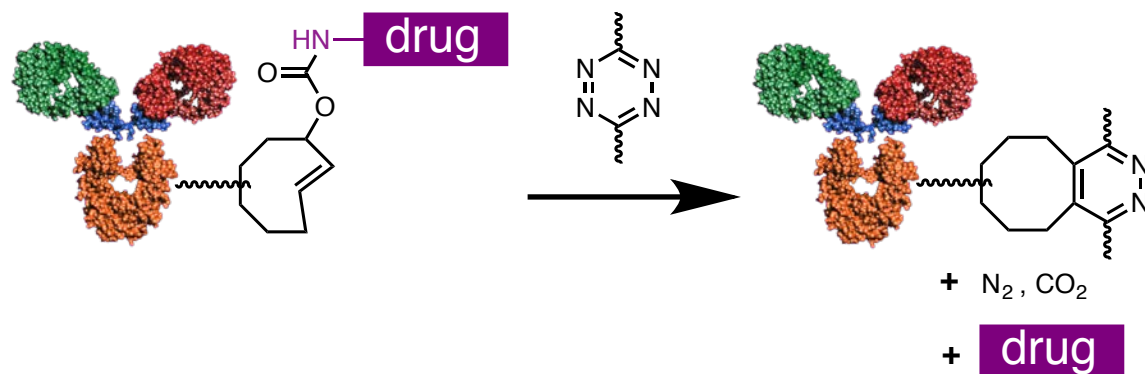
# “Click-to-release” ADC approach to cancer therapy

# Moving from Click to Click-to-Release

Click tags for Pretargeted Radioimmunoimaging & Radioimmunotherapy (RIT)

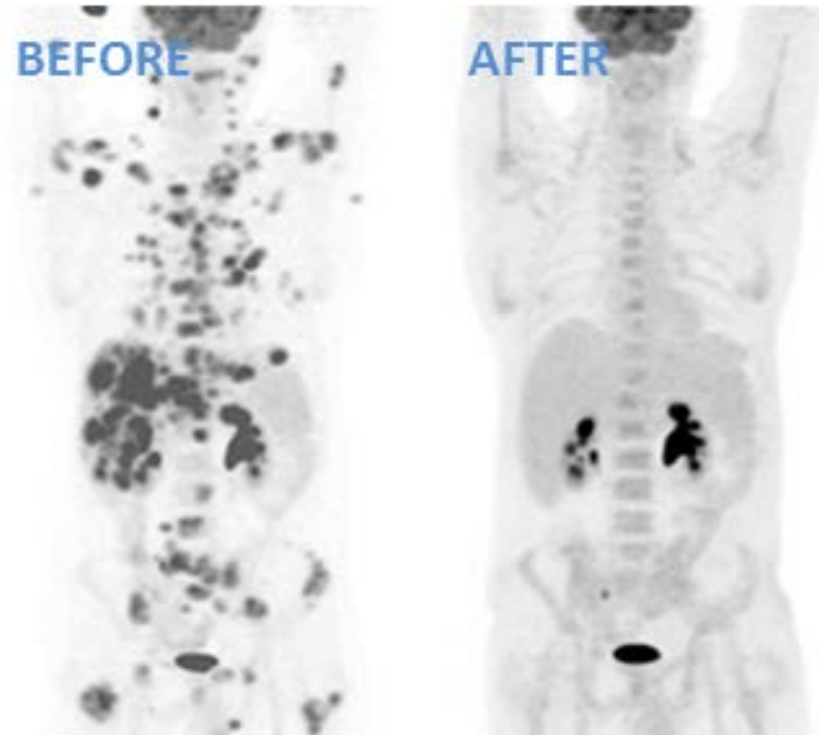
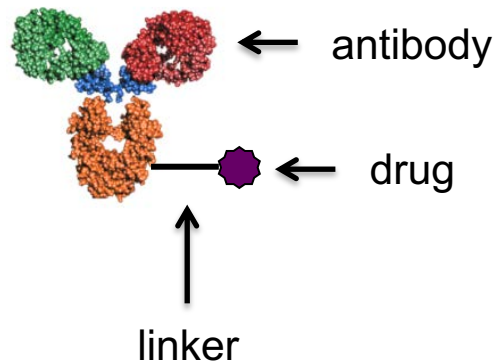


Click-cleavable linkers for non-internalizing Antibody-Drug Conjugates (ADC)



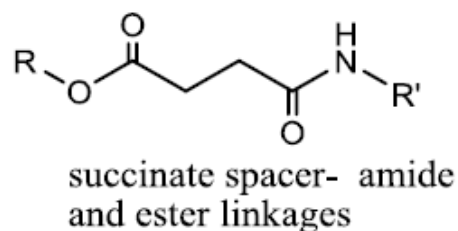
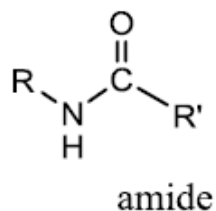
# Antibody-Drug Conjugates (ADCs)

Highly potent biopharmaceuticals that use the targeting ability of antibodies to selectively bind to tumor cells where the conjugated cancer drug is released

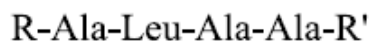


FDG PET scan before and 2 weeks after ADC treatment of patient with 70 Non-Hodgkin's Lymphoma tumors

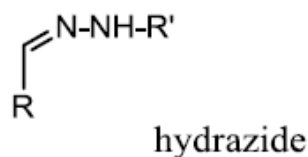
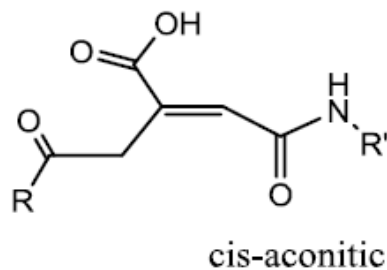
## Non cleavable linkers



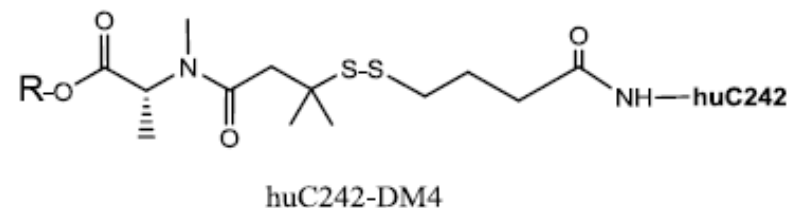
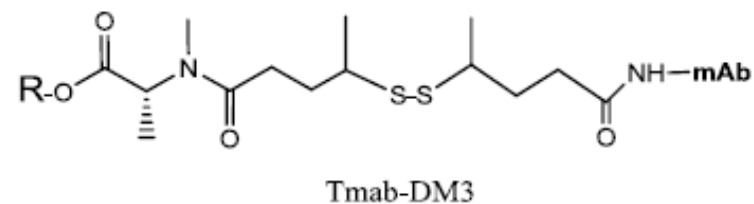
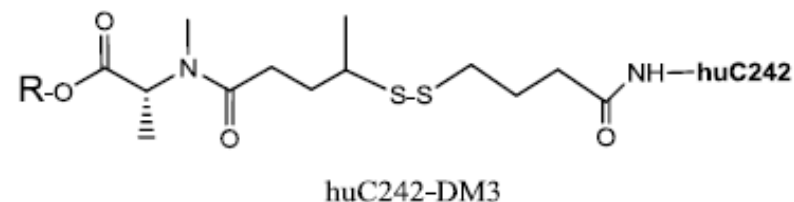
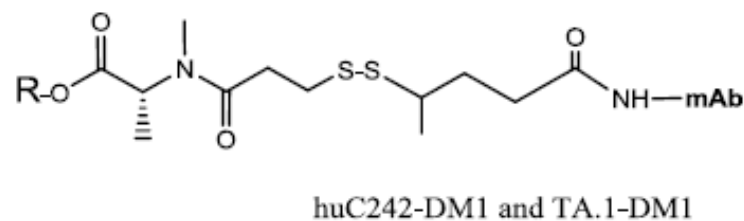
## Peptidase-labile linkers



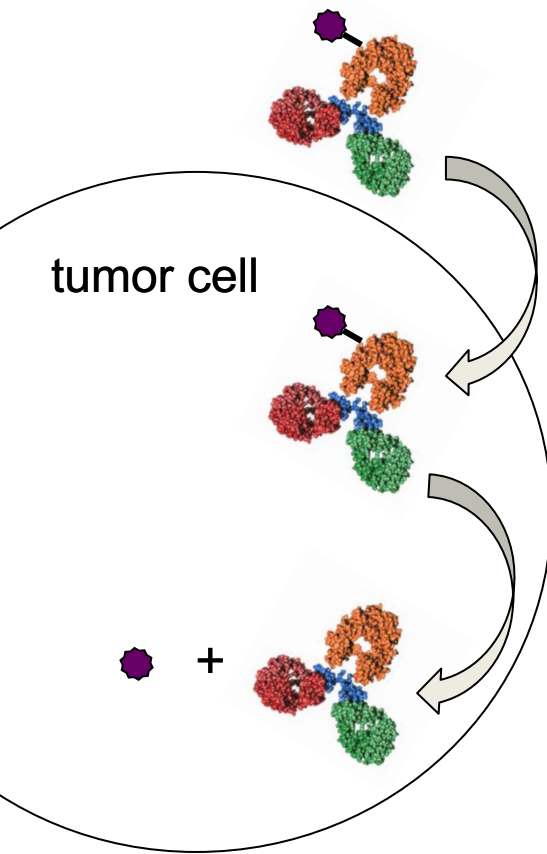
## Acid labile linkers



## Thiol labile linkers



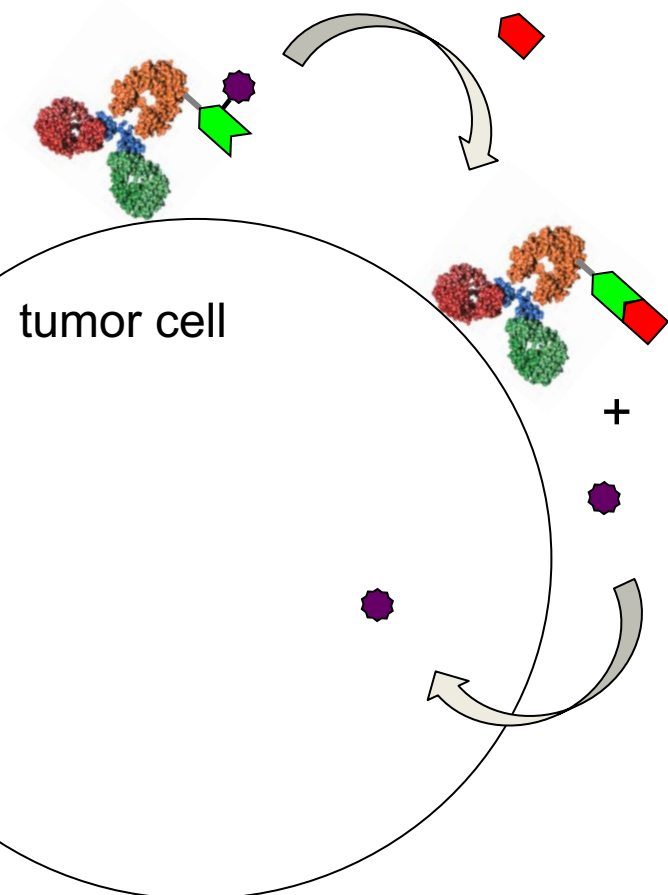
Current systems are based on **intracellular toxin release** by enzymes, thiols or pH



## Issues

- Limited to efficiently internalizing receptors
- Shortage of suitable ADC targets in solid tumors
- Less effective in heterogeneous or poorly penetrated tumors, i.e. solid tumors

# Click-to-Release ADCs



- Stable ADC linker, cleaved by a chemical probe in vivo
- 2 steps: after ADC has cleared from blood, probe is administered, triggering toxin release @ tumor
- Modification of in vivo validated pretargeting tech
- Expands the range of ADC targets: non-internalizing receptors, extracellular matrix constituents, stroma, etc
- Advantageous in heterogeneous or poorly penetrated tumors
- Universal & temporally controlled release independent from tumor biology
- Well suited for mAb fragments, or full mAbs in combination with a clearing agent

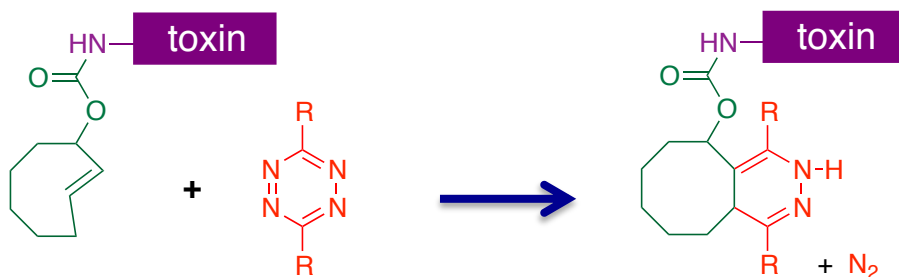
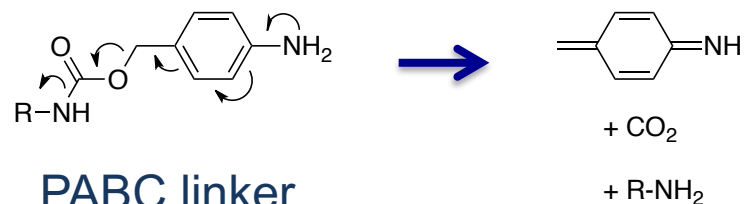
 linker  
 activator  
 toxin

# Moving from Click to Unclick

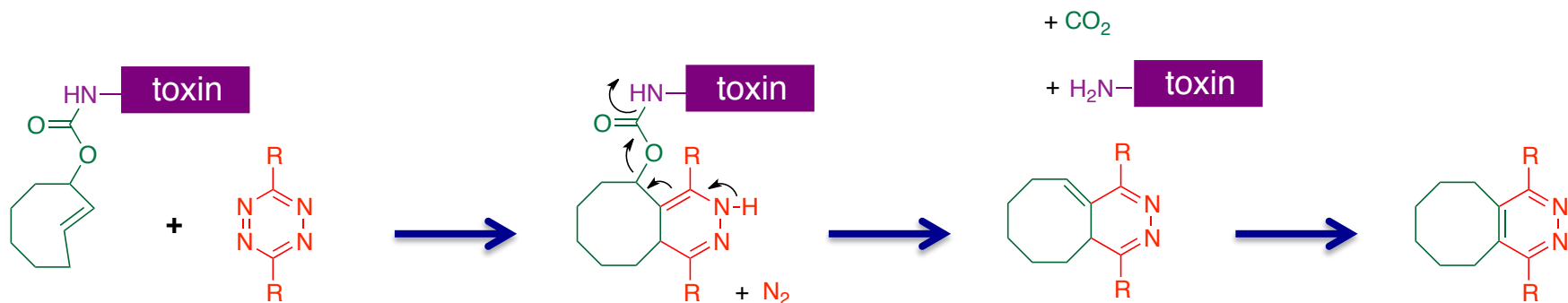
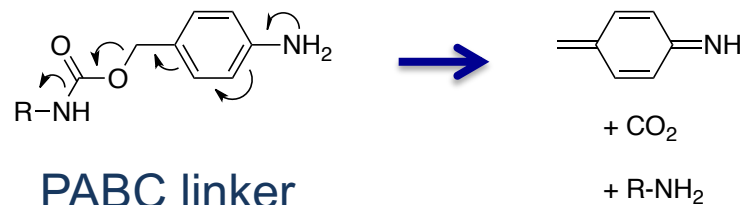




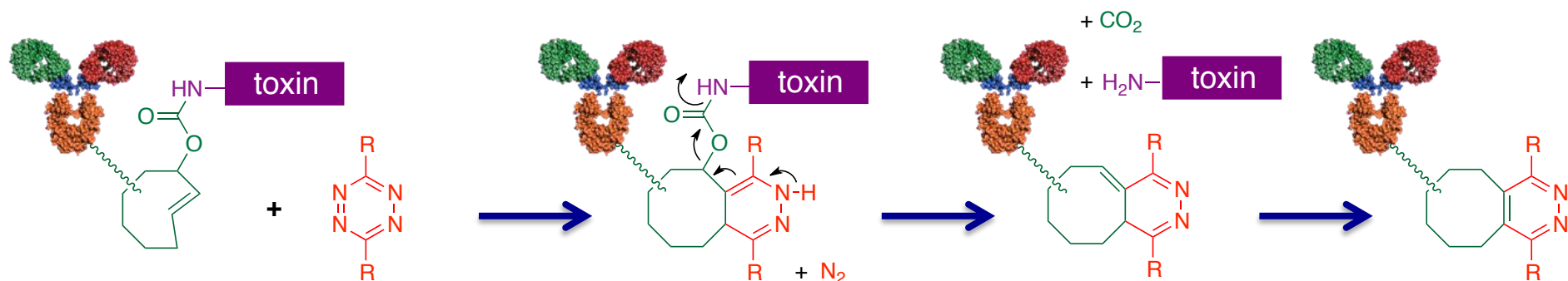
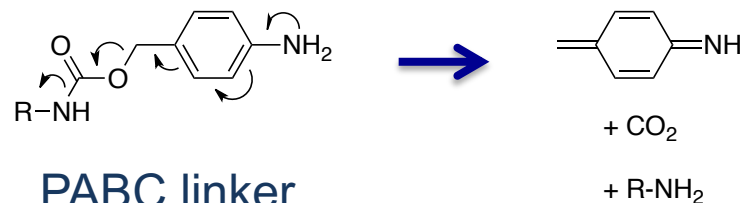
# Moving from Click to Unclick



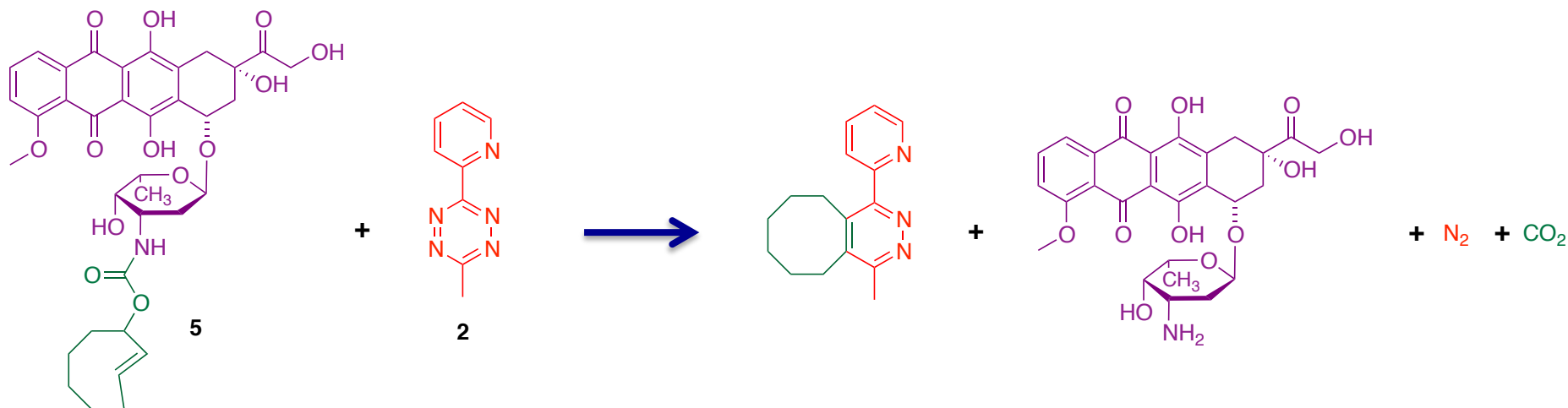
# Moving from Click to Unclick



# Moving from Click to Unclick



# Proof of concept: Dox release in vitro



IC<sub>50</sub> values determined in A431 cell line, from concentration range incubated 72 h @ 37°C

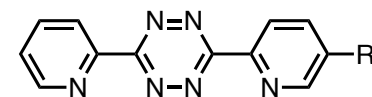
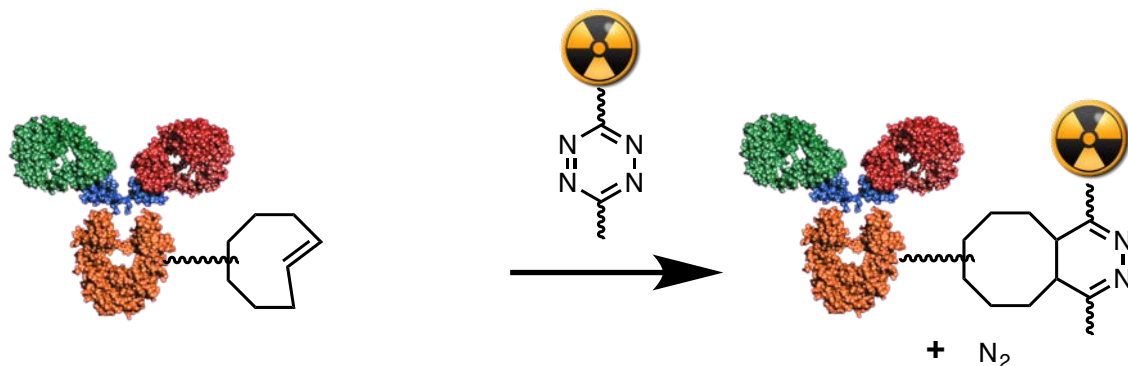
Compound	IC <sub>50</sub> (μM)
Doxorubicin	0.020 ± 0.002
Doxorubicin-tag	3.017 ± 0.486
Doxorubicin-tag + 1.5 eq. probe	0.137 ± 0.012
Probe	> 100

- Doxorubicin can be deactivated by conjugation to *trans*-cyclooctene tag and...
- .. activated again by equimolar reaction with tetrazine probe in cell culture

# Designing a suitable activator for in vivo application

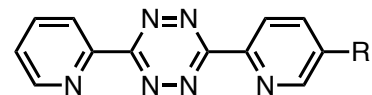
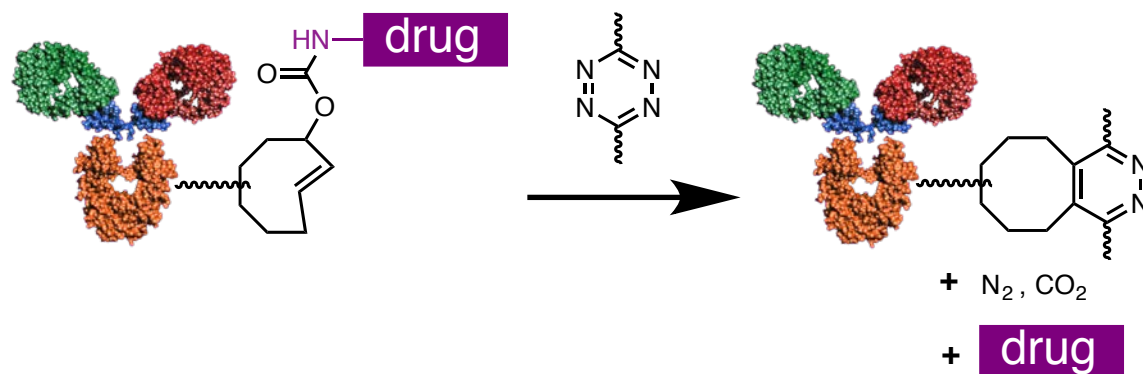
# Click-to-Release - lower reactivities

Click tags for Pretargeted Radioimmunoimaging & Radioimmunotherapy (RIT)

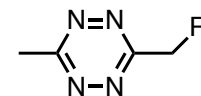


- $k_2 > 1.3 \times 10^5 \text{ M}^{-1}\text{s}^{-1}$

Click-cleavable linkers for non-internalizing Antibody-Drug Conjugates (ADC)



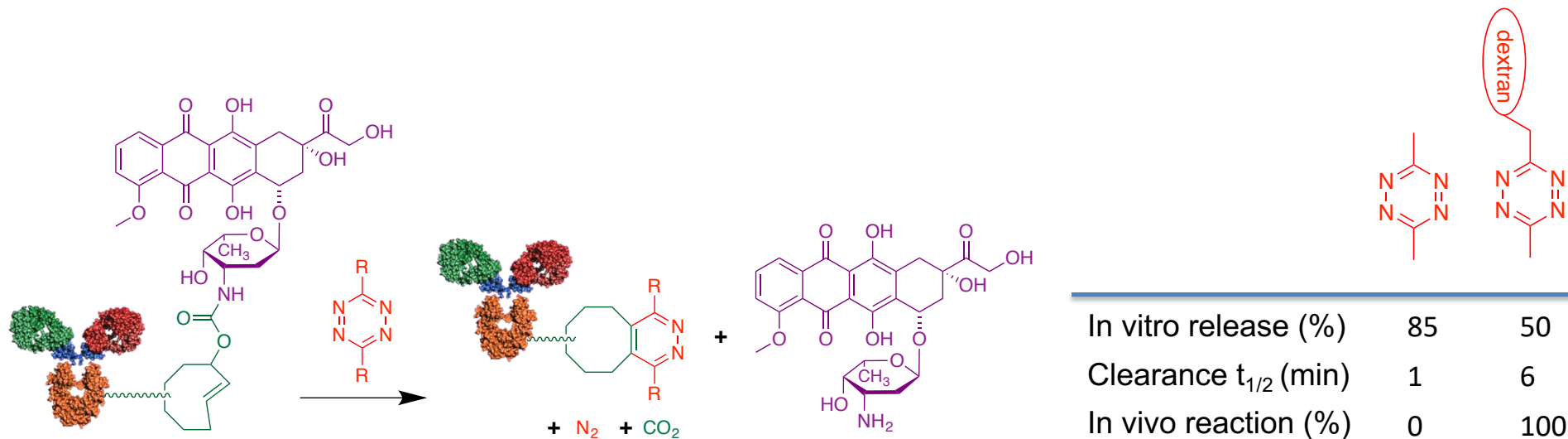
- $k_2 \sim 4 \times 10^3 \text{ M}^{-1}\text{s}^{-1}$
- release: 10 %



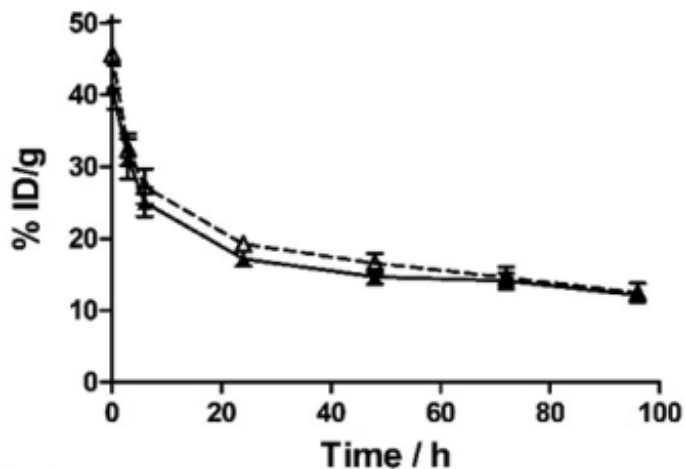
- $k_2 \sim 55 \text{ M}^{-1}\text{s}^{-1}$
- release: 85 %

**(we can use a large excess of activator!)**

# Proof of principle ADC: CC49-TCO-doxorubicin



## ADC blood clearance

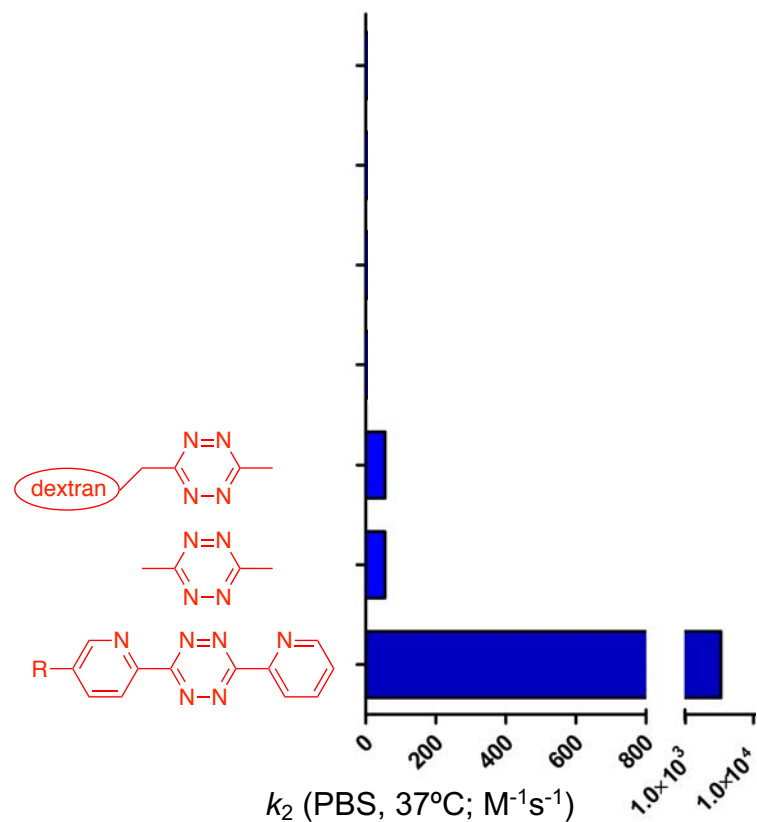


- High tumor uptake of CC49-TCO-dox, 100 % reaction with activator, good recovery of released dox, but..
- Slow mAb clearance requires use of clearing agent
- high dose of activator needed (30 mgs)
- Incomplete release

CC49 (dashed): t<sub>1/2</sub> 30.4 h  
CC49-TCO-Dox (solid): t<sub>1/2</sub> 26.9 h



# Click-to-Release: reactivity vs. release



release: 50 % / dose: 10,000 eq, 268 mg/kg

release: 90 % / dose: >100,000 eq

release: 10 % / dose: 50 eq

## *In vivo* proof-of-concept for Click-to-Release recently published in Nature Communications (2018)



### ARTICLE

DOI: [10.1038/s41467-018-03880-y](https://doi.org/10.1038/s41467-018-03880-y)

OPEN

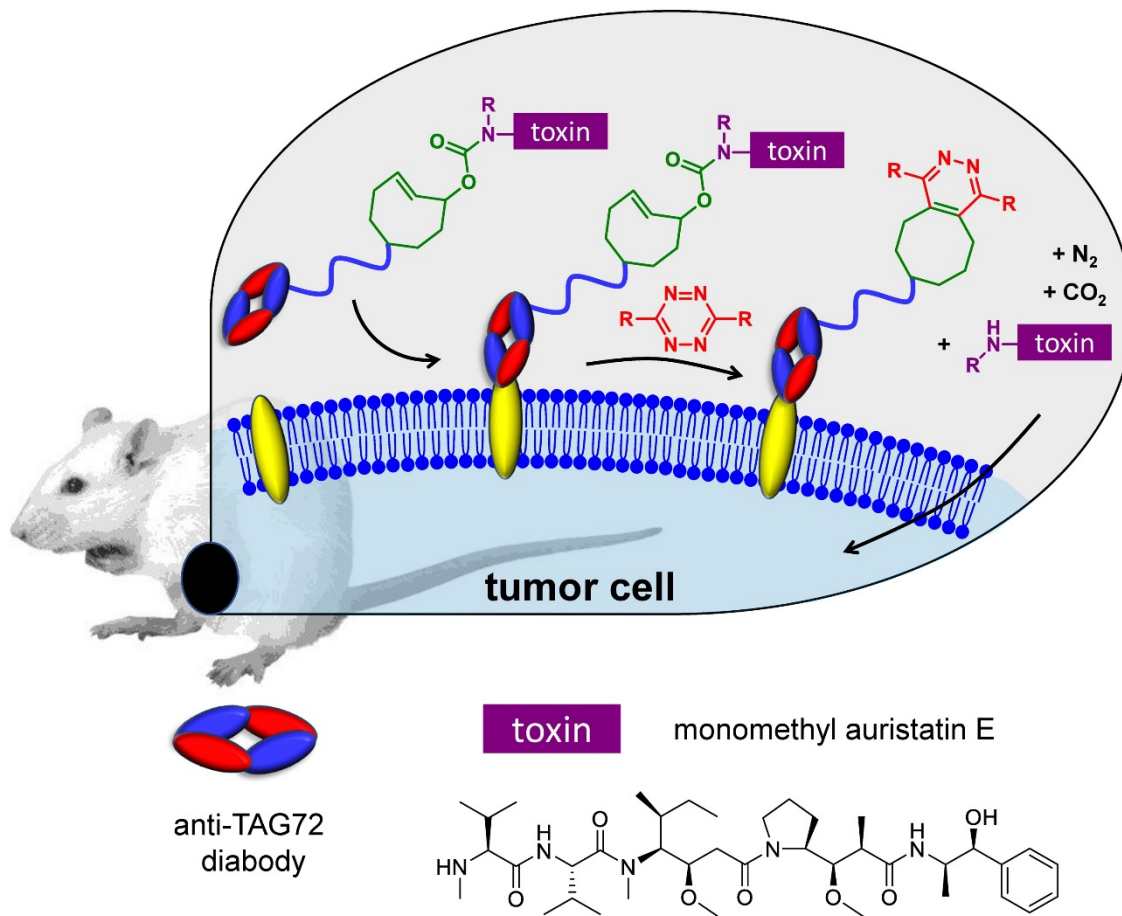
## Chemically triggered drug release from an antibody-drug conjugate leads to potent antitumour activity in mice

Raffaella Rossin<sup>1</sup>, Ron M. Versteegen<sup>2</sup>, Jeremy Wu<sup>3</sup>, Alisher Khasanov<sup>4</sup>, Hans J. Wessels<sup>5</sup>, Erik J. Steenbergen<sup>6</sup>, Wolter ten Hoeve<sup>7</sup>, Henk M. Janssen<sup>2</sup>, Arthur H.A.M. van Onzen<sup>1</sup>, Peter J. Hudson<sup>3</sup> & Marc S. Robillard<sup>1</sup>

Current antibody-drug conjugates (ADCs) target internalising receptors on cancer cells leading to intracellular drug release. Typically, only a subset of patients with solid tumours has sufficient expression of such a receptor, while there are suitable non-internalising receptors and stroma targets. Here, we demonstrate potent therapy in murine tumour models using a non-internalising ADC that releases its drugs upon a click reaction with a chemical activator, which is administered in a second step. This was enabled by the development of a diabody-based ADC with a high tumour uptake and very low retention in healthy tissues, allowing systemic administration of the activator 2 days later, leading to efficient and selective activation throughout the tumour. In contrast, the analogous ADC comprising the protease-cleavable linker used in the FDA approved ADC Adcetris is not effective in these tumour models. This first-in-class ADC holds promise for a broader applicability of ADCs across patient populations.

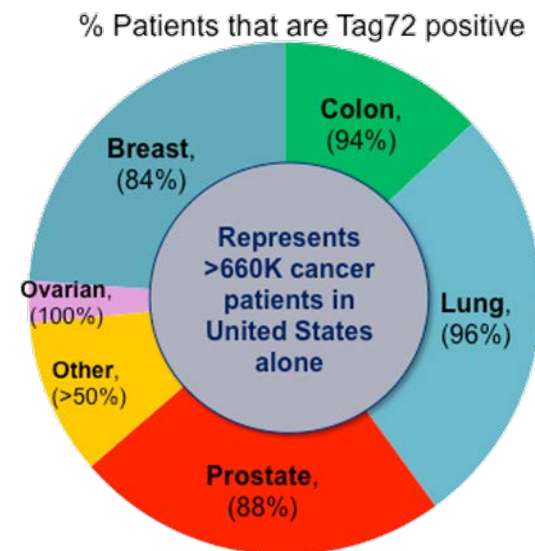
# Click-to-Release of MMAE from TAG72-targeted diabody

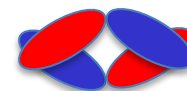
Objectives: faster clearing ADC, improved activator, more potent drug



TAG72:

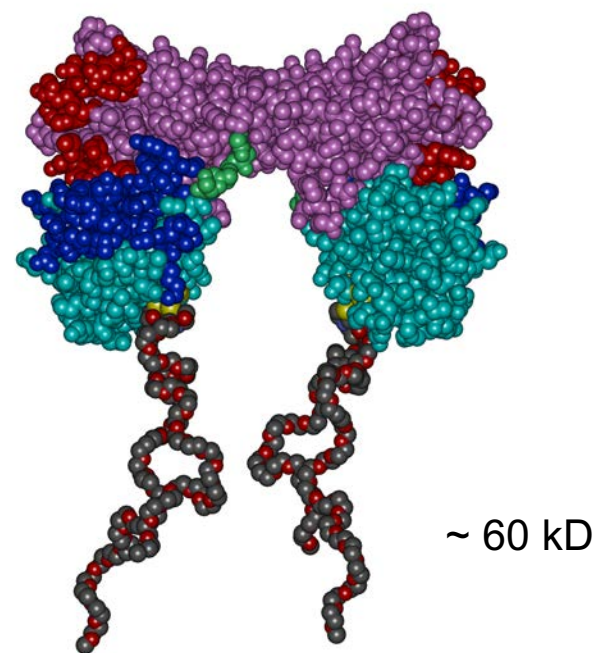
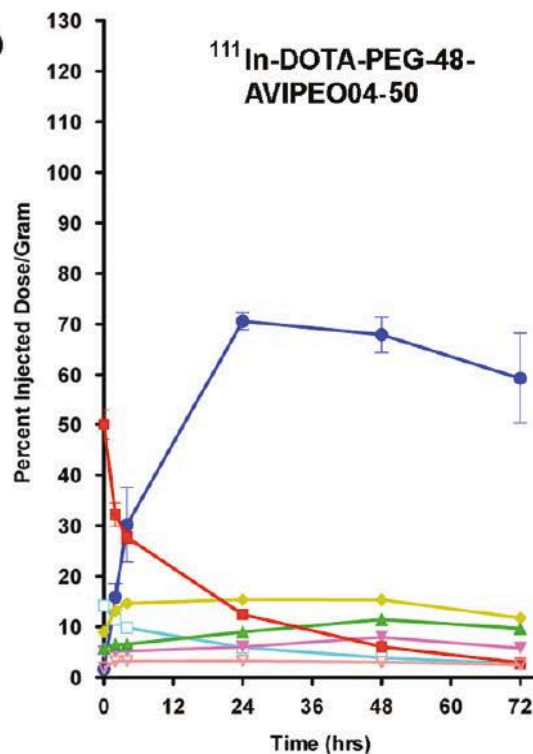
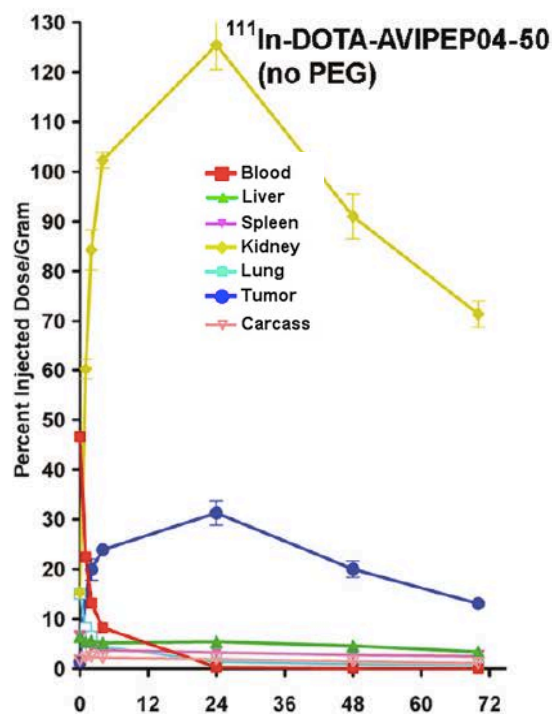
extracellular cell membrane  
target, non-internalizing, low  
shedding





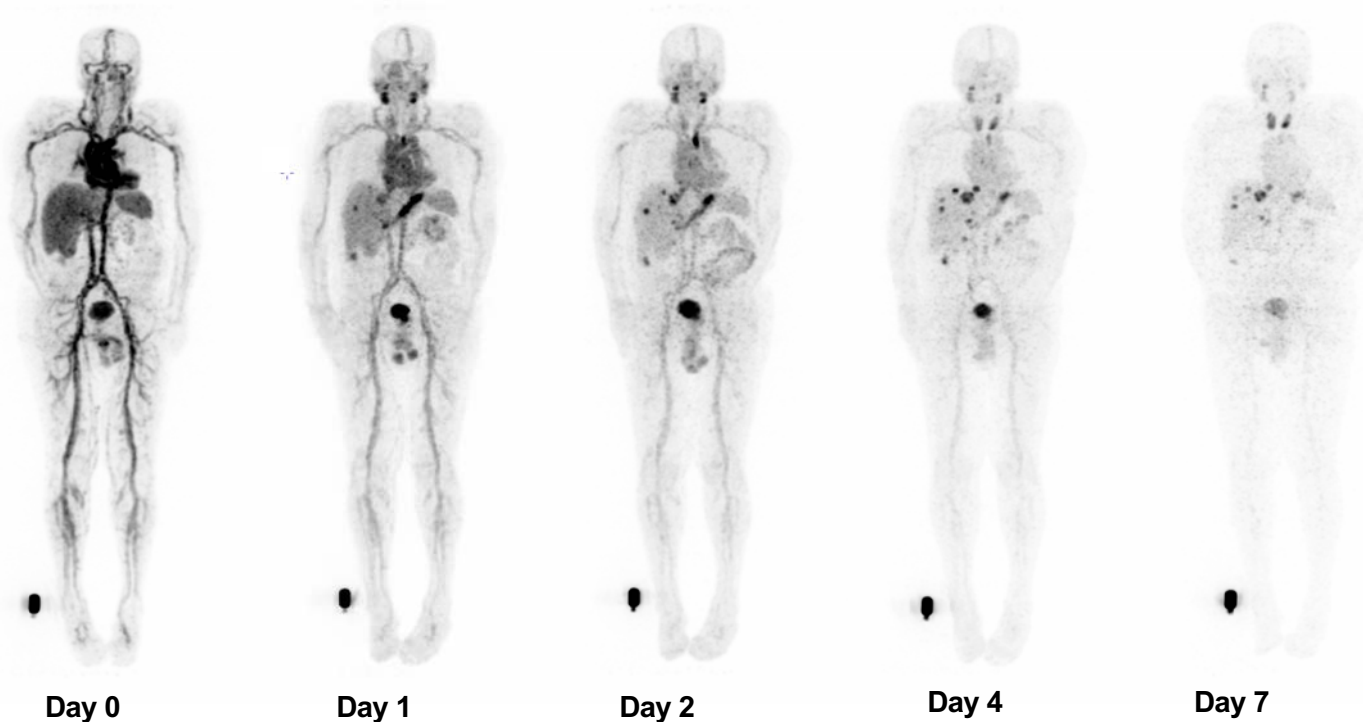
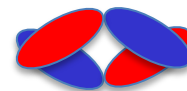
no PEG

4 x PEG<sub>48</sub>



LS174T-tumor-bearing mice injected with <sup>111</sup>In-DOTA-AVP04-50 with / without 4 x PEG<sub>48</sub> conjugated via cysteines

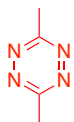
# Phase 1 imaging study with $^{124}\text{I}$ -PEG-AVP0458 prostate and colorectal cancer patients



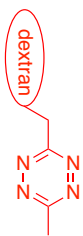
- $T_{\alpha}$  5hr from blood and  $T_{\beta}$  from whole body 45hr
- High tumor:blood ratios - 22:1 at 7 days p.i.
- Reproducible pharmacokinetics across two dose levels (1 and 10 mg/m<sup>2</sup>)

# Slowing down the tetrazine clearance with a chelator

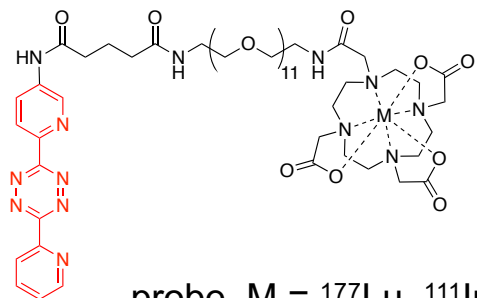
1<sup>st</sup> generation  
activators



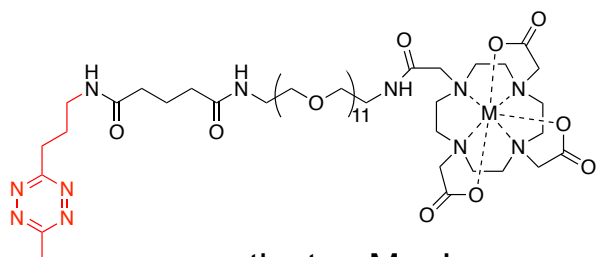
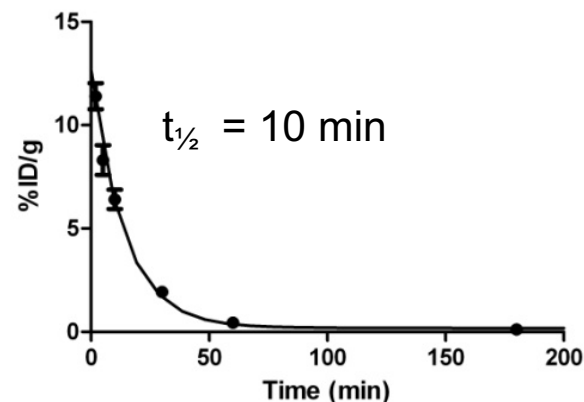
$t_{1/2}$  1 min



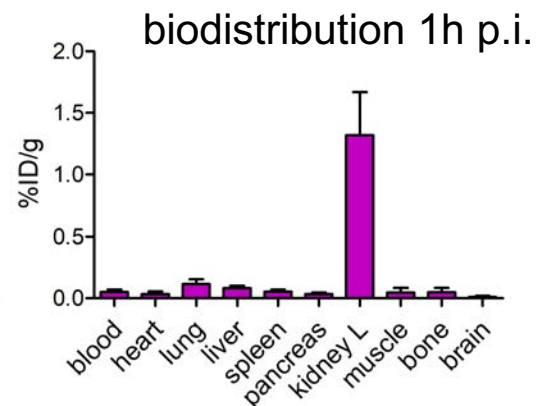
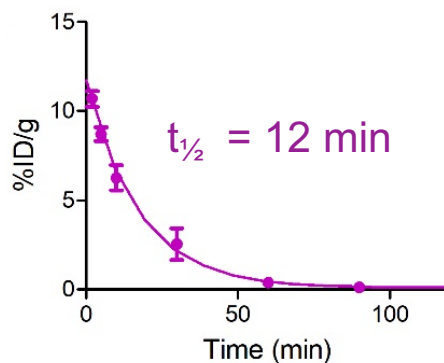
$t_{1/2}$  6 min



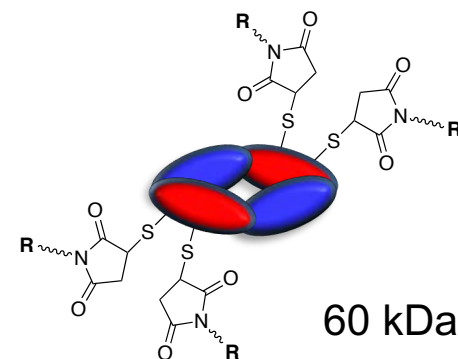
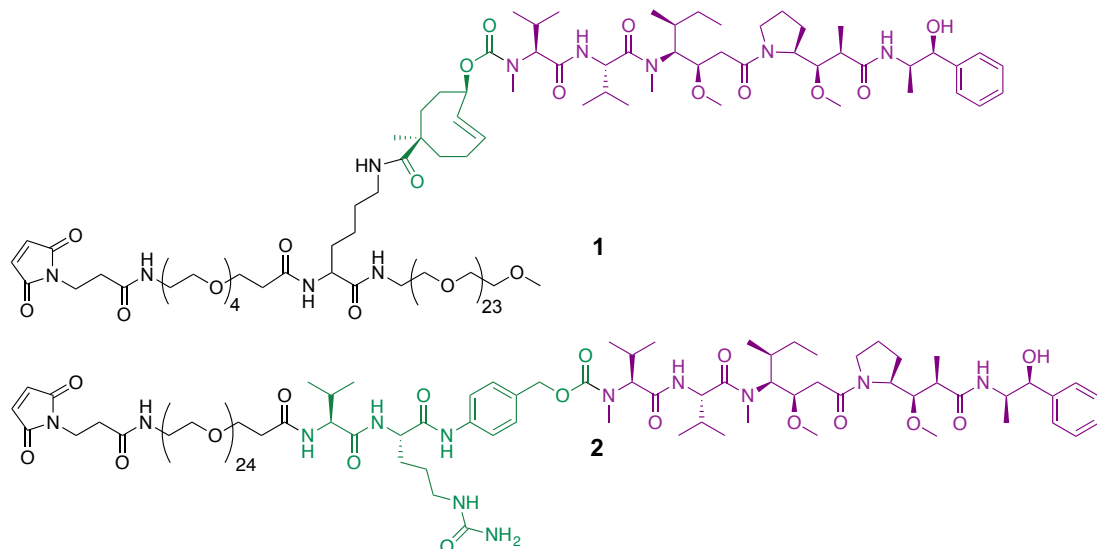
probe, M =  $^{177}\text{Lu}$ ,  $^{111}\text{In}$



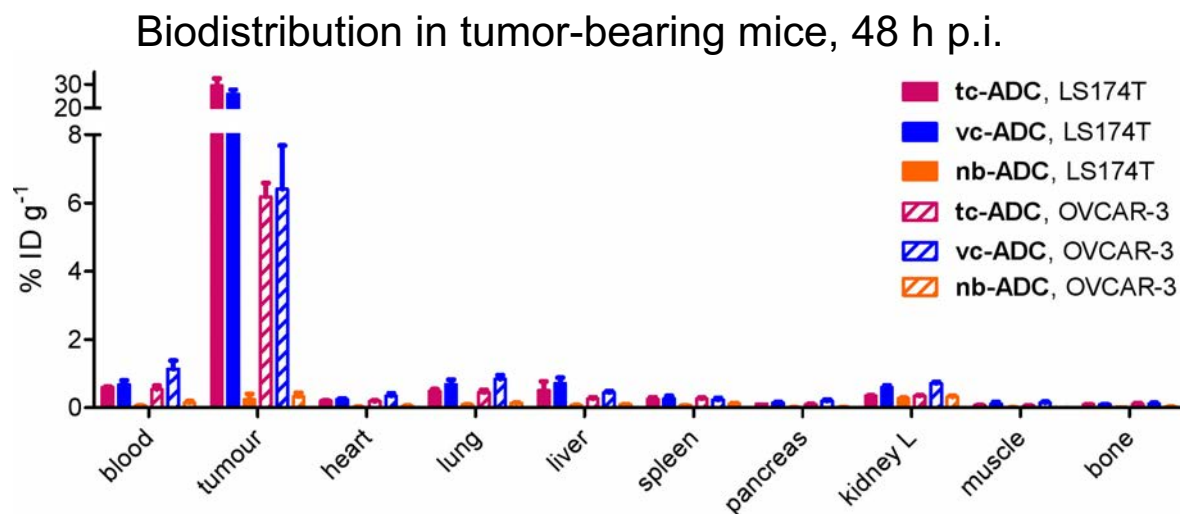
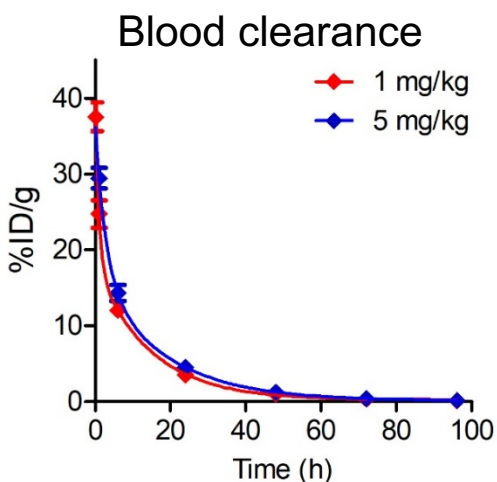
new activator, M = Lu



# Anti-TAG72 diabody ADC and controls



ADC	diabody	R
<b>tc-ADC</b>	anti-TAG72	-TCO-MMAE (1)
<b>vc-ADC</b>	anti-TAG72	-val-cit-MMAE (2)
<b>nb-ADC</b>	anti-PSMA	-TCO-MMAE (1)

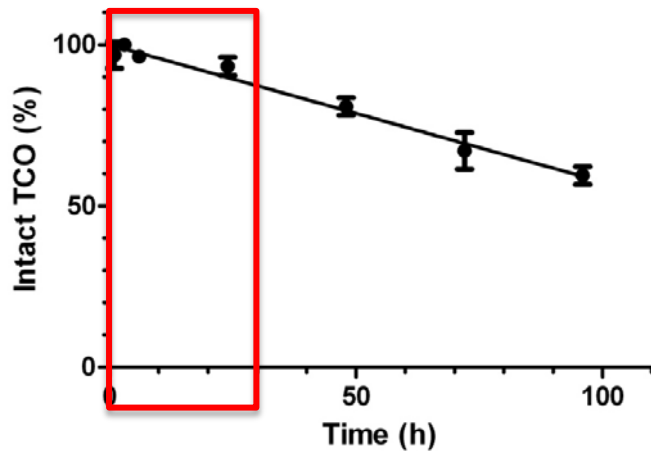


Fast clearance allows 2 day interval between diabody and activator

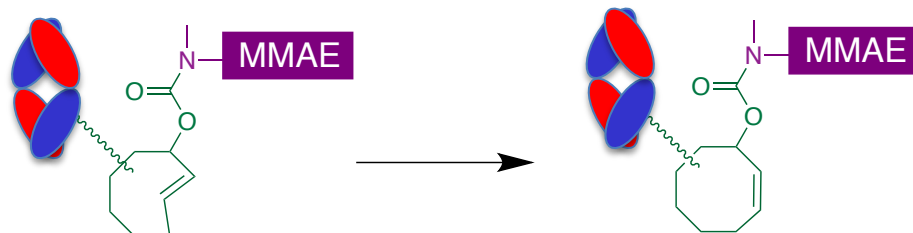


# Slow linker deactivation of TCO ADC in circulation

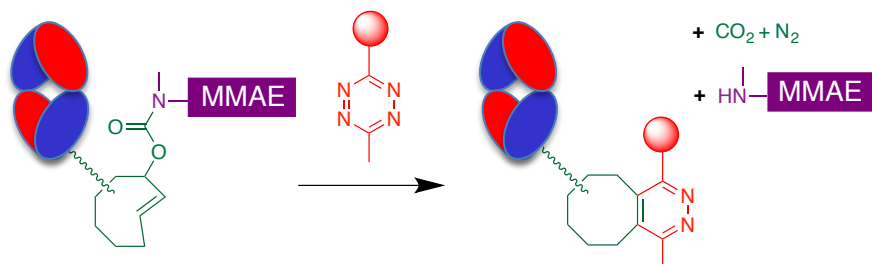
In vivo TCO stability in blood in mice (n=3) assessed ex vivo with  $^{177}\text{Lu}$ -DOTA-tetrazine, corrected for  $^{125}\text{I}$ -ADC blood clearance



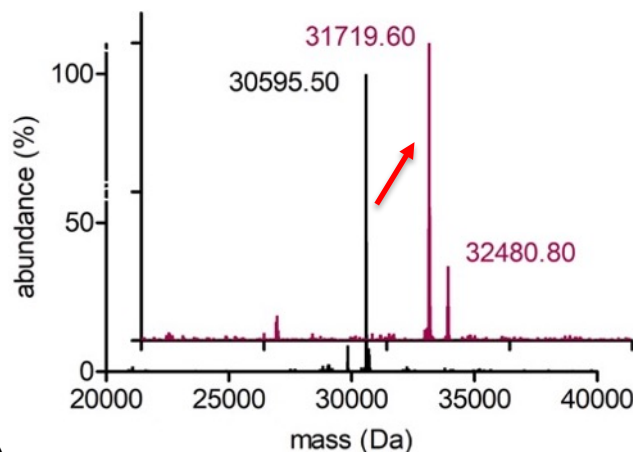
Stability  $t_{1/2} = 5$  days



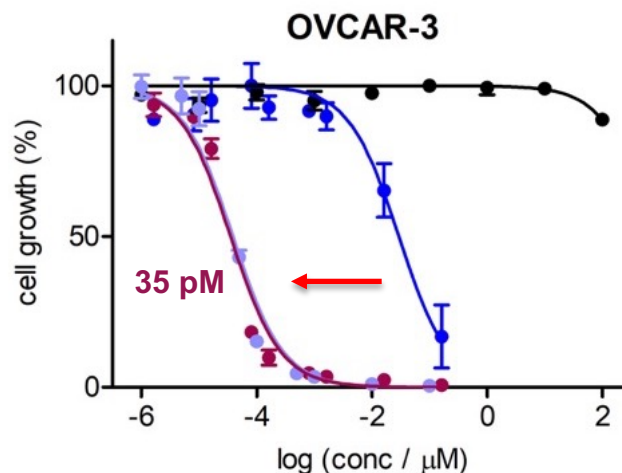
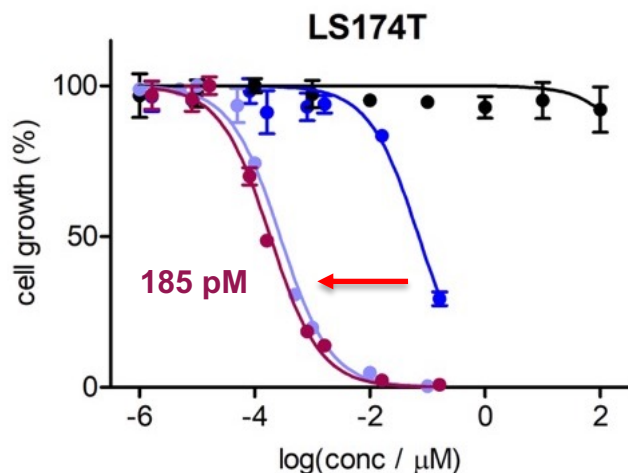
# Reaction of ADC with activator leads to instantaneous drug release and 1000-fold increased cytotoxicity



90 % release after 1h in PBS

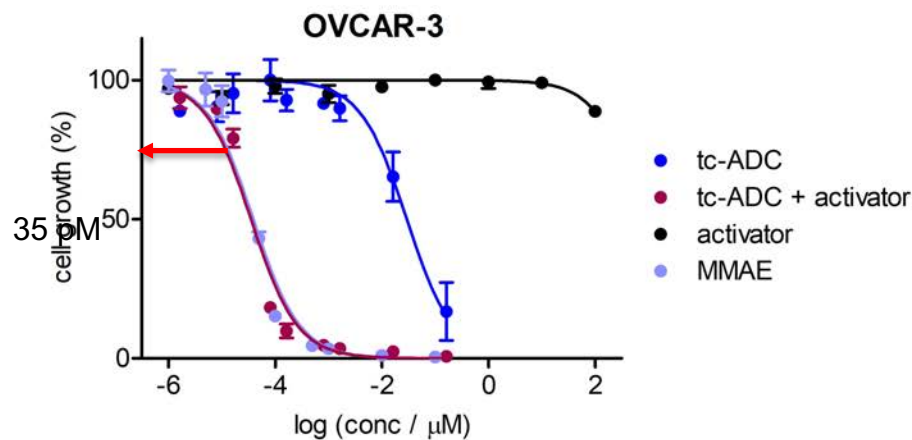
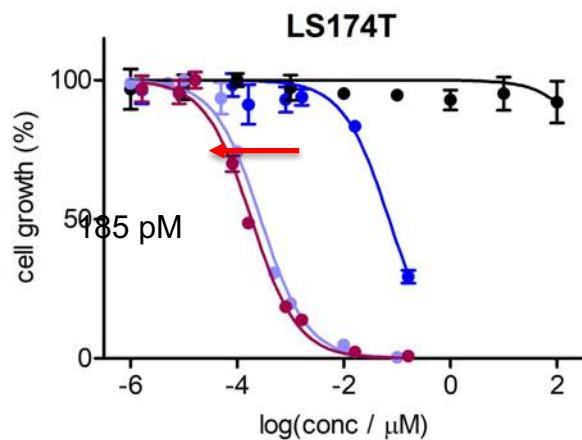
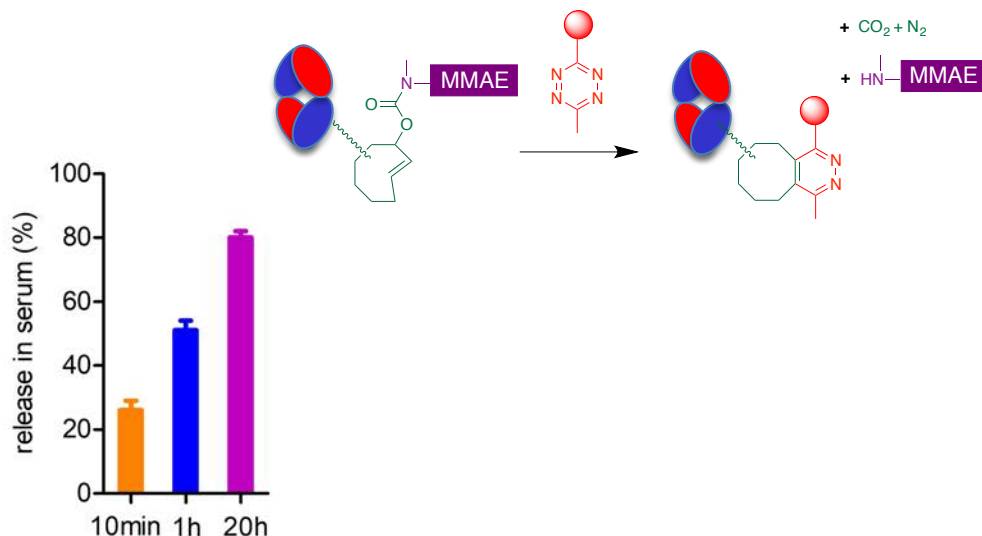
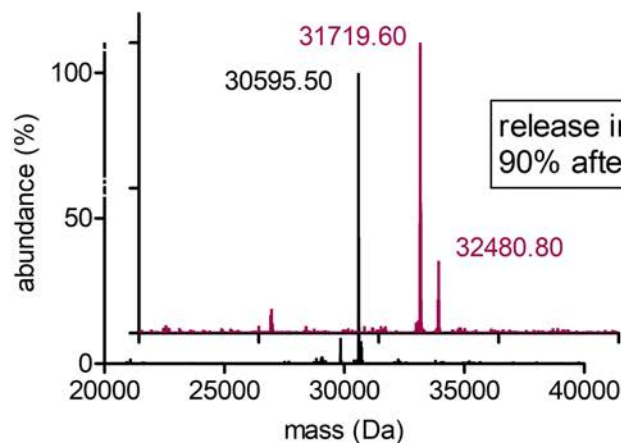


Cytotoxicity in colorectal (LS174T) and ovarian (OVCAR-3) cancer cell culture

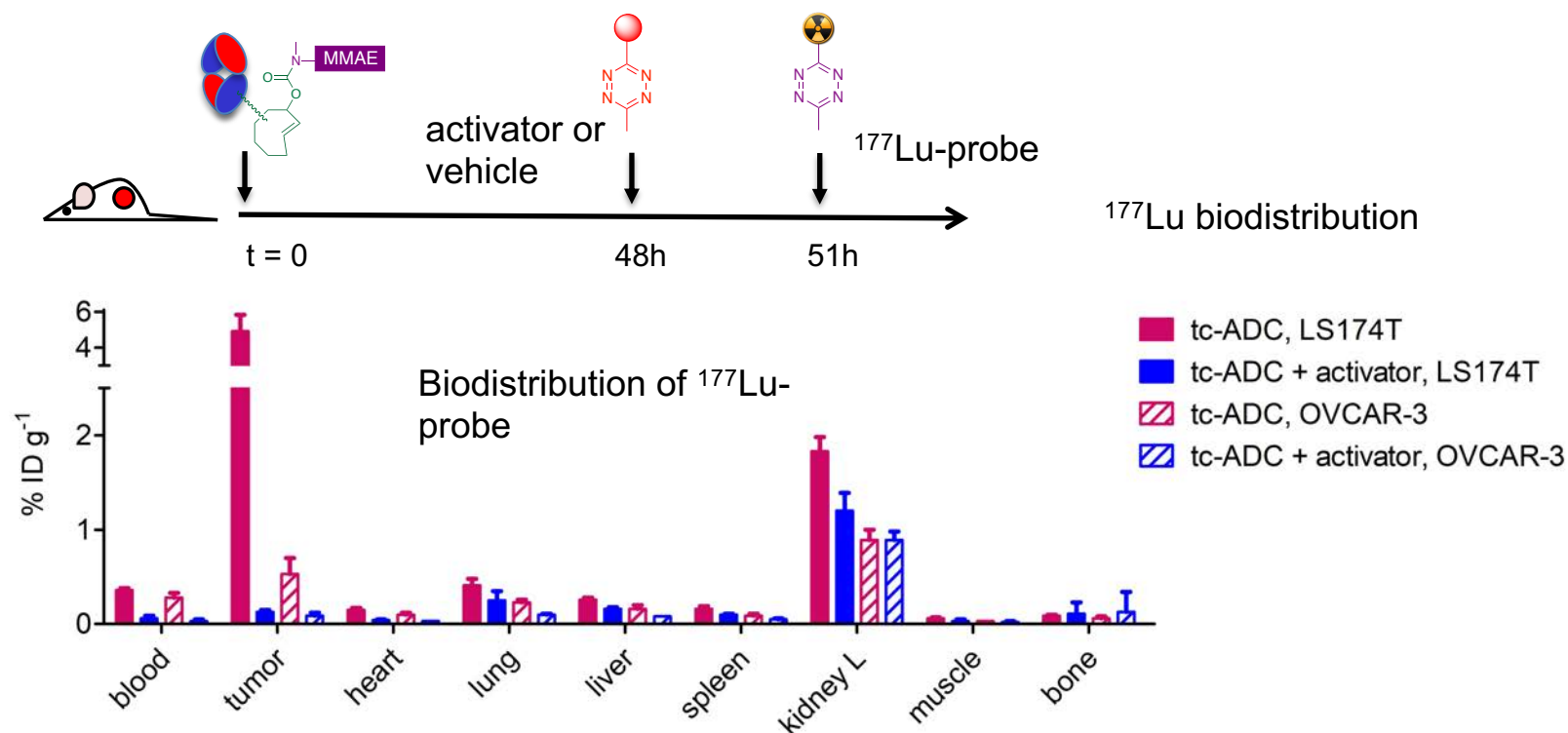


- tc-ADC
- tc-ADC + activator
- activator
- MMAE

# ADC activation: rapid drug release & 1000-fold higher cytotoxicity

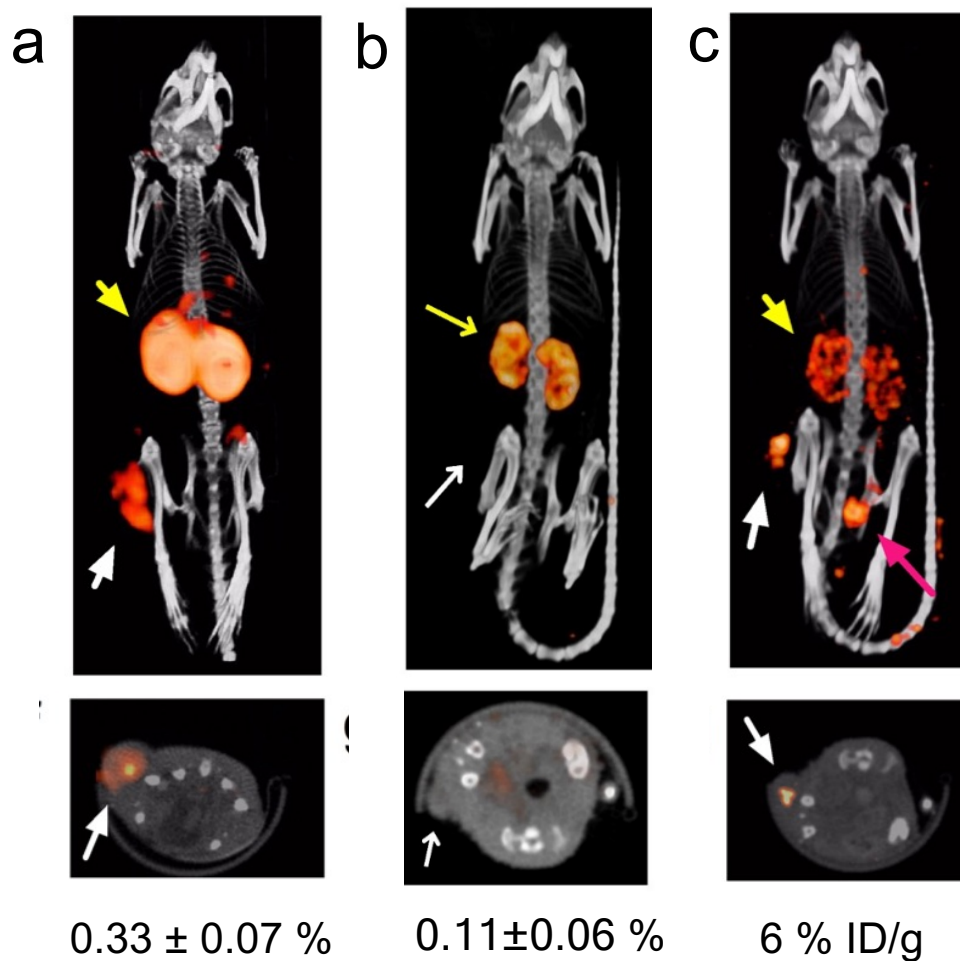
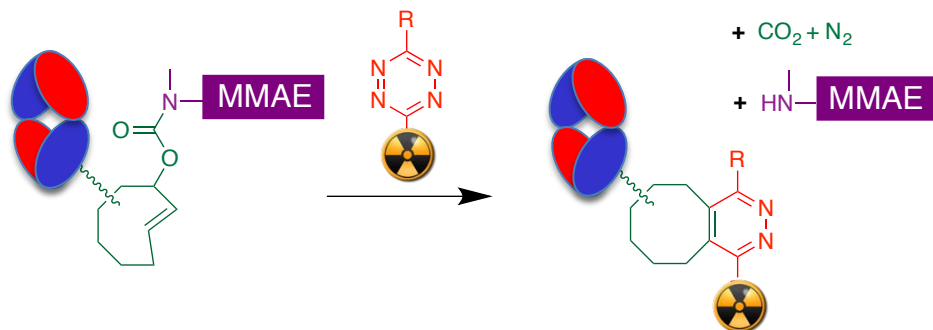


# Complete reaction of tumor-bound ADC with excess tetrazine

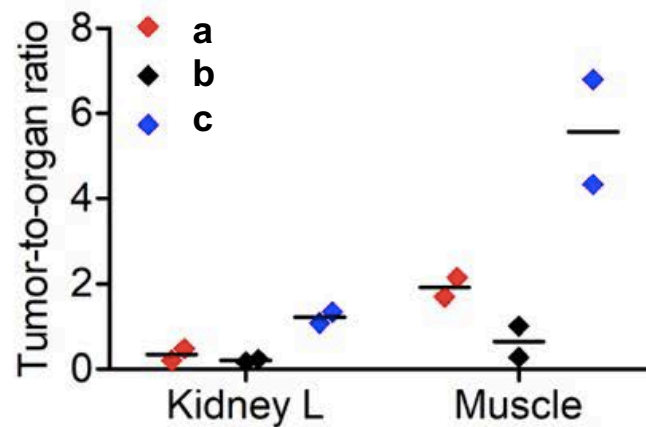


tumor-bearing mice (n=4) injected with 1) tc-ADC (0.033  $\mu\text{mol/kg}$ ) , 2) vehicle or activator (0.335 mmol/kg; 48h), 3)  $^{177}\text{Lu}$ -probe (0.335  $\mu\text{mol/kg}$ ; 51 h), biodistribution @ 54 h.

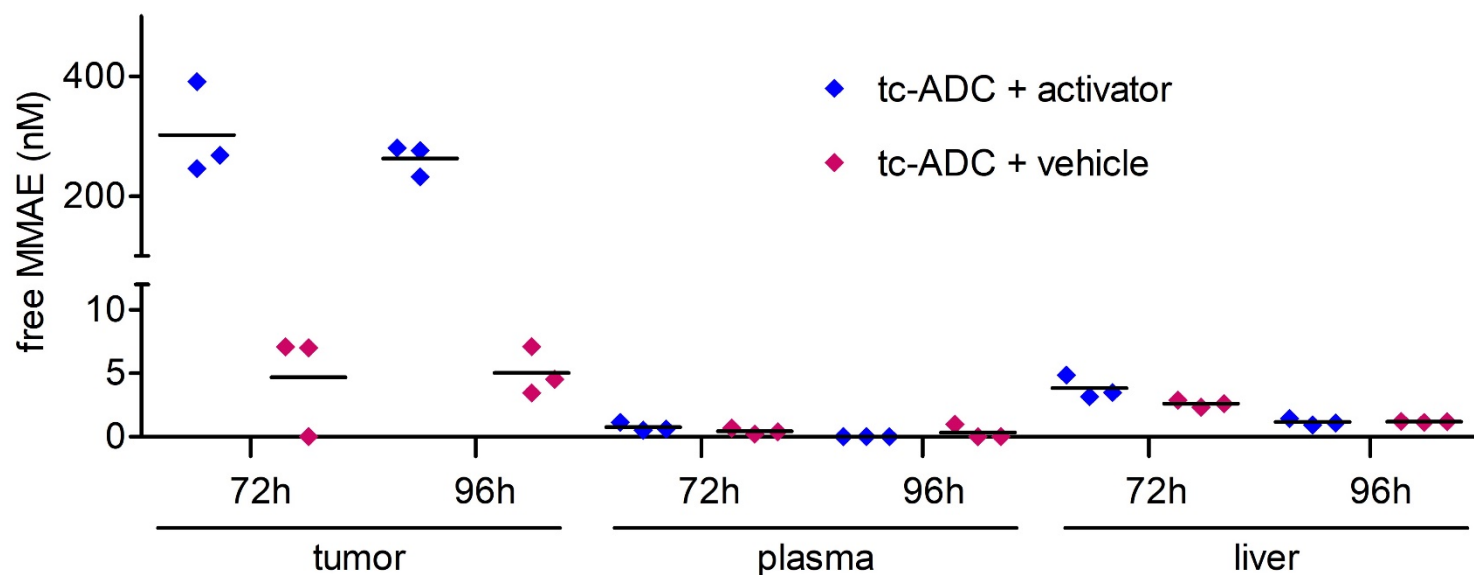
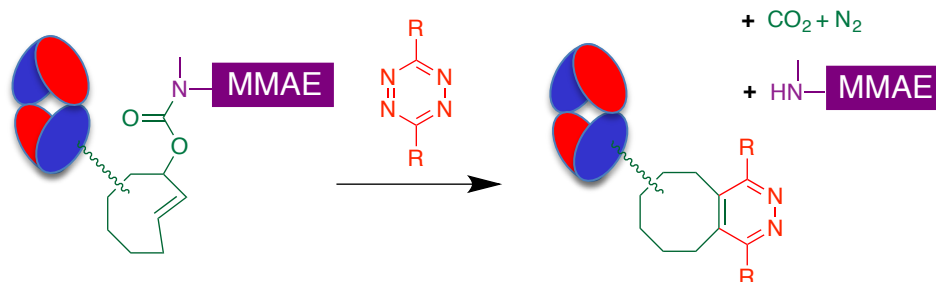
LS174T-mice inj. 1) tc-ADC; 2) 1 eq  $^{111}\text{In}$ -Tz @ 48h; 3) imaging/biodistribution @ 51h



- a) tc-ADC +  $^{111}\text{In}$ -activator
- b)  $^{111}\text{In}$ -activator
- c) tc-ADC +  $^{111}\text{In}$ -probe

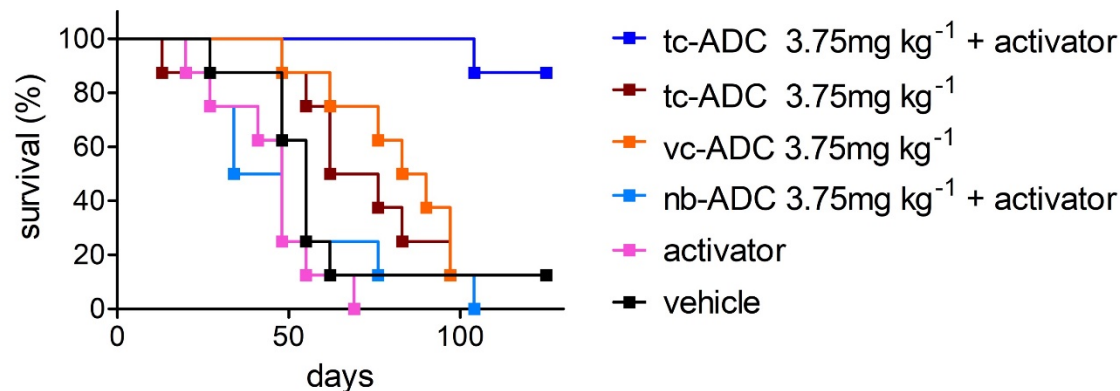
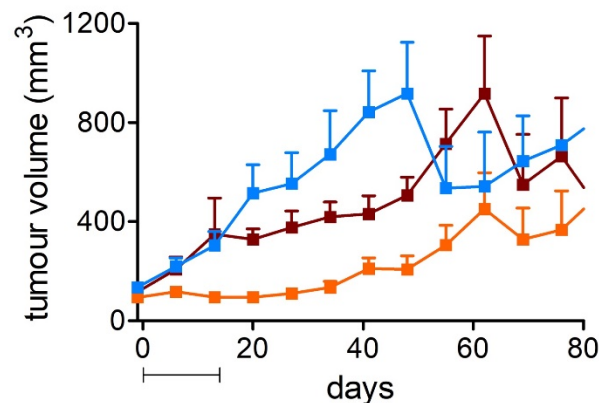
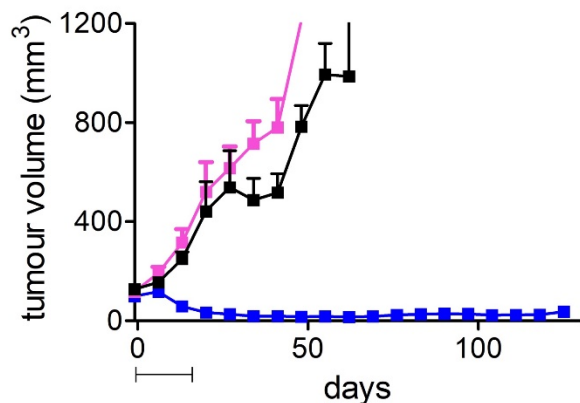


# Free MMAE concentrations in vivo



LS174T-mice inj. 1) tc-ADC (0.033  $\mu\text{mol/kg}$ ) , 2) activator (0.335 mmol/kg; 48h), biodistribution @ 72 and 96 h, MMAE extraction

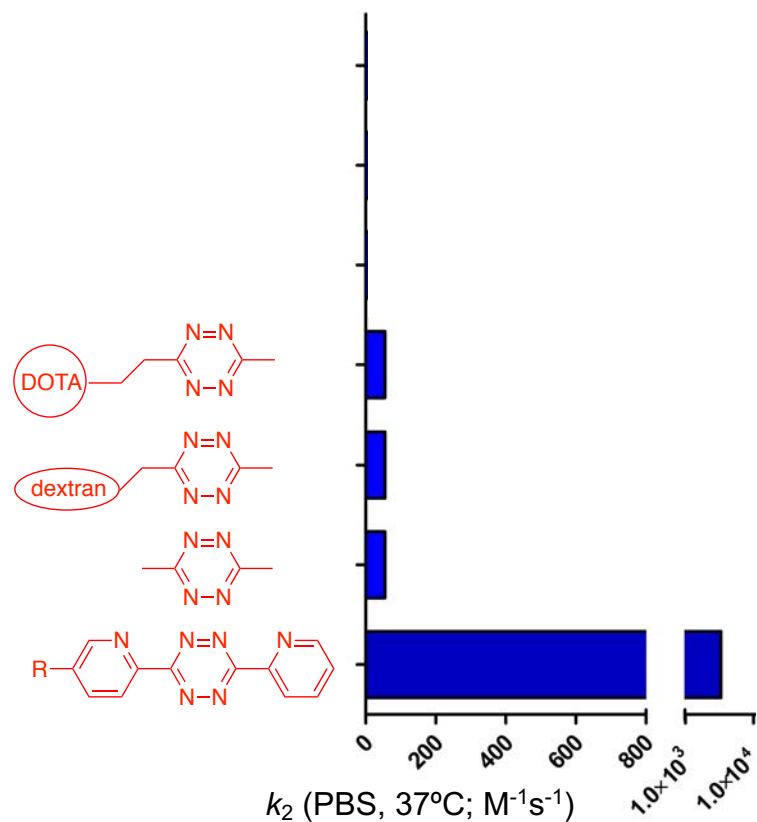
# Therapeutic efficacy in OVCAR-3 tumor bearing mice



ADC	diabody	R
<b>tc-ADC</b>	anti-TAG72	-TCO-MMAE (1)
<b>vc-ADC</b>	anti-TAG72	-val-cit-MMAE (2)
<b>nb-ADC</b>	anti-PSMA	-TCO-MMAE (1)

Mice inj. within 2 weeks with 4 cycles of 1) tc-ADC, 2) activator (0.335 mmol/kg; 48h)

# Click-to-Release: reactivity vs. release



release: 90 % / dose: 10,000 eq, 36 mg/kg

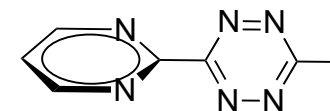
release: 50 % / dose: 10,000 eq, 268 mg/kg

release: 90 % / dose: >100,000 eq

release: 10 % / dose: 50 eq

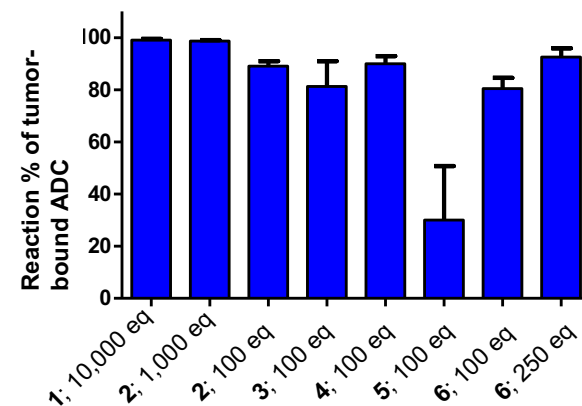
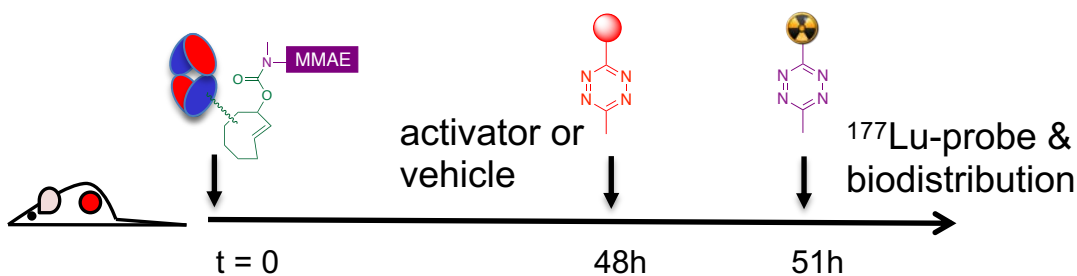
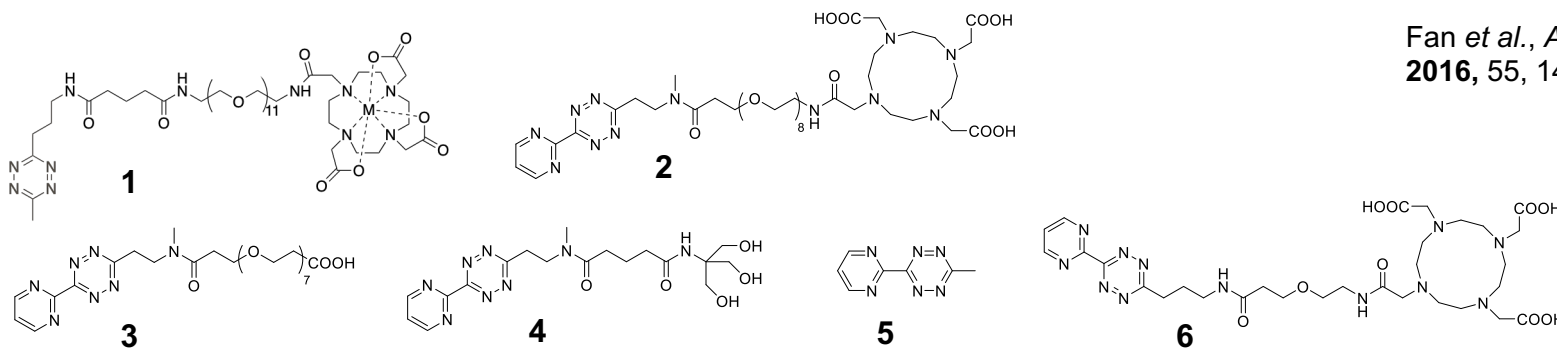


# Pyrimidine tetrazines: boosted reactivity, slightly lower release

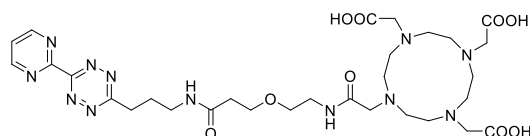


- $k_2 \sim 1000 \text{ M}^{-1}\text{s}^{-1}$
- release: 65-75 %

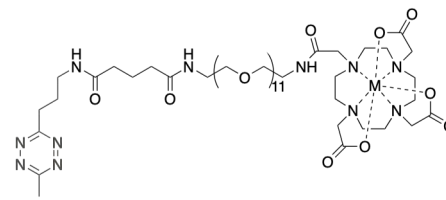
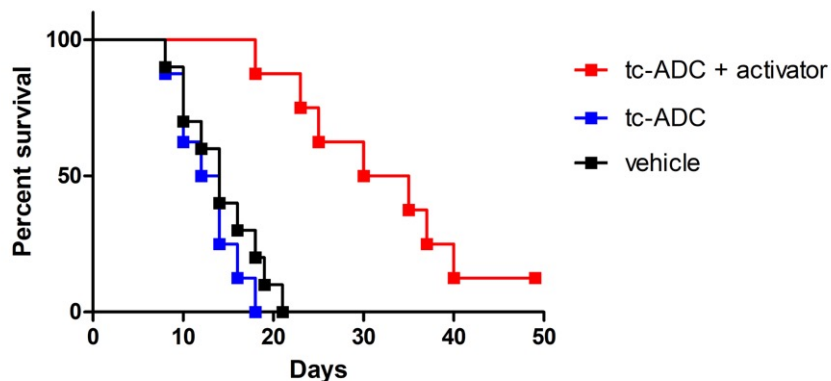
Fan et al., *Angew. Chem. Int. Ed.*  
**2016**, 55, 14046



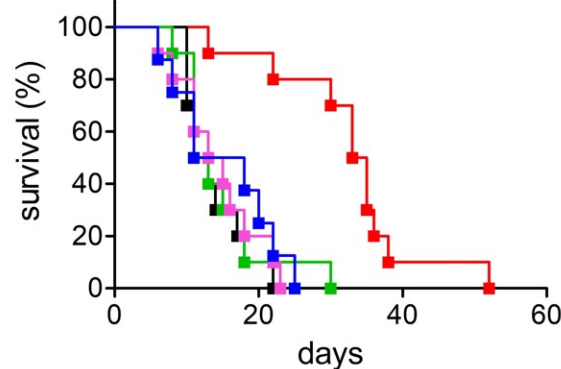
# Same therapeutic efficacy of pyrimidine tetrazine at 20-fold lower dose



500 eq



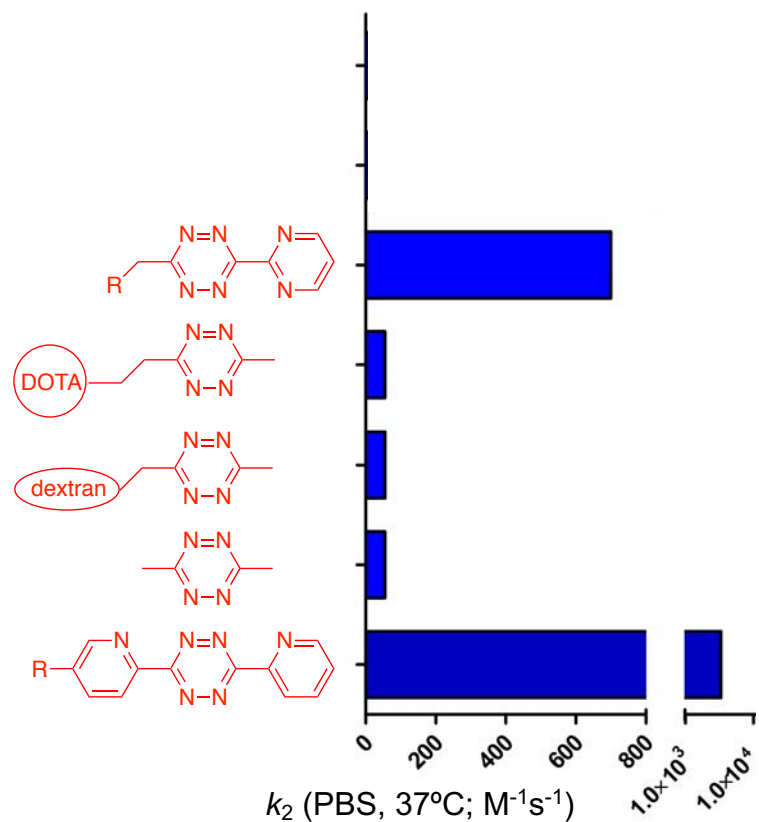
10,000 eq



Rossin *et al.*, *Nature Commun.* **2018**, 9, 1484

LS174T tumored mice inj. i.v. within 2 weeks with 4 cycles of 1) tc-ADC (3 mg/Kg), 2) activator (17  $\mu$ mol/kg; 48h)

# Click-to-Release: reactivity vs. release



release: 70 % / dose: 500 eq, 0.6 mg/kg

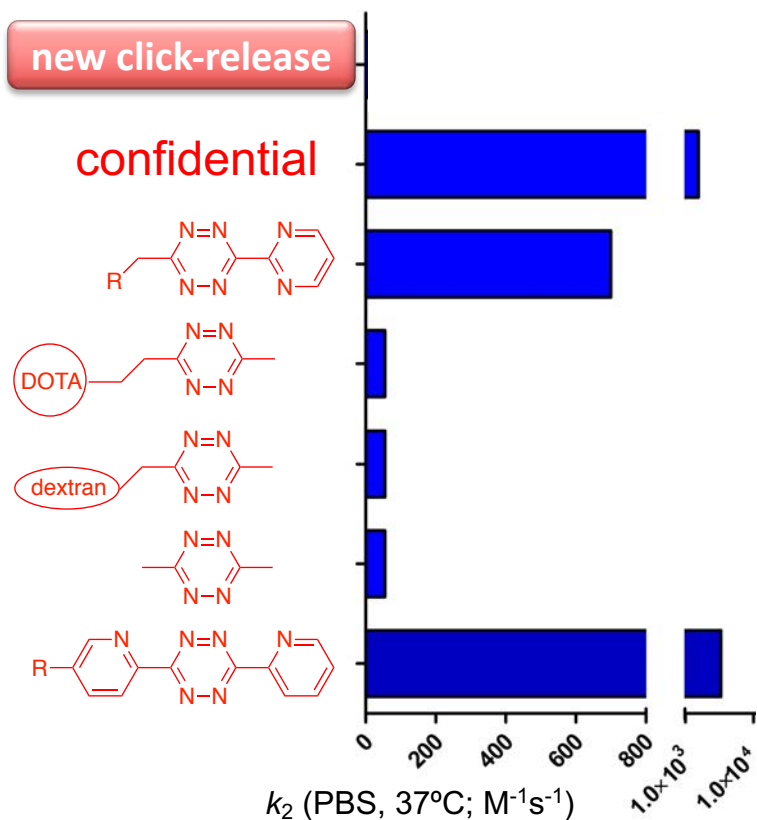
release: 90 % / dose: 10,000 eq, 36 mg/kg

release: 50 % / dose: 10,000 eq, 268 mg/kg

release: 90 % / dose: >100,000 eq

release: 10 % / dose: 50 eq

# Click-to-Release: reactivity vs. release



release: 100 % / dose: 250 eq, 0.3 mg/kg

release: 70 % / dose: 500 eq, 0.6 mg/kg

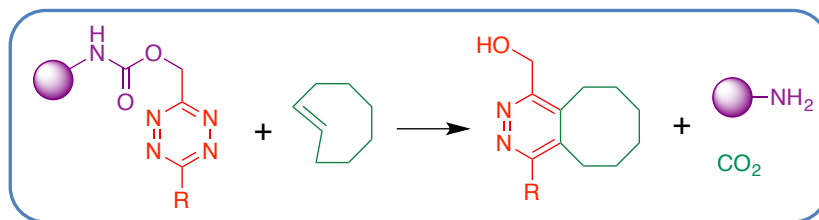
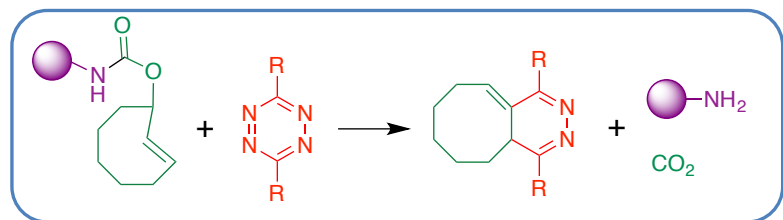
release: 90 % / dose: 10,000 eq, 36 mg/kg

release: 50 % / dose: 10,000 eq, 268 mg/kg

release: 90 % / dose: >100,000 eq

release: 10 % / dose: 50 eq

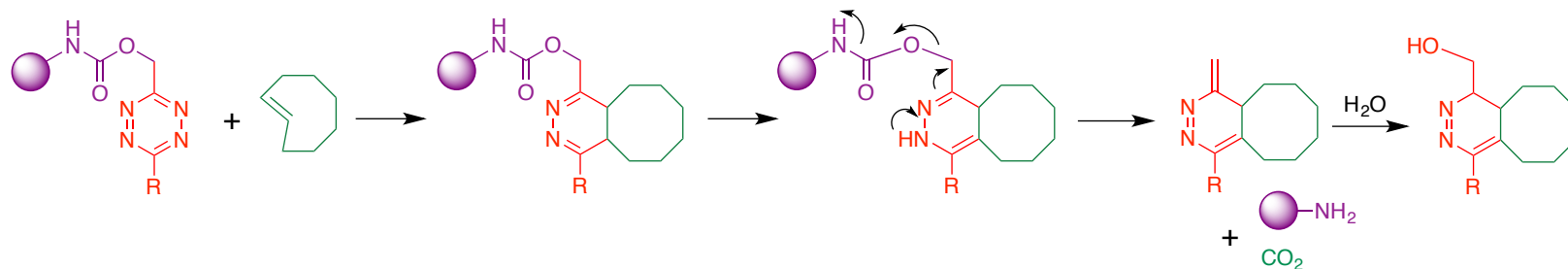
# New click-to-release reaction: swapping TCO and tetrazine



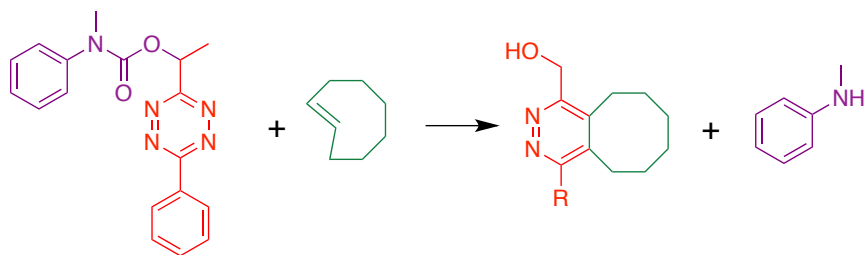
Allows use of highly reactive sTCO as activator to boost reactivity of system 100-fold



## Envisioned mechanism

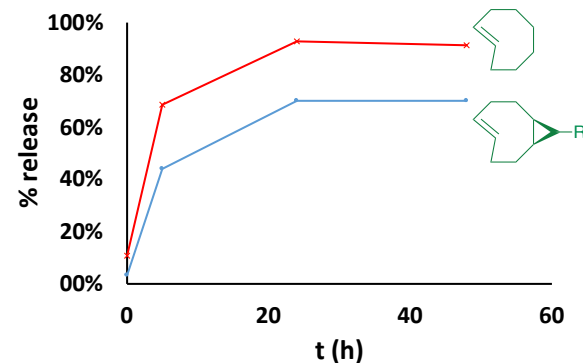


# New click-to-release reaction: swapping TCO and tetrazine

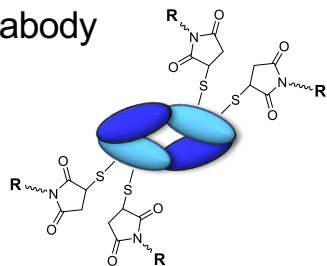


20% MeCN/PBS@37°C:

Stability  $t_{1/2}$ : 75 days  
Release  $t_{1/2}$ : 10 h

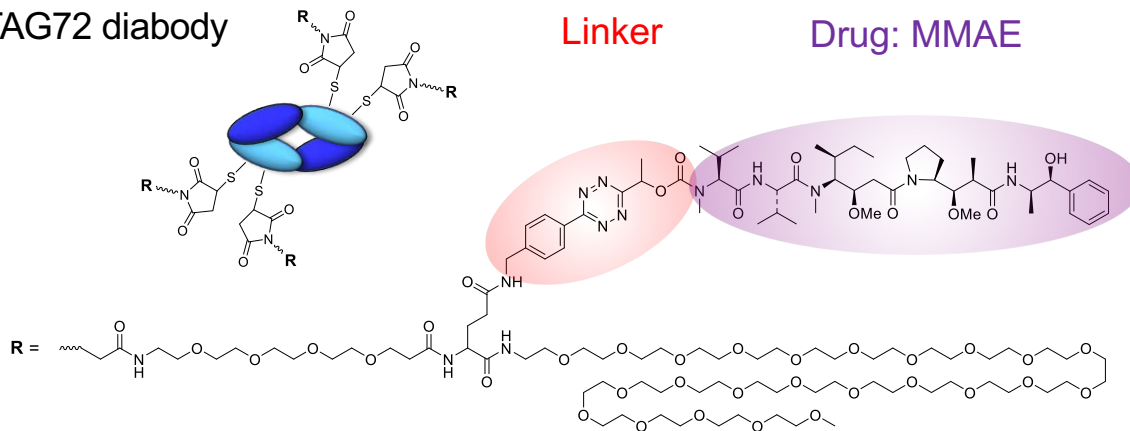


TAG72 diabody

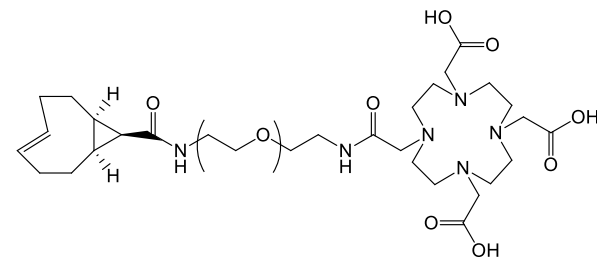


Linker

Drug: MMAE



Activator

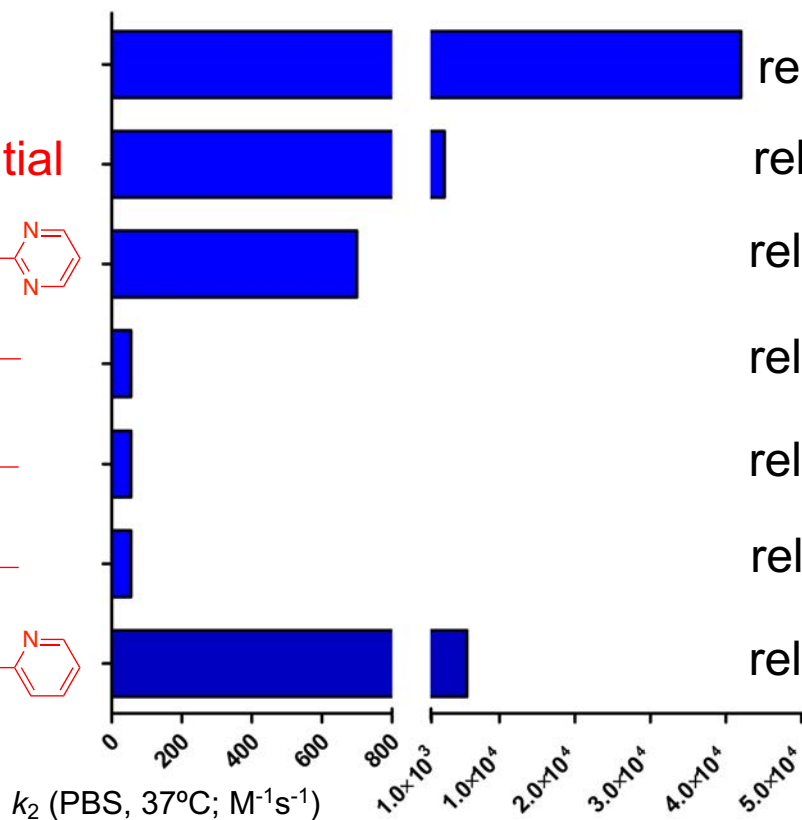
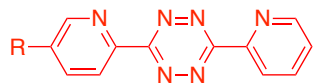
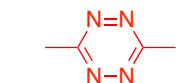
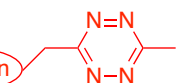
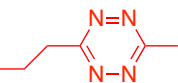
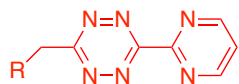


Reactivity ( $k_2$ ): 42000 M<sup>-1</sup>s<sup>-1</sup>  
Max release: 70 % (within 24 h)

# Click-to-Release: reactivity vs. release



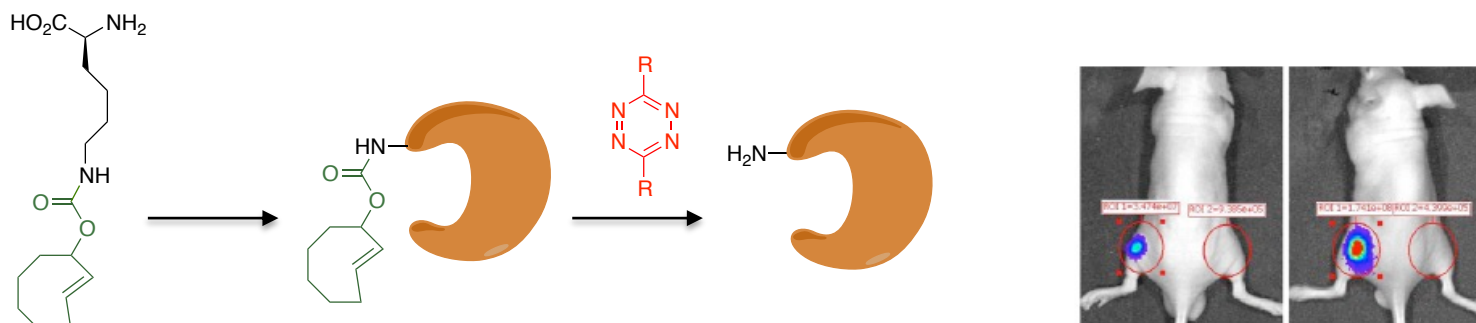
confidential



## Other applications of the IEDDA pyridazine elimination reaction

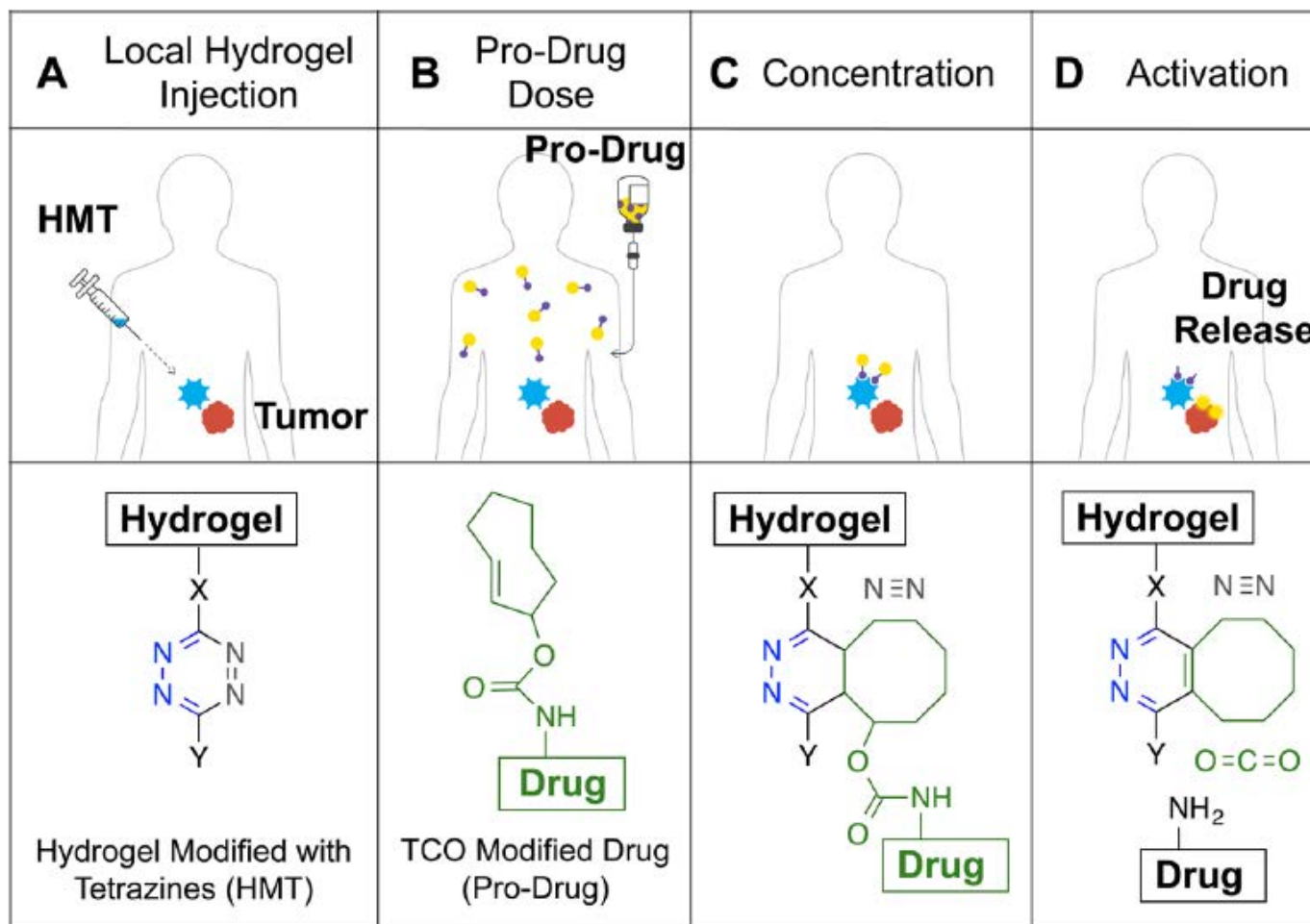


Genetic incorporation of masked lysine in luciferase and its in vivo intracellular unmasking with tetrazine



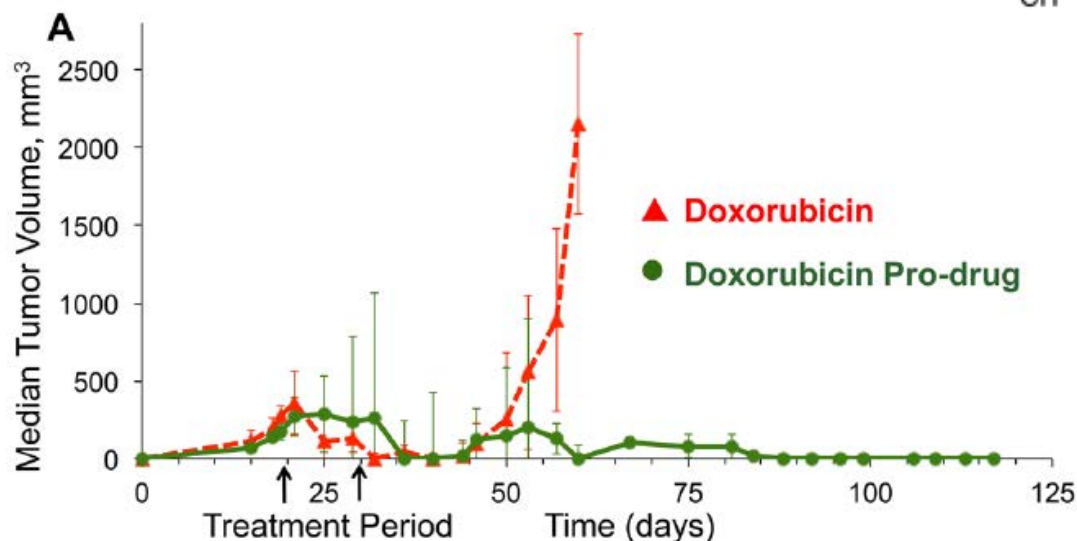
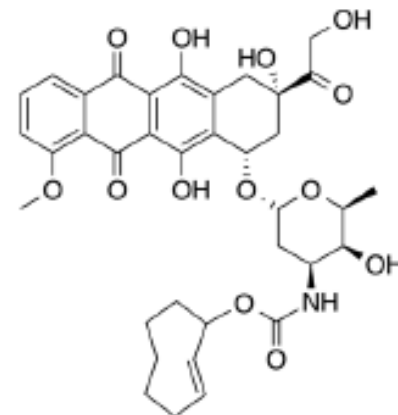
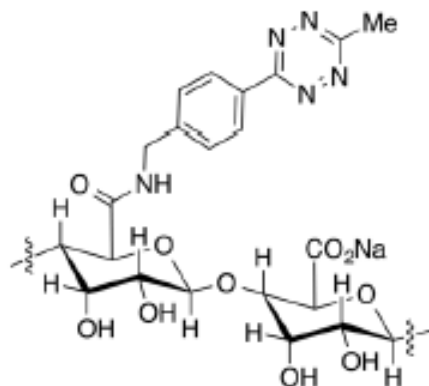
Incorporation of a single cleavable TCO at the right position can deactivate the protein. Protein activity can be restored on demand, also intracellularly, upon treatment with a tetrazine activator.

# Click-to-release approach to local drug activation



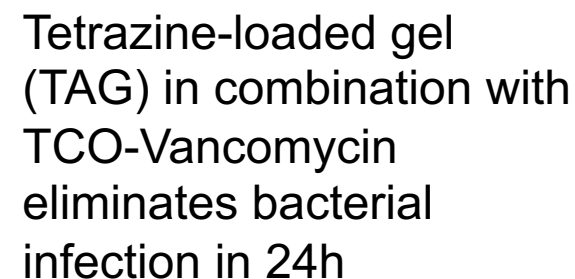
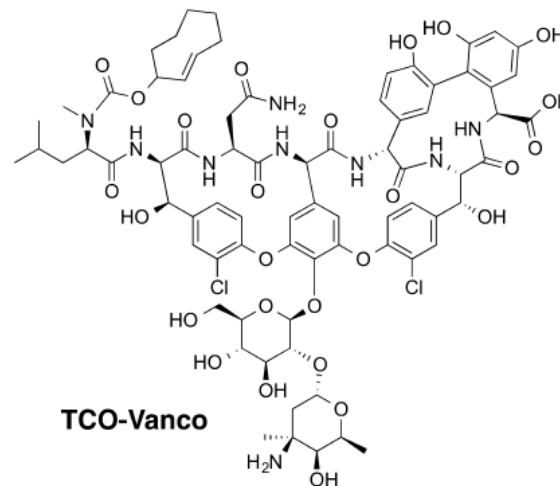
# Click-to-release approach to local drug activation

Nude mice bearing HT-1080 soft tissue sarcoma locally injected with a Tz-alginate gel followed by TCO-Dox prodrug i.v.



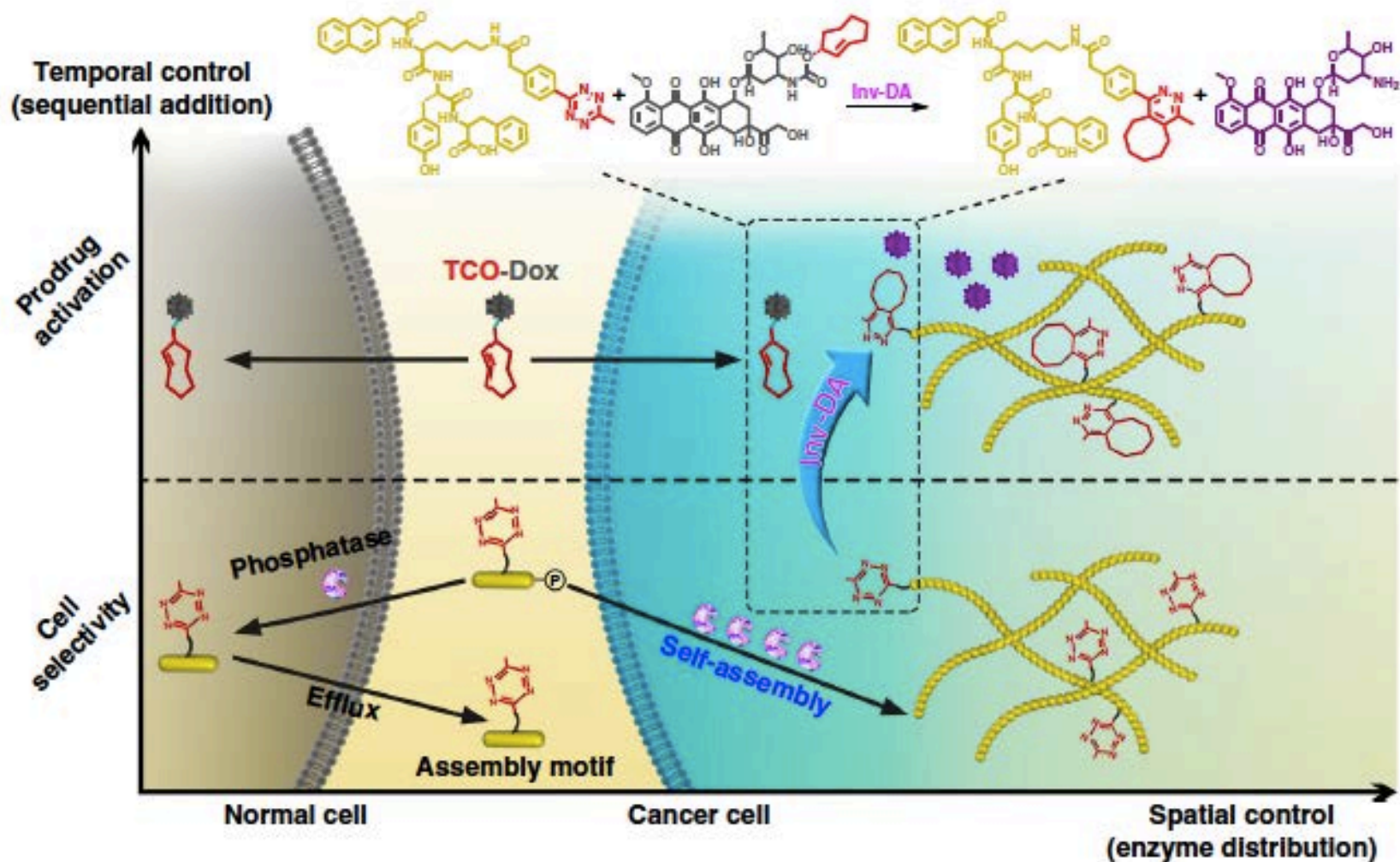
5/10 tumors treated  
(the remaining mice were  
euthanized with > 2 gr  
tumors)

Promising approach for  
**treatment of tumor  
resection areas** (to eliminate  
residual tumor cells)



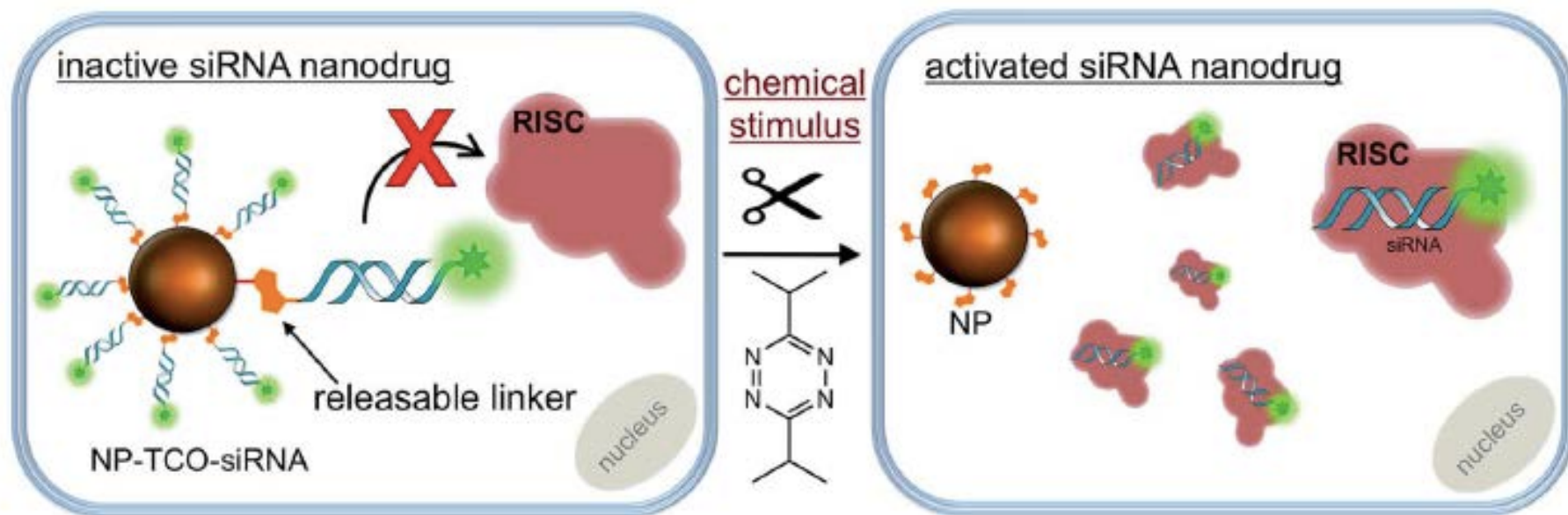
# Enzymatic Tz accumulation followed by pro-drug uncaging

Tumor bearing mice injected with 3 cycles of 50 mg/kg Tz + 30mg/kg TCO-Dox





## Cleavage of siRNA from nanoparticle carrier

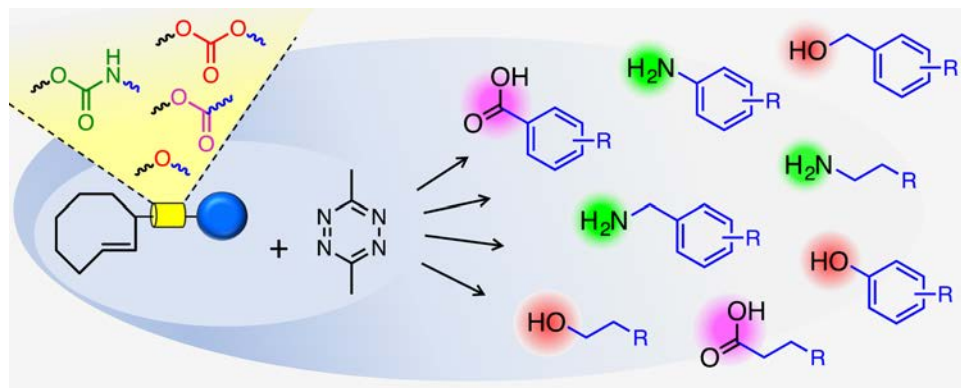


Khan *et al.*, *Chemical Science* **2017**, 8, 5705

and many more...

# Summary – Click-to-release

- Click-cleavable ADCs expand ADC scope to non-internalizing targets & platform technology for controlled payload delivery to TME
- Diabody ADC: high tumor uptake, fast clearance, low systemic exposure
- High intratumor MMAE levels, minimal washout, no toxicity observed
- Potential for homogenous drug distribution and activation of tumor-residing immune cells
- Platform technology with a variety of application, also in synthetic chemistry
- Broad chemical scope:

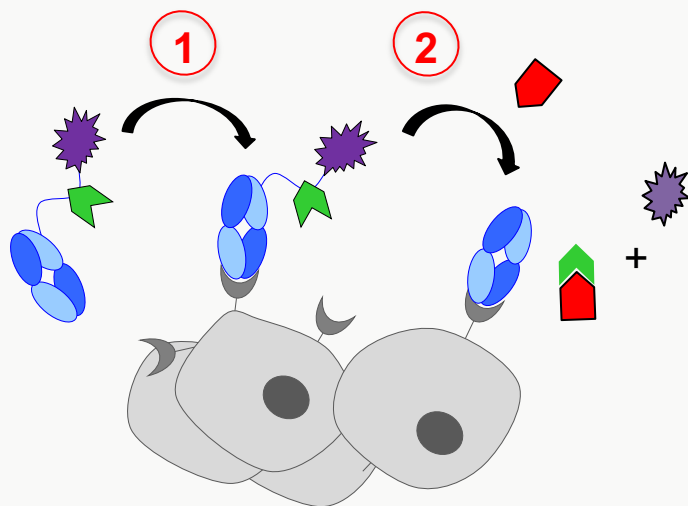


# Tagworks' Click-to-Release

A platform technology that can be used with a broad array of immune modulators

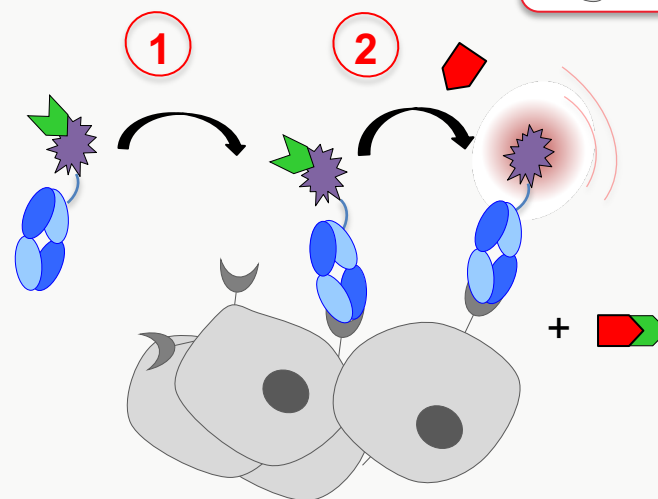
## Applications:

### Drug Release

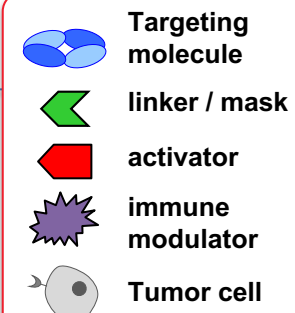


Examples: TLR agonists, STING agonists, cytokines, etc

### Drug unmasking

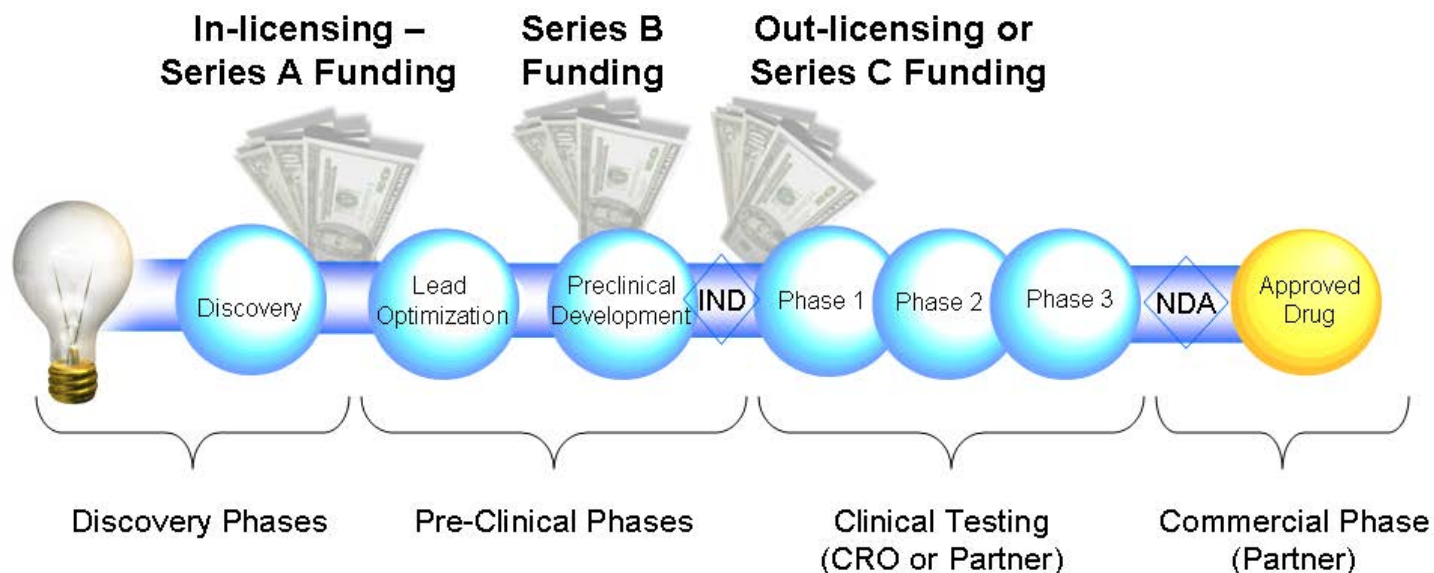


Examples: masked T-cell engagers, immunocytokines, etc



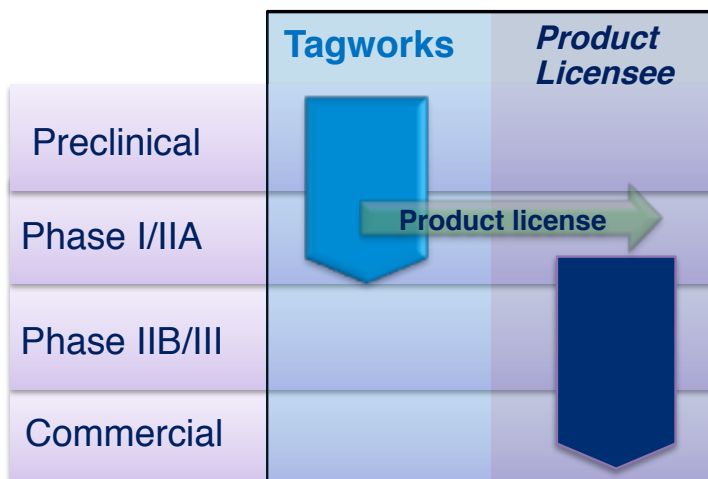


# Drug discovery pipeline



## Our business plan

### Product Development and Licensing



### Technology Licenses and Services

